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(54) **COMPOUNDS AND METHODS FOR MODULATING COMMUNICATION AND VIRULENCE IN QUORUM SENSING BACTERIA**

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(51) **Int. Cl.**

C07D 309/30 (2006.01)
A01N 43/08 (2006.01)

(52) **U.S. Cl.** **549/291; 514/471**

(58) **Field of Classification Search** **549/291; 514/471**

See application file for complete search history.

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(57) **ABSTRACT**

The present invention provides compositions and methods for modulating the communication and virulence of quorum sensing bacteria. In various exemplary embodiments, the invention provides a combinatorial library of quorum sensing compounds including synthetic analogs of naturally occurring and non-naturally occurring acyl-homoserine lactone (AHL) analogs, and methods of synthesizing and using these compounds.

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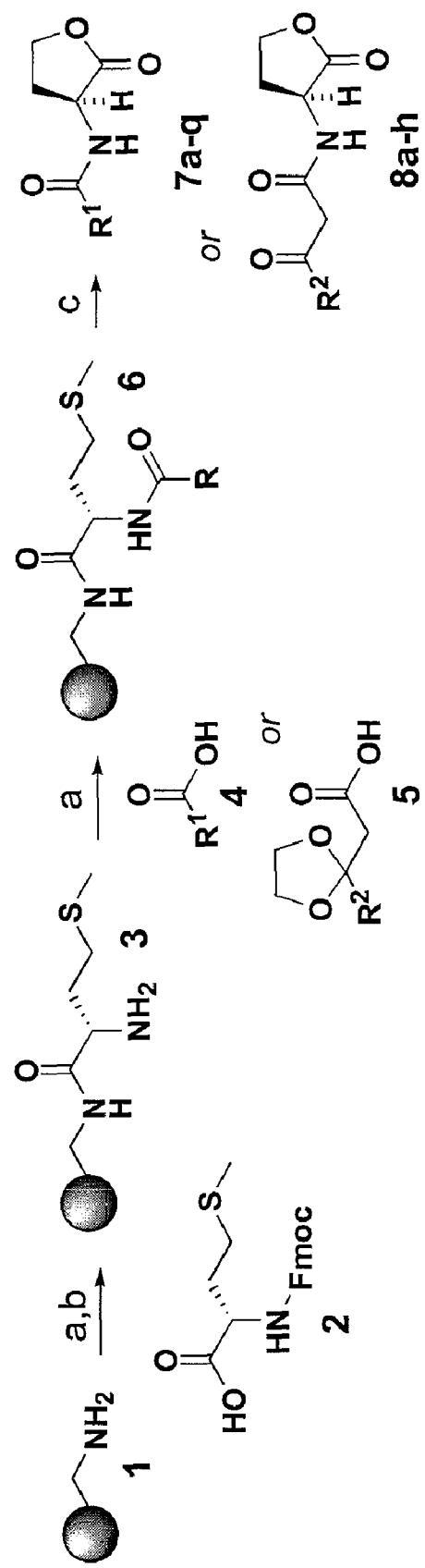


FIG. 1A

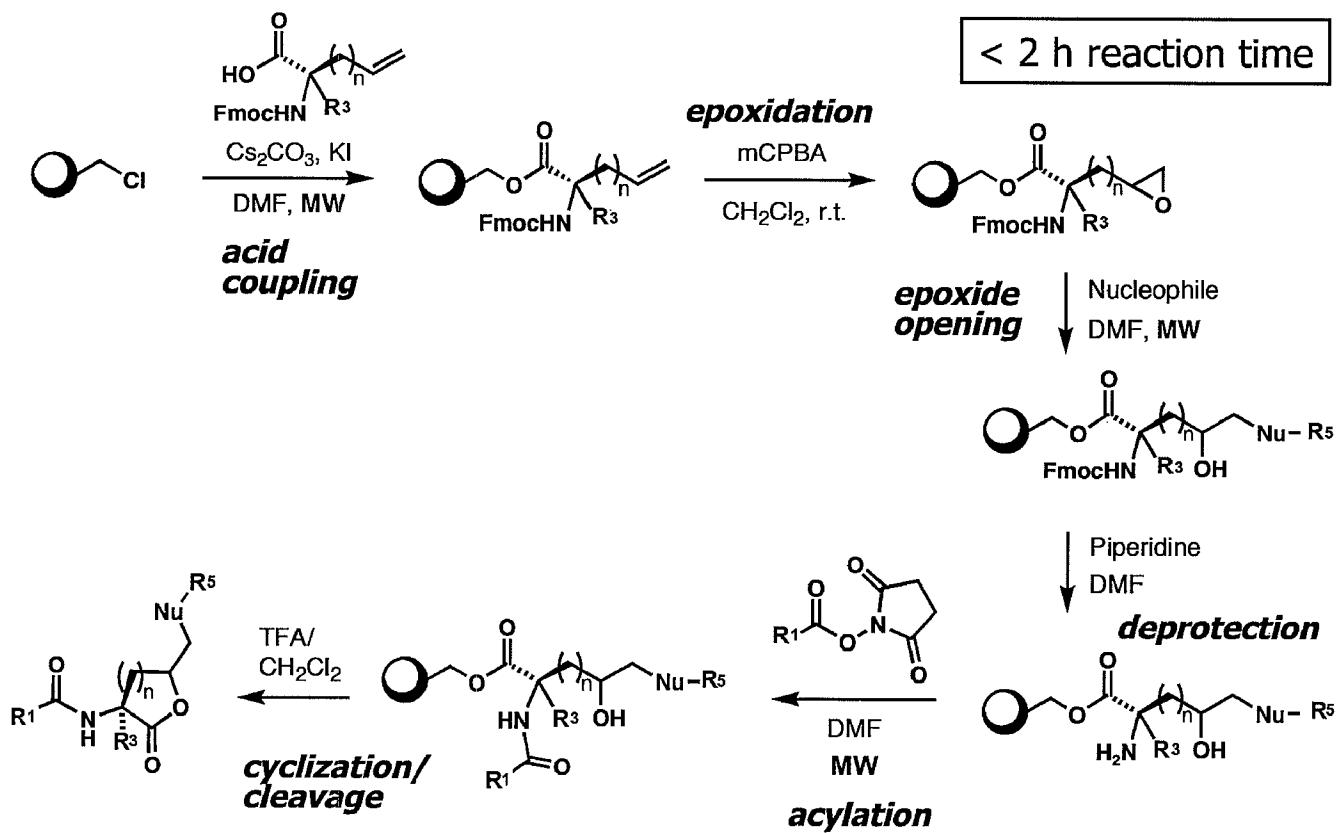


FIG. 1B

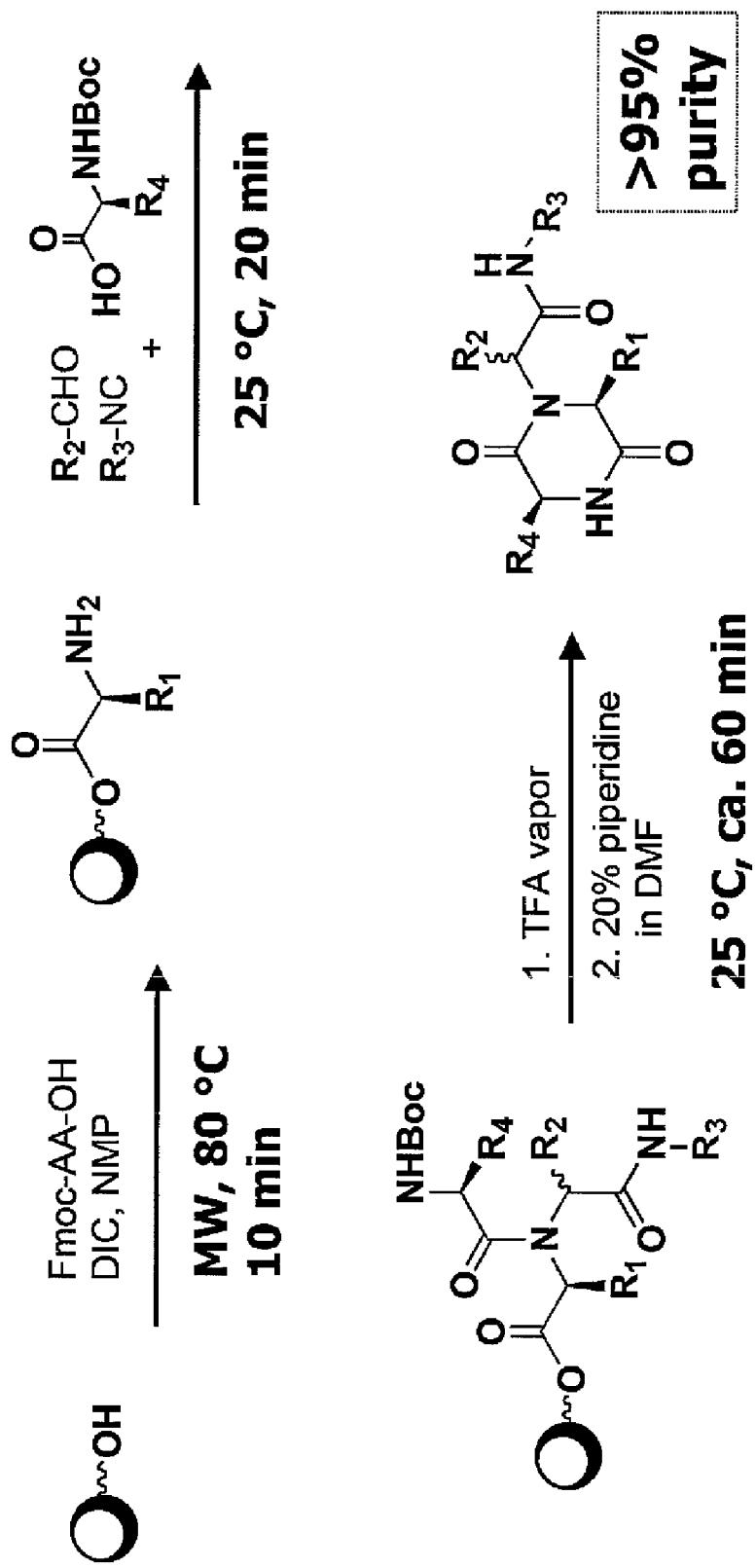


FIG. 1C

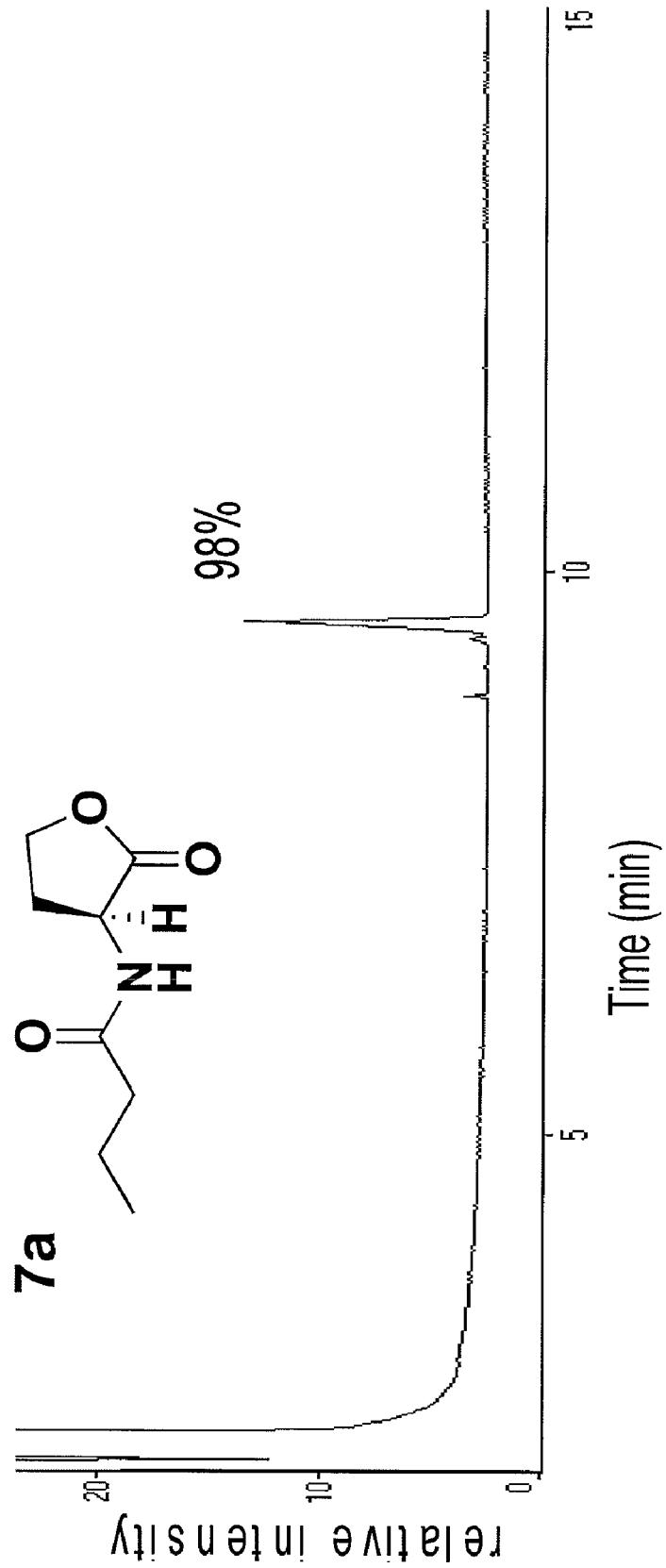


FIG. 2A

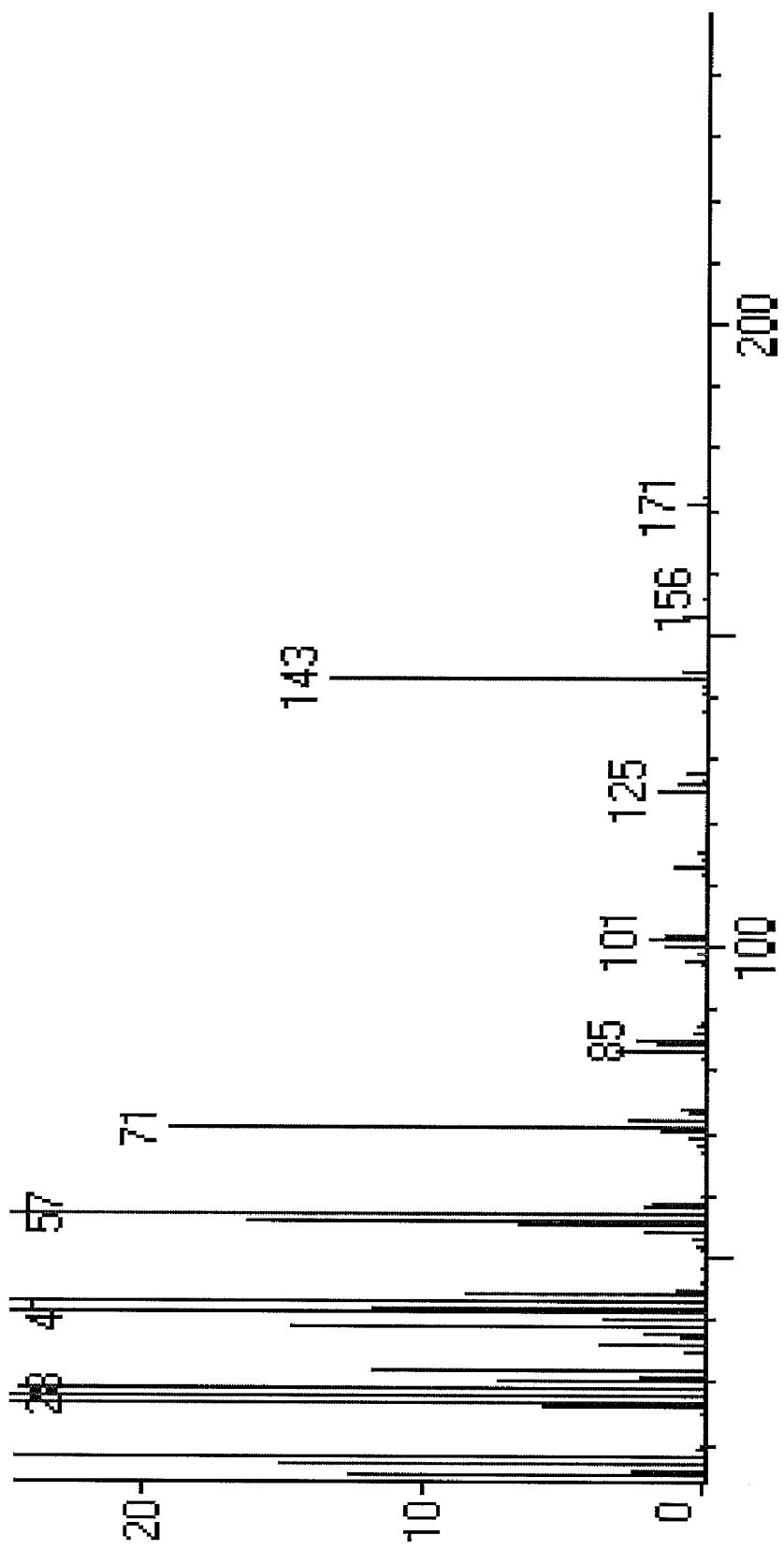


FIG. 2B

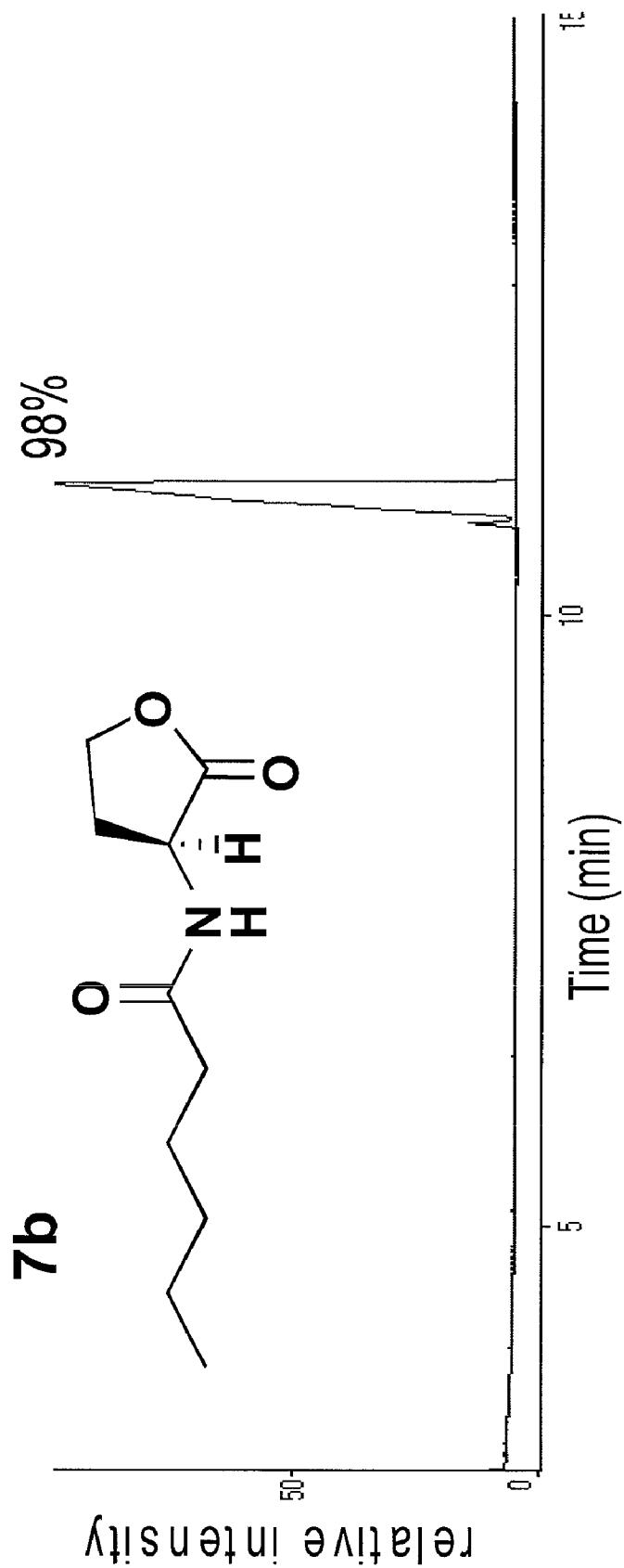


FIG. 3A

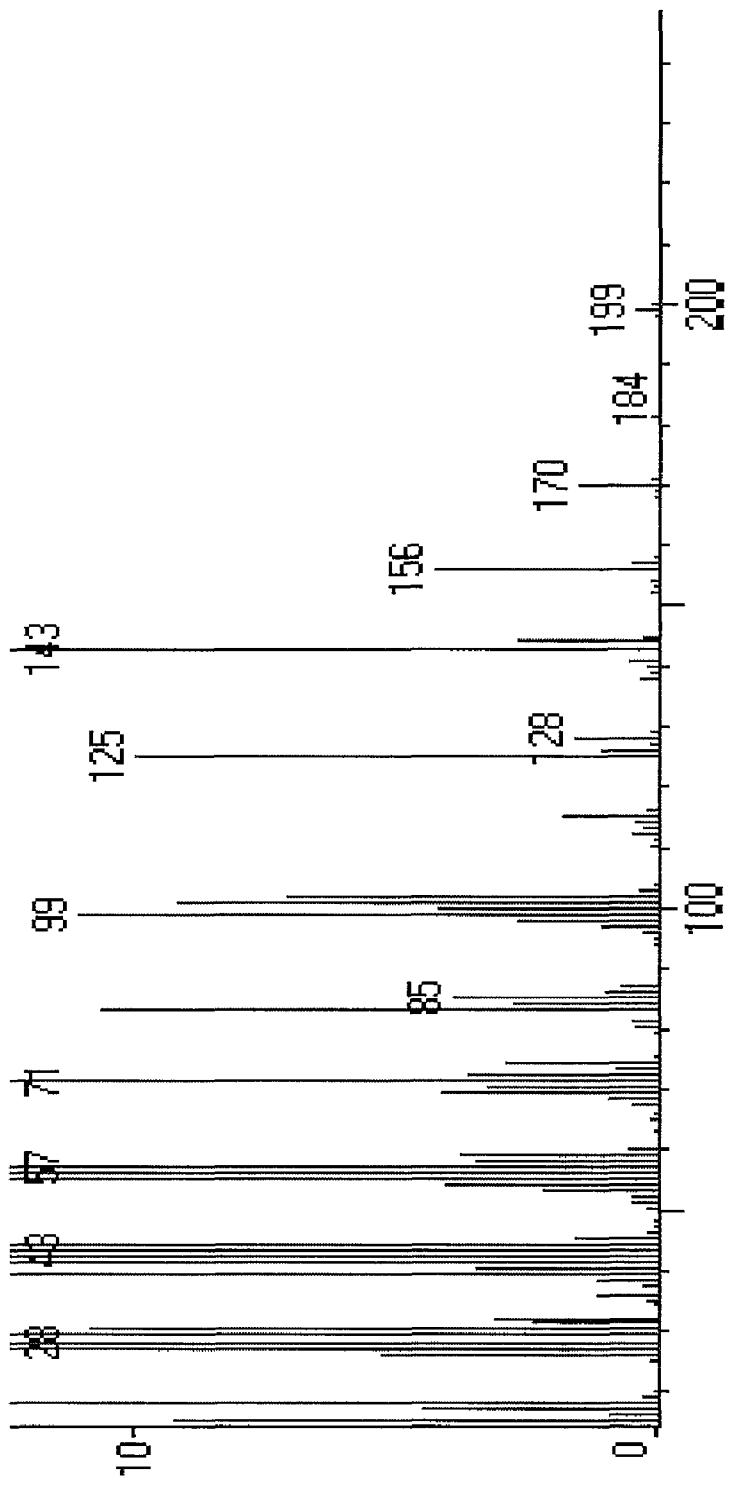


FIG. 3B

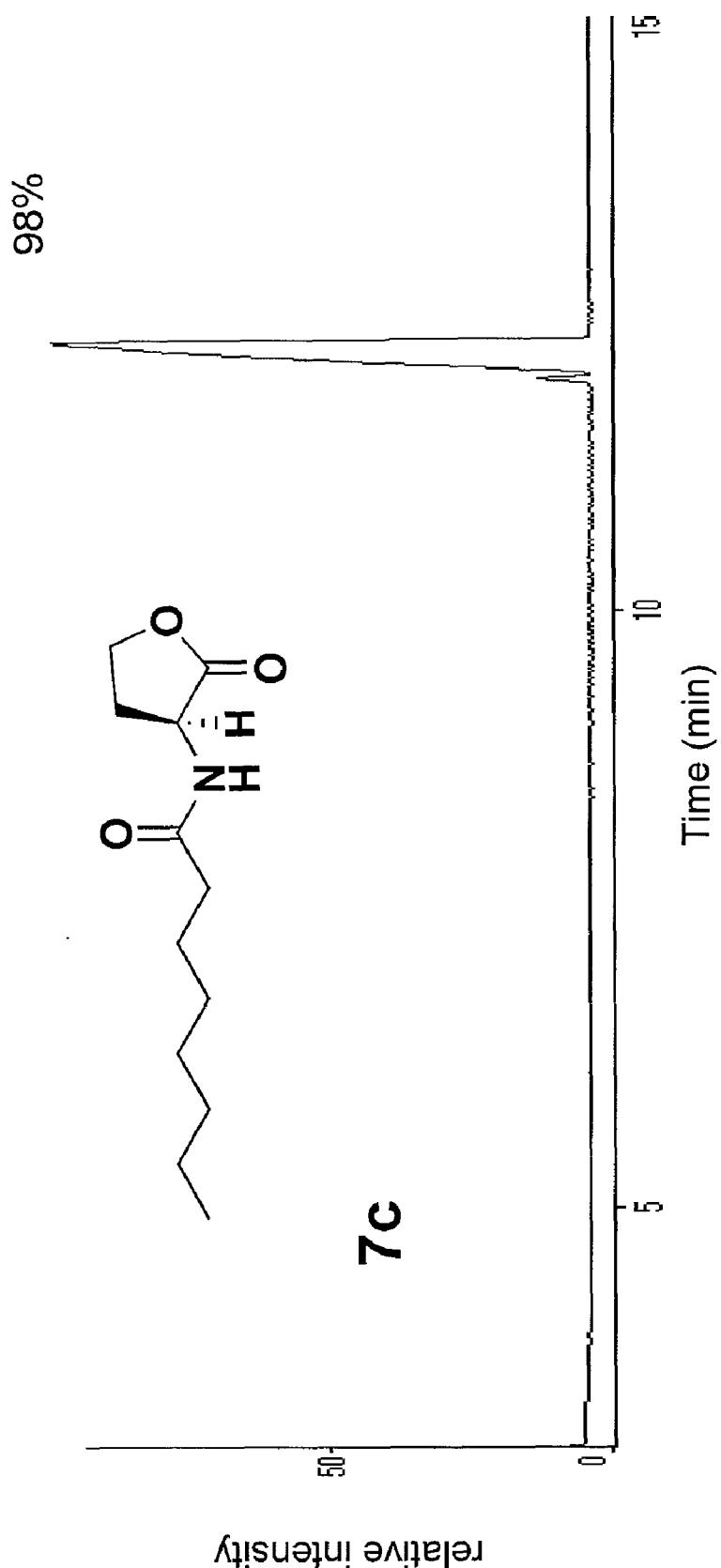


FIG. 4A

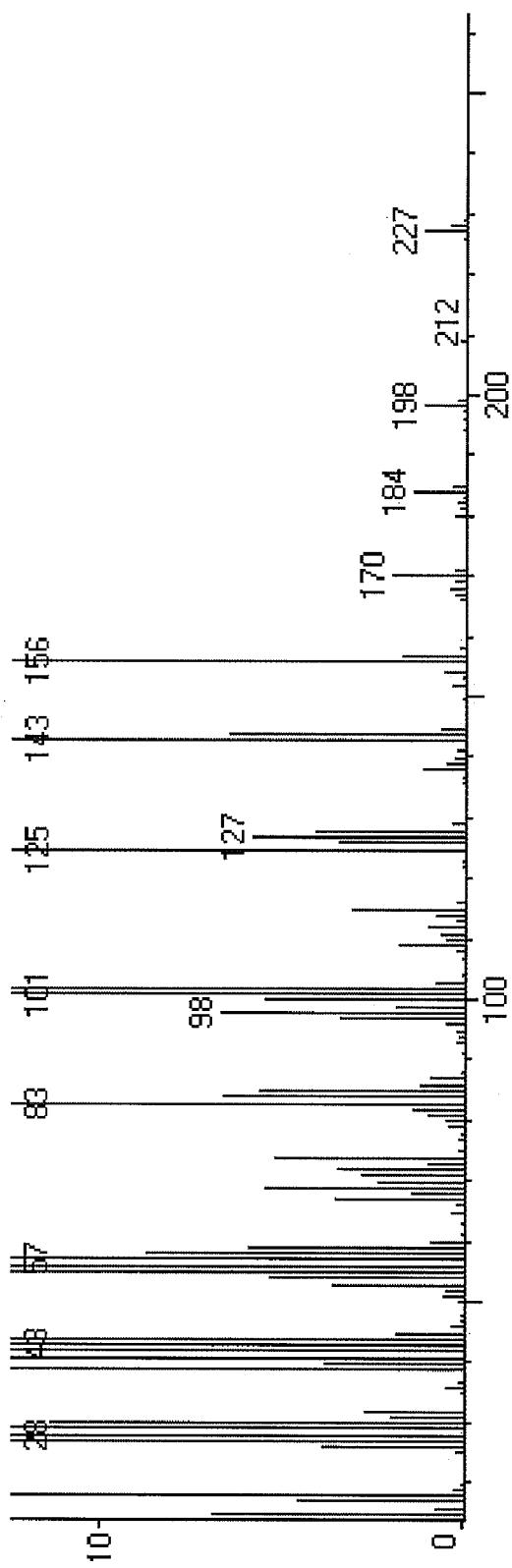


FIG. 4B

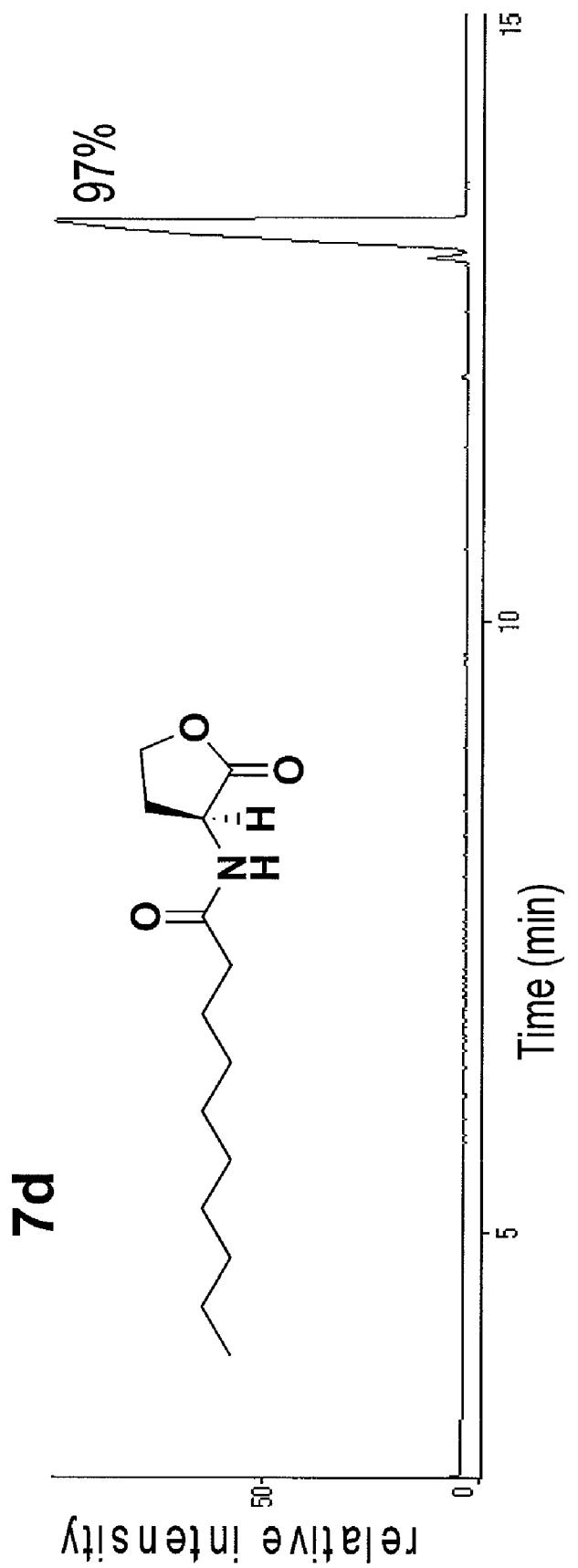


FIG. 5A

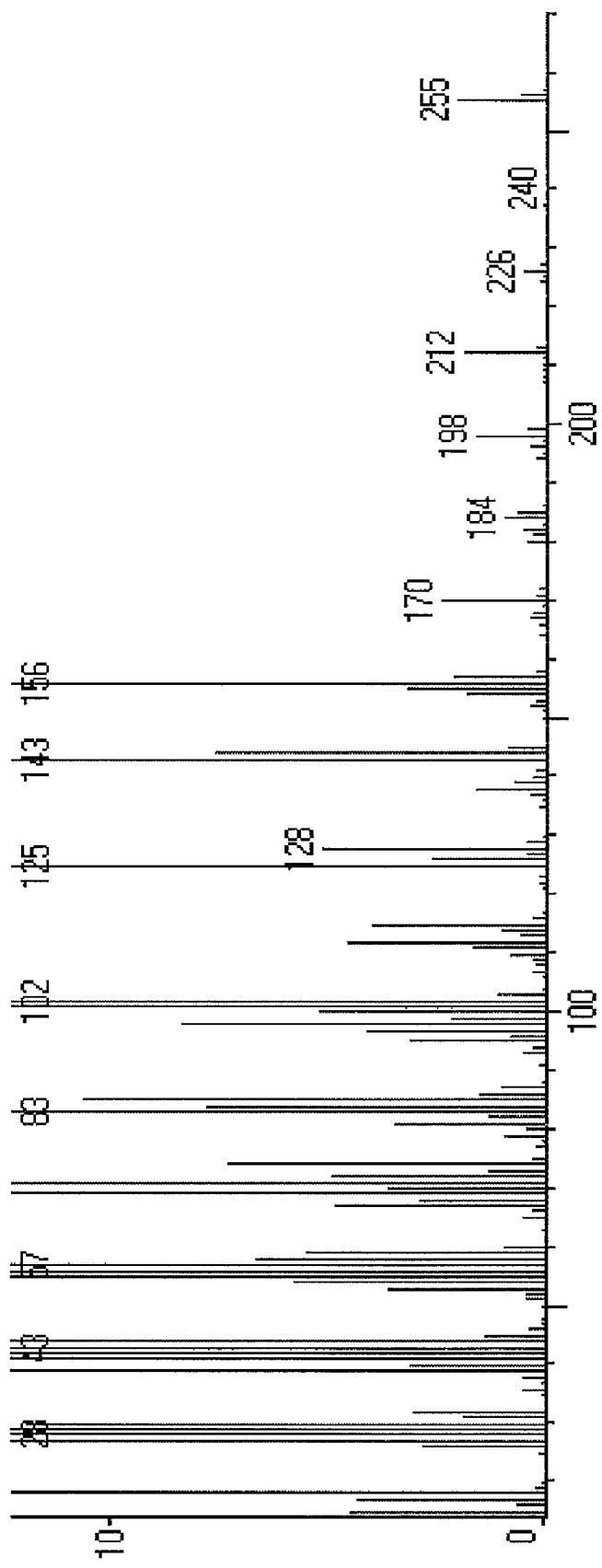


FIG. 5B

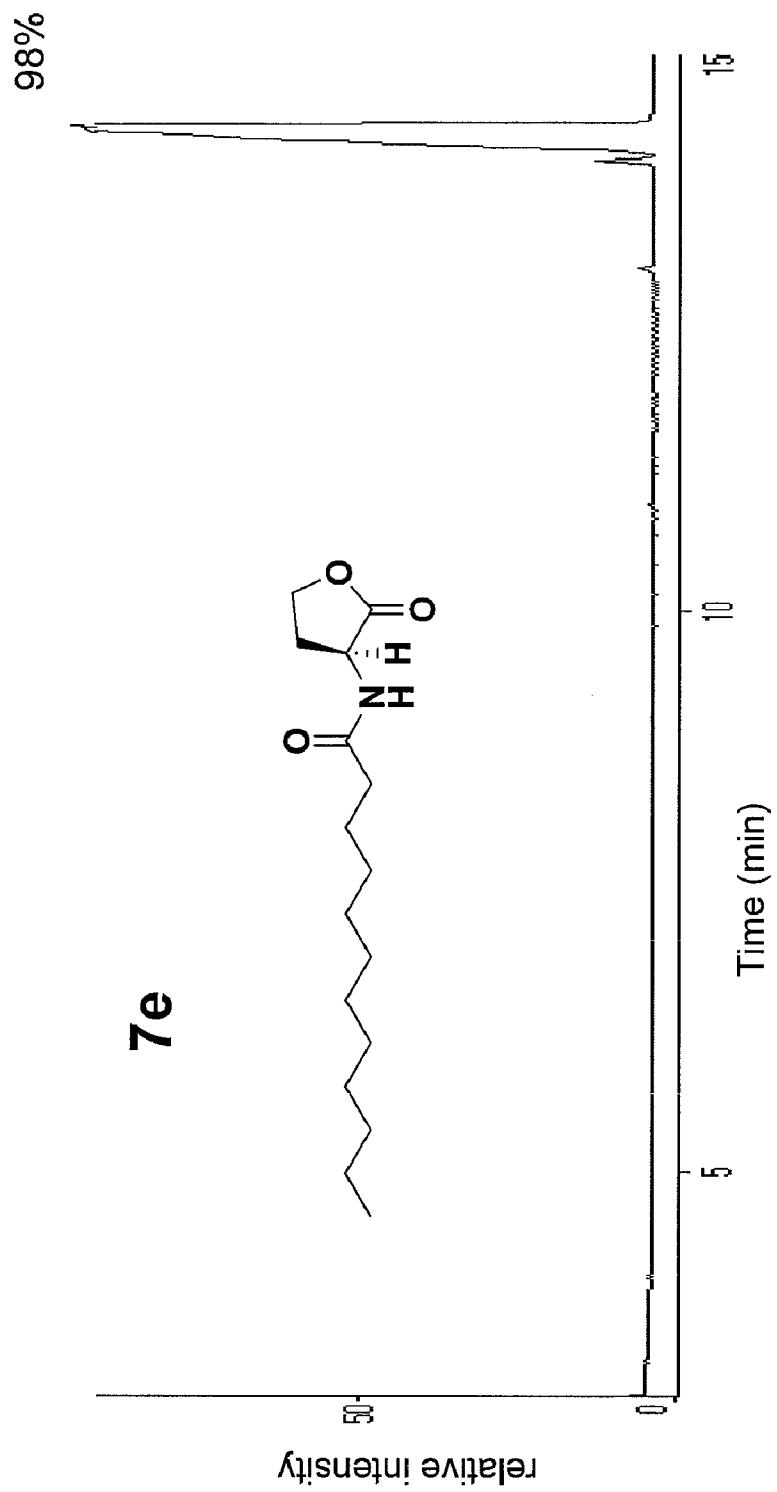


FIG. 6A

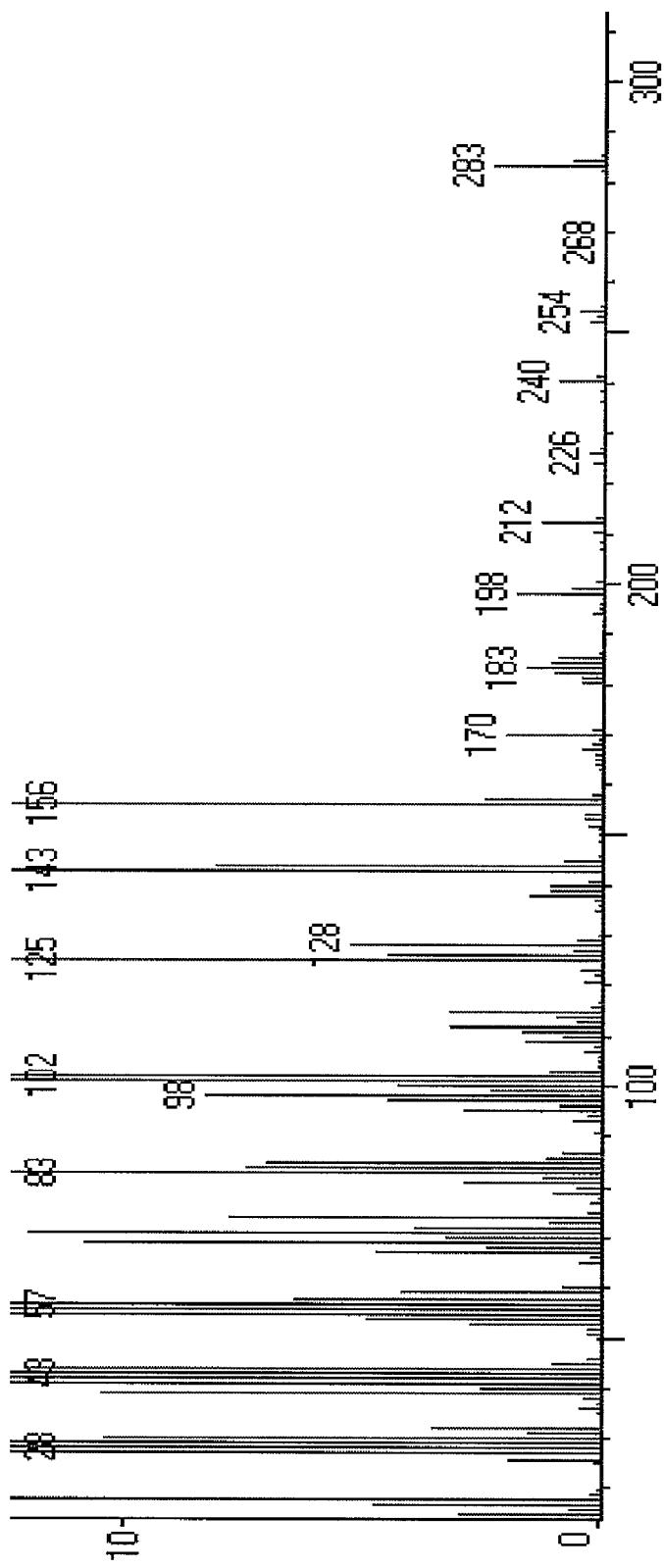


FIG. 6B

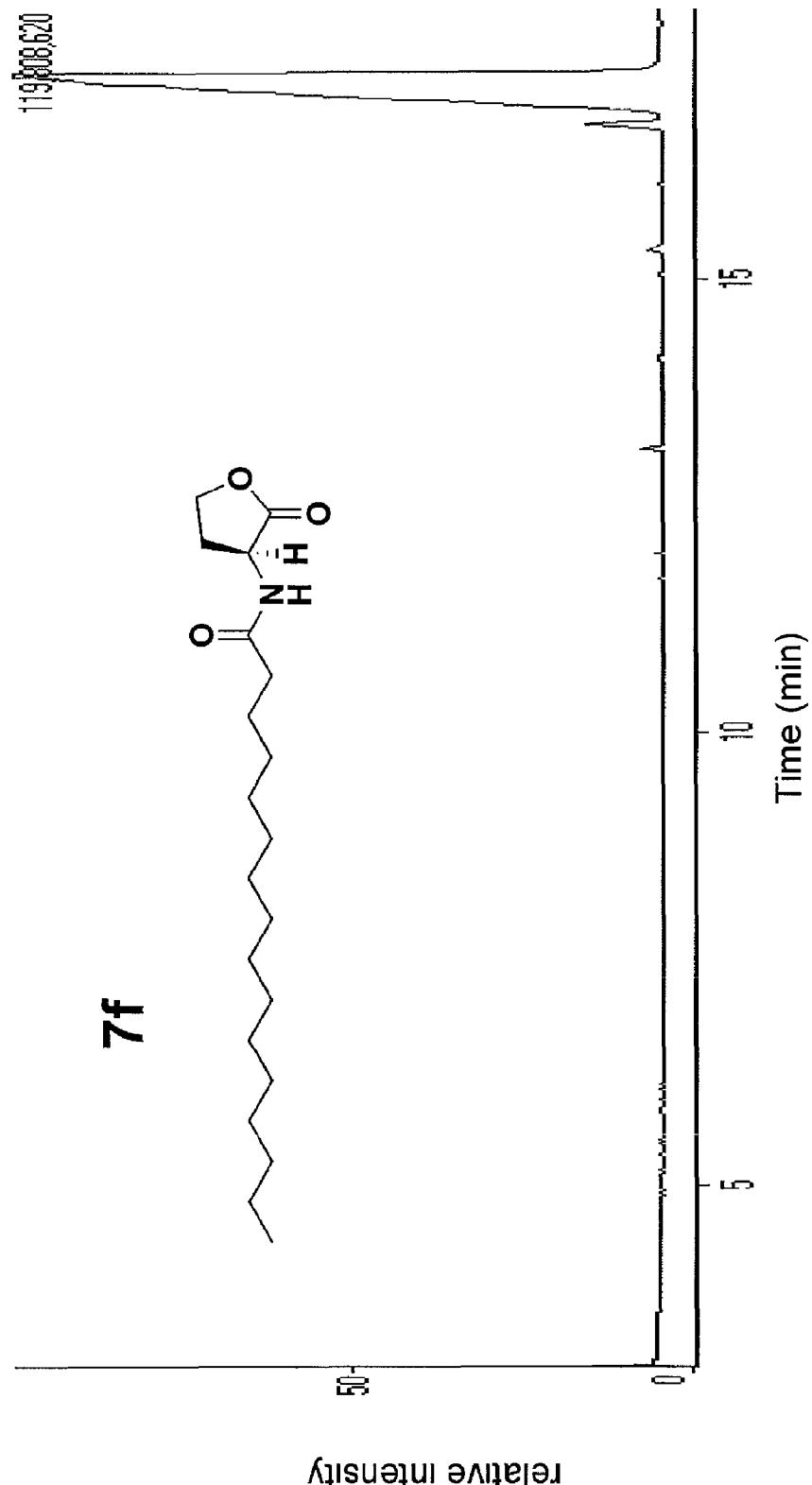


FIG. 7A

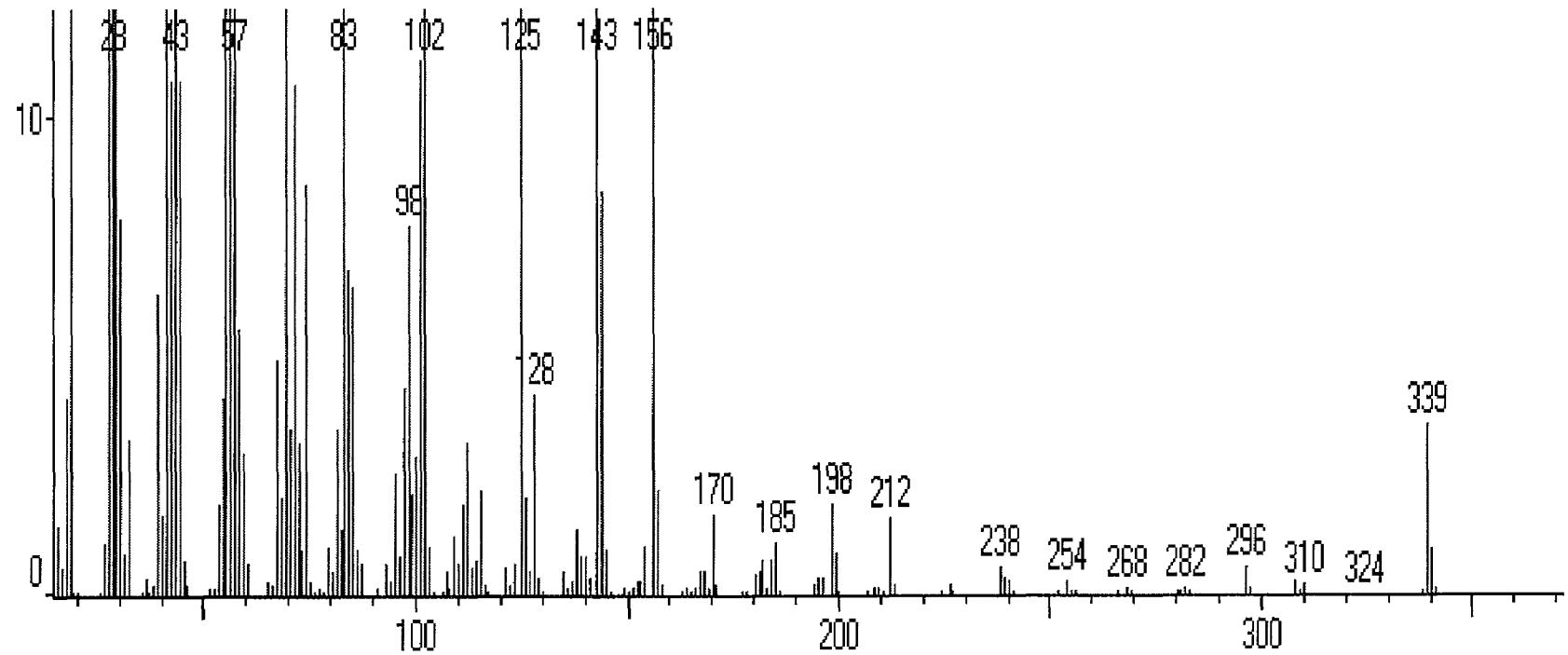


FIG. 7B

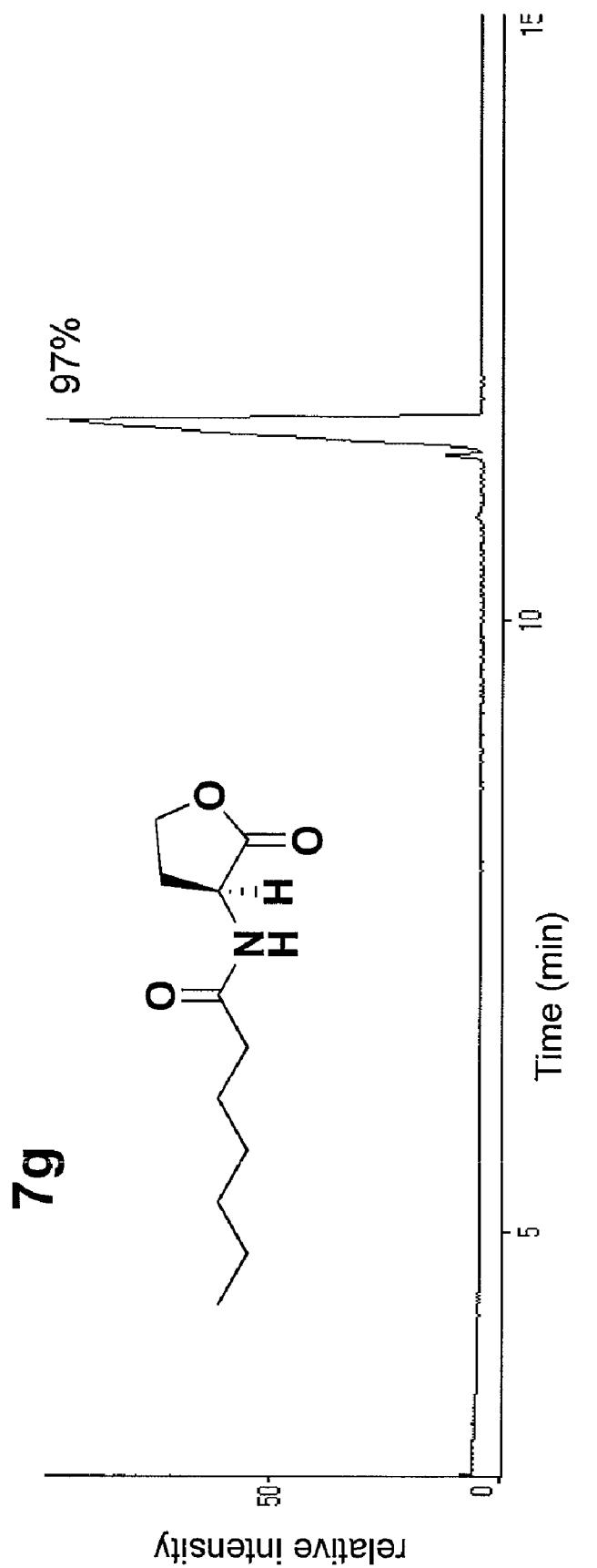


FIG. 8A

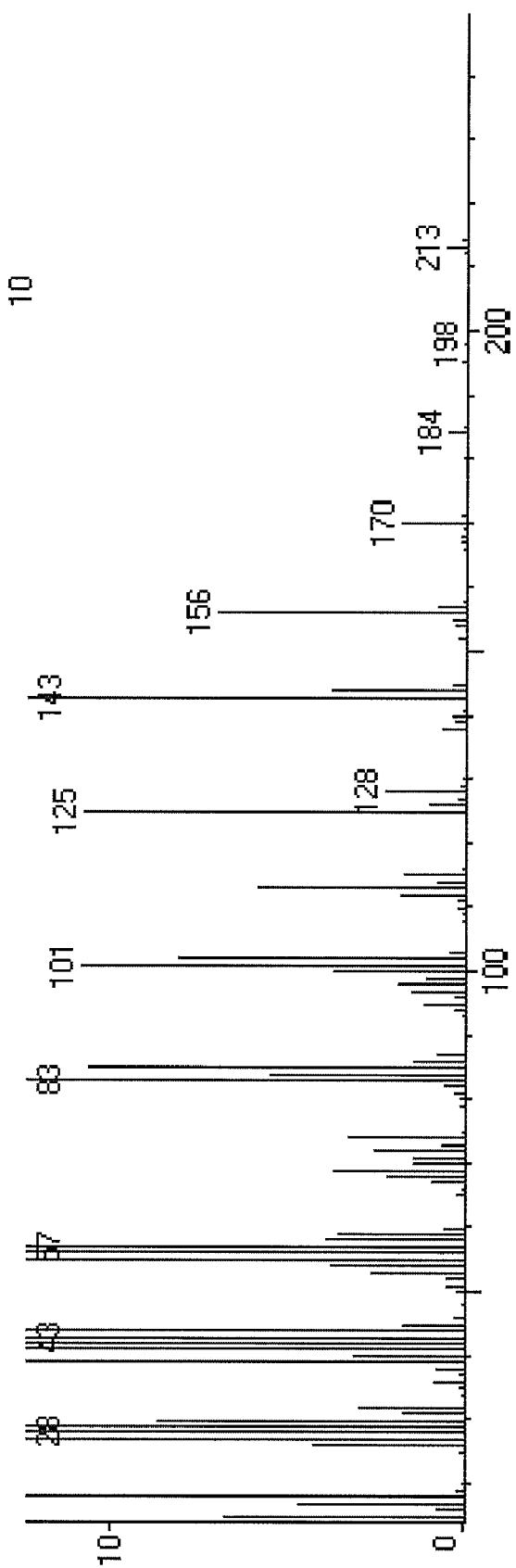


FIG. 8B

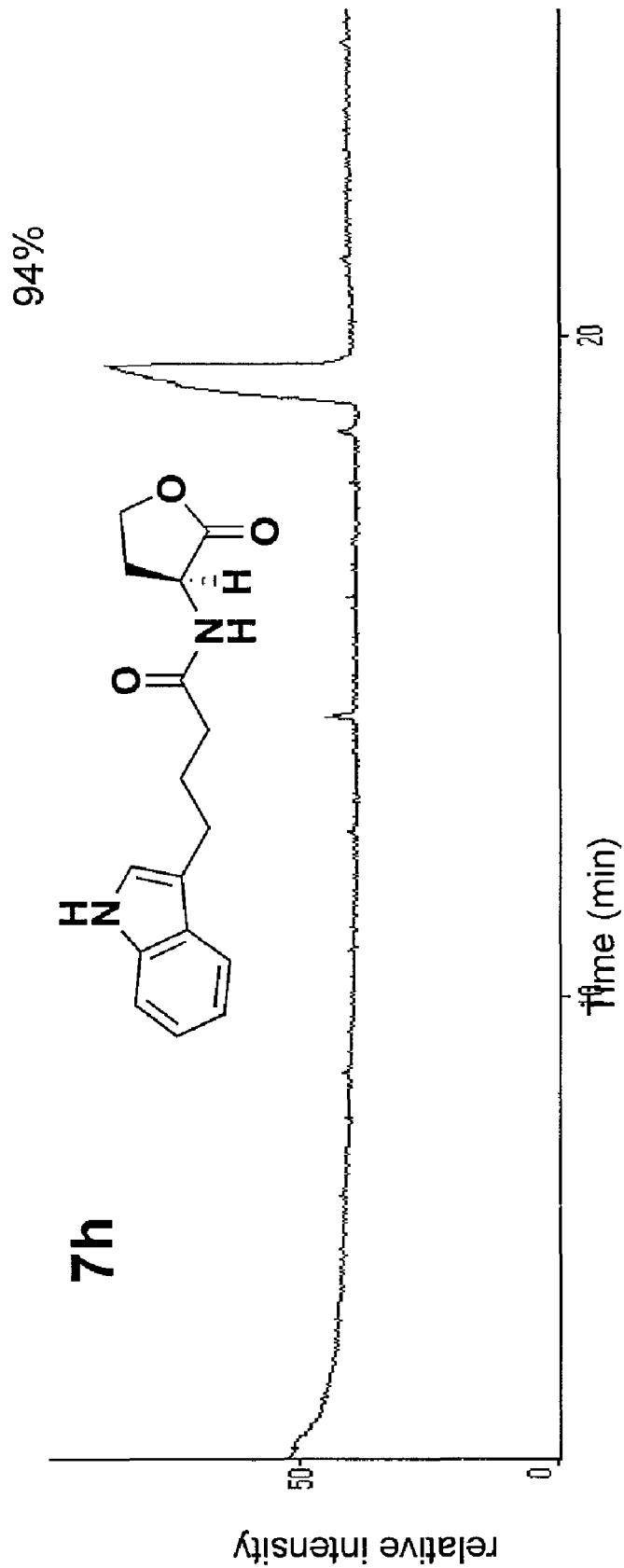


FIG. 9A

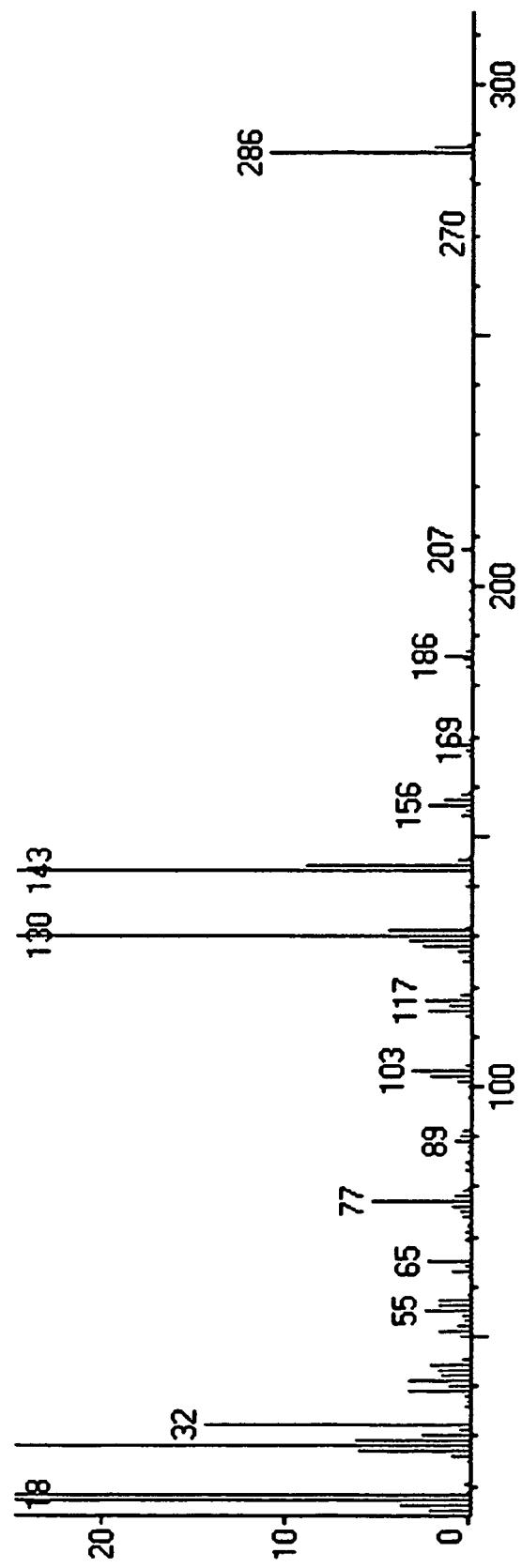


FIG. 9B

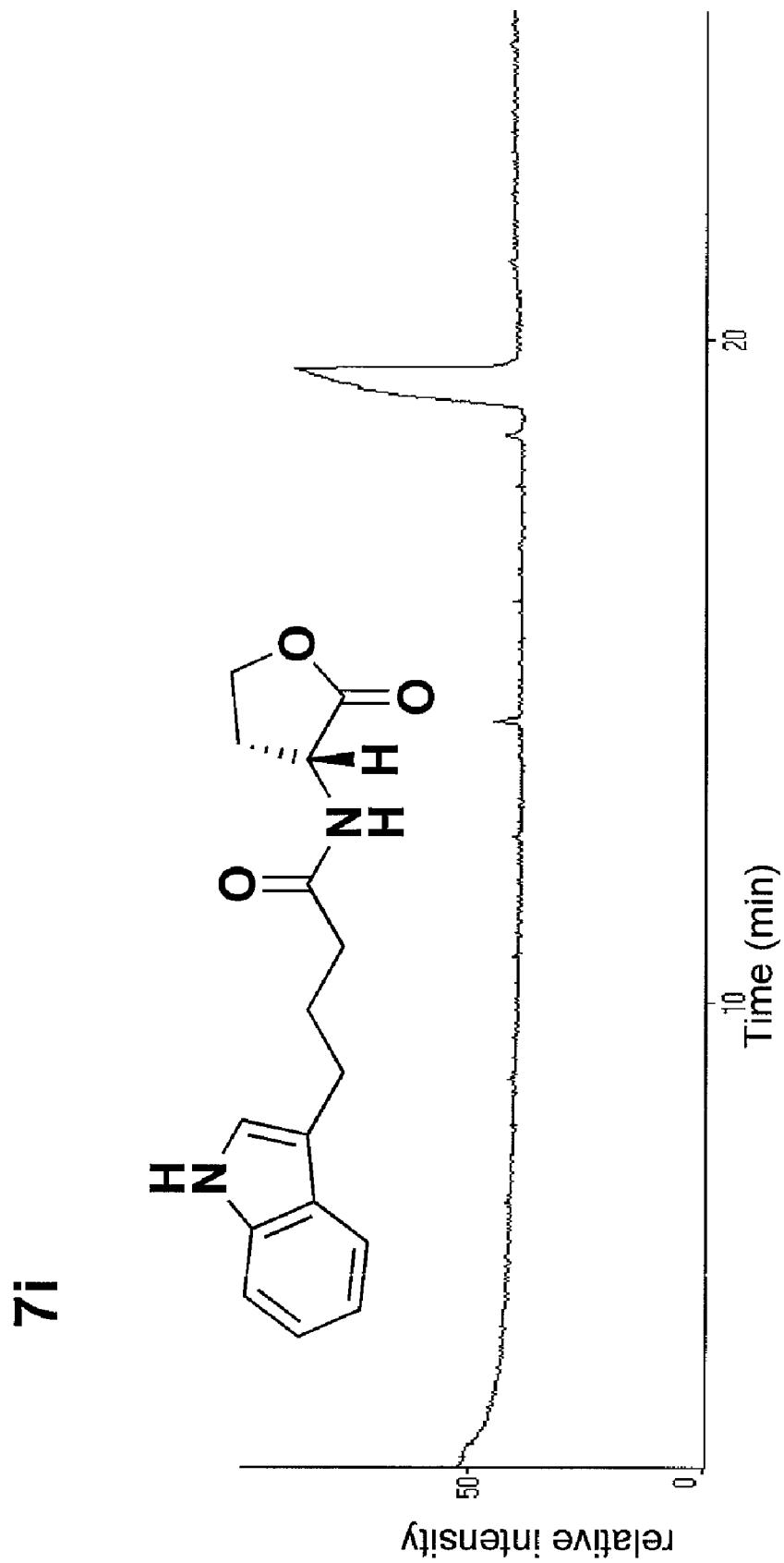


FIG. 10A

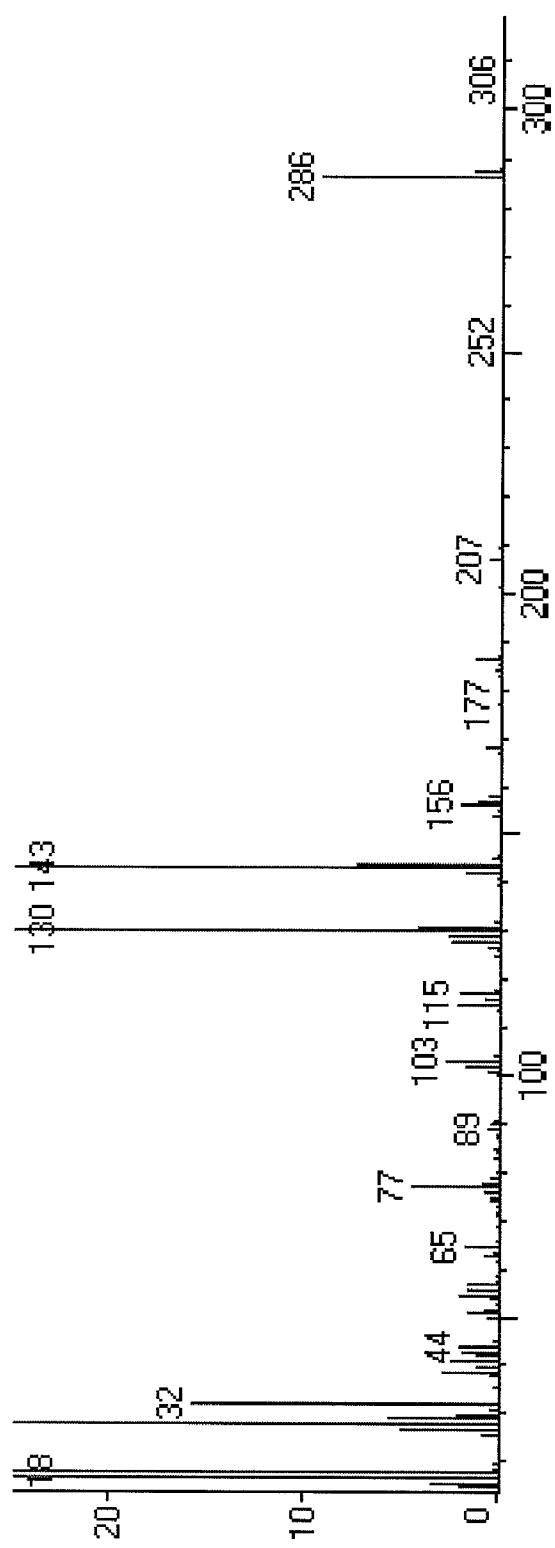


FIG. 10B

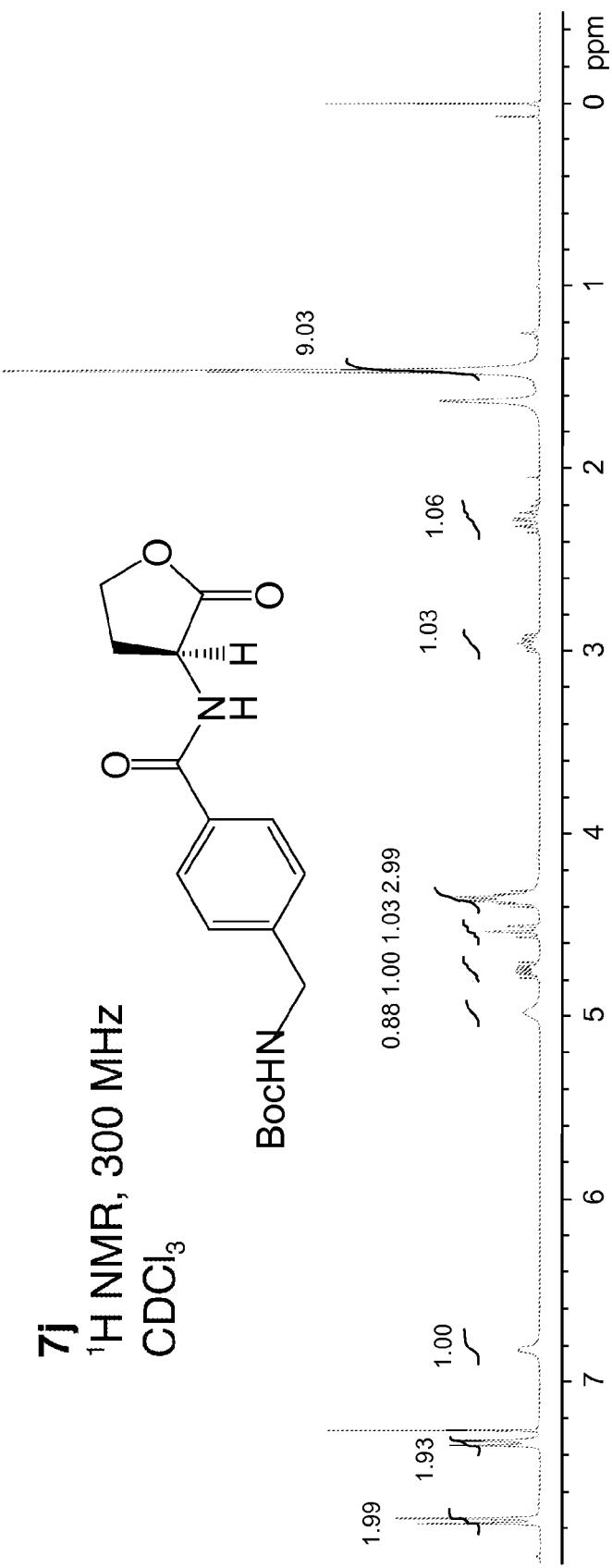


FIG. 11

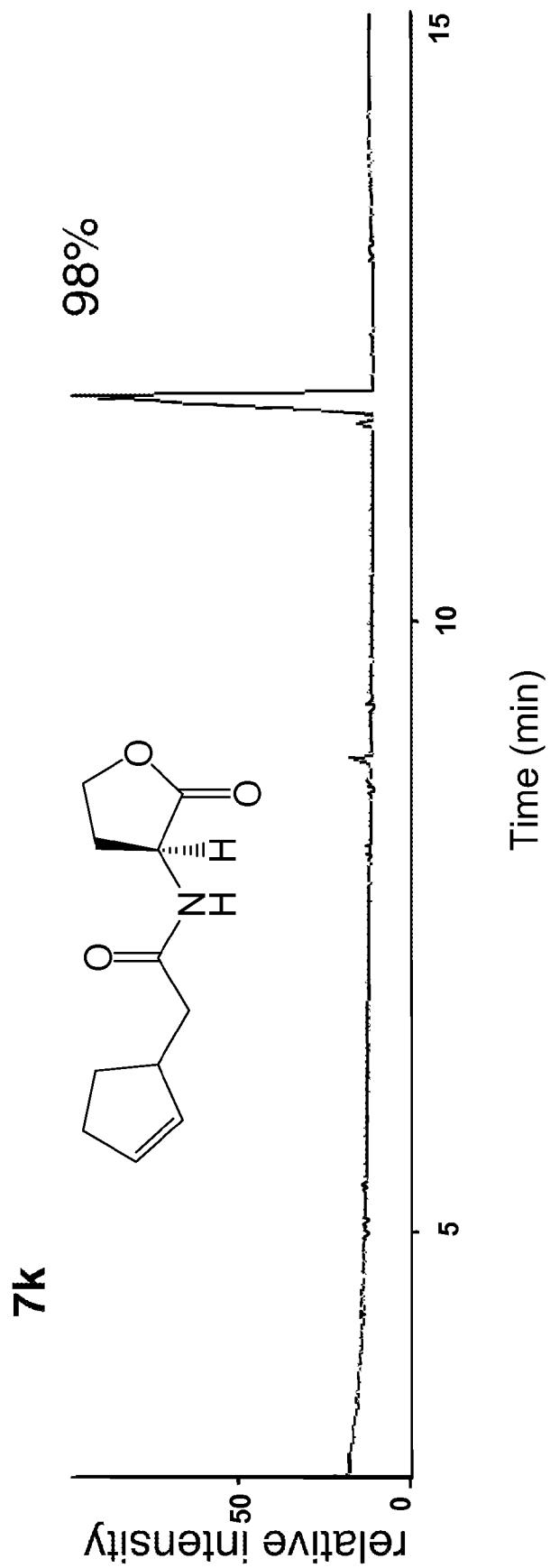


FIG. 12A

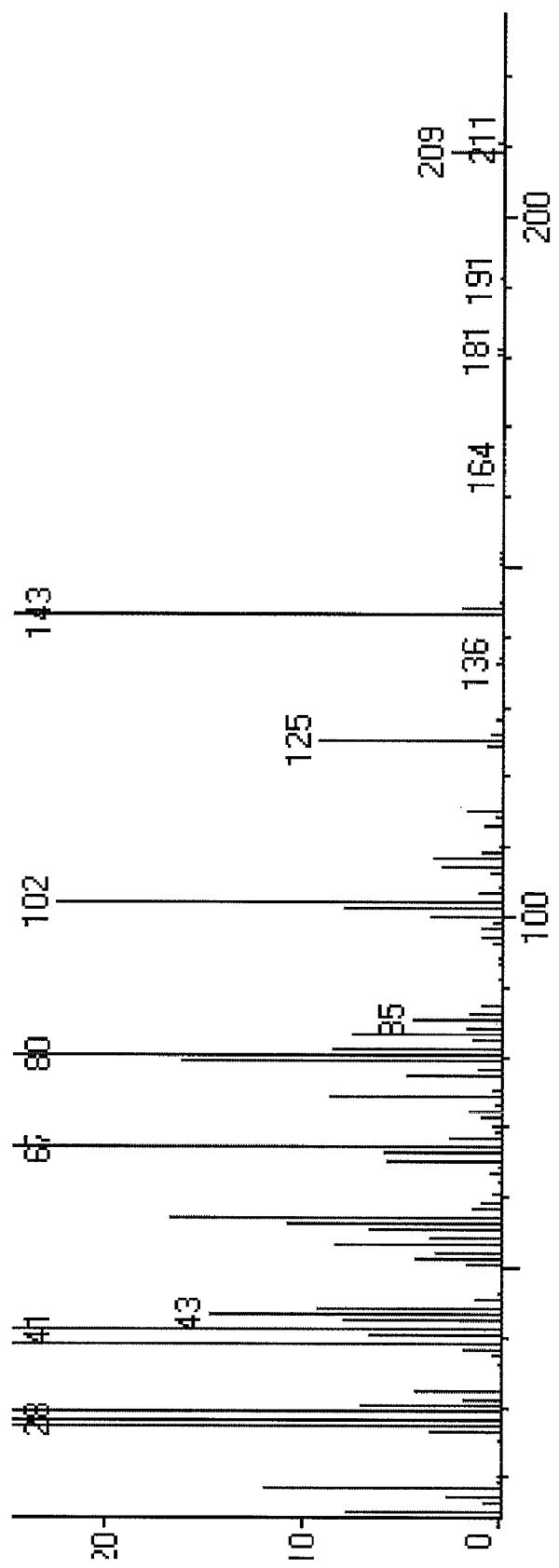


FIG. 12B

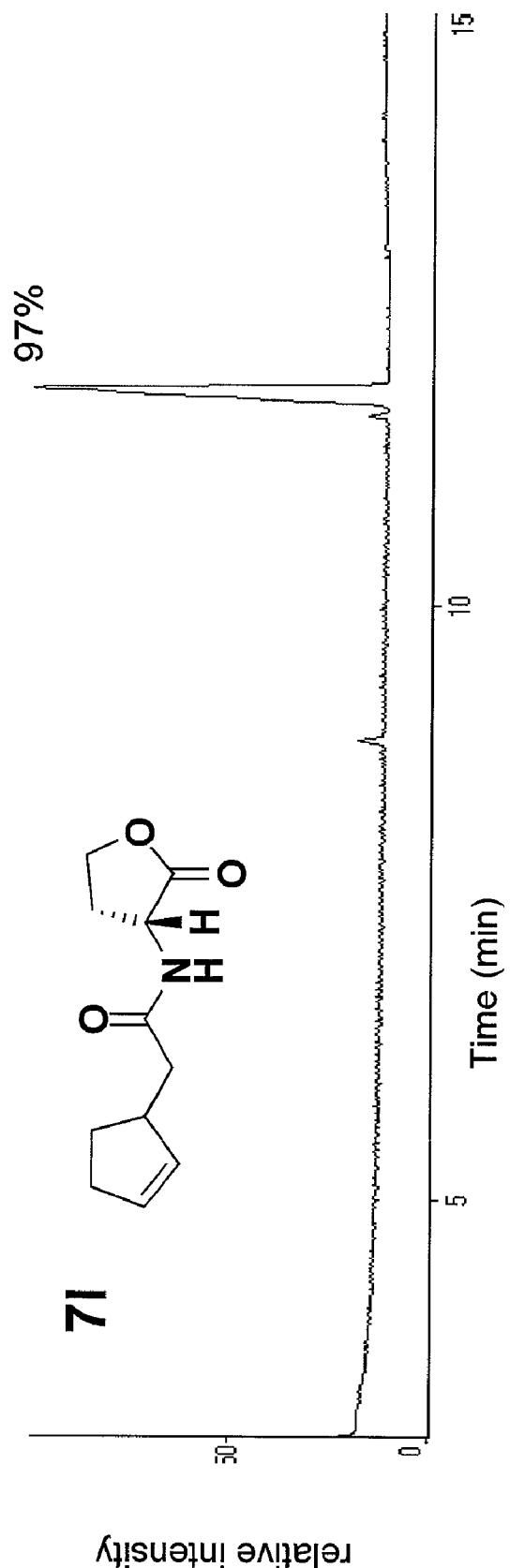


FIG. 13A

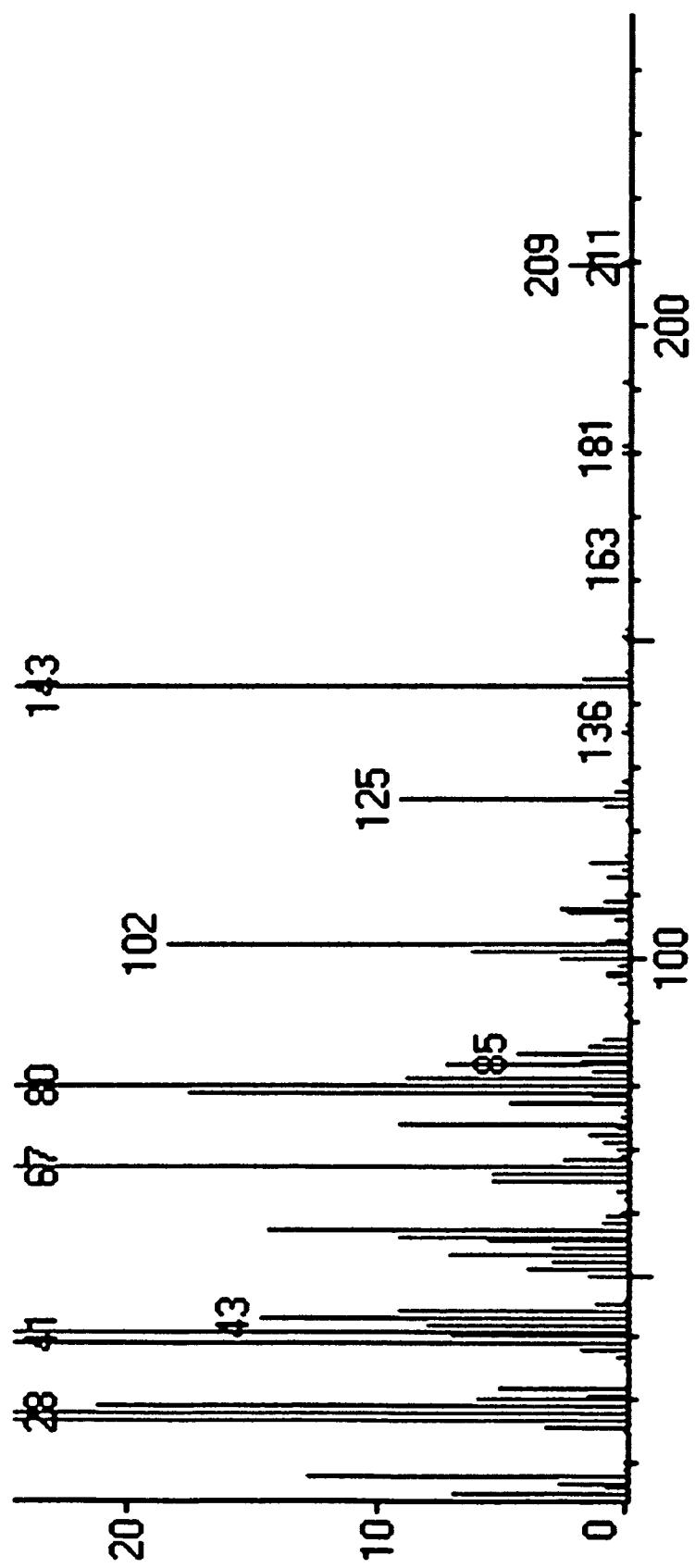


FIG. 13B

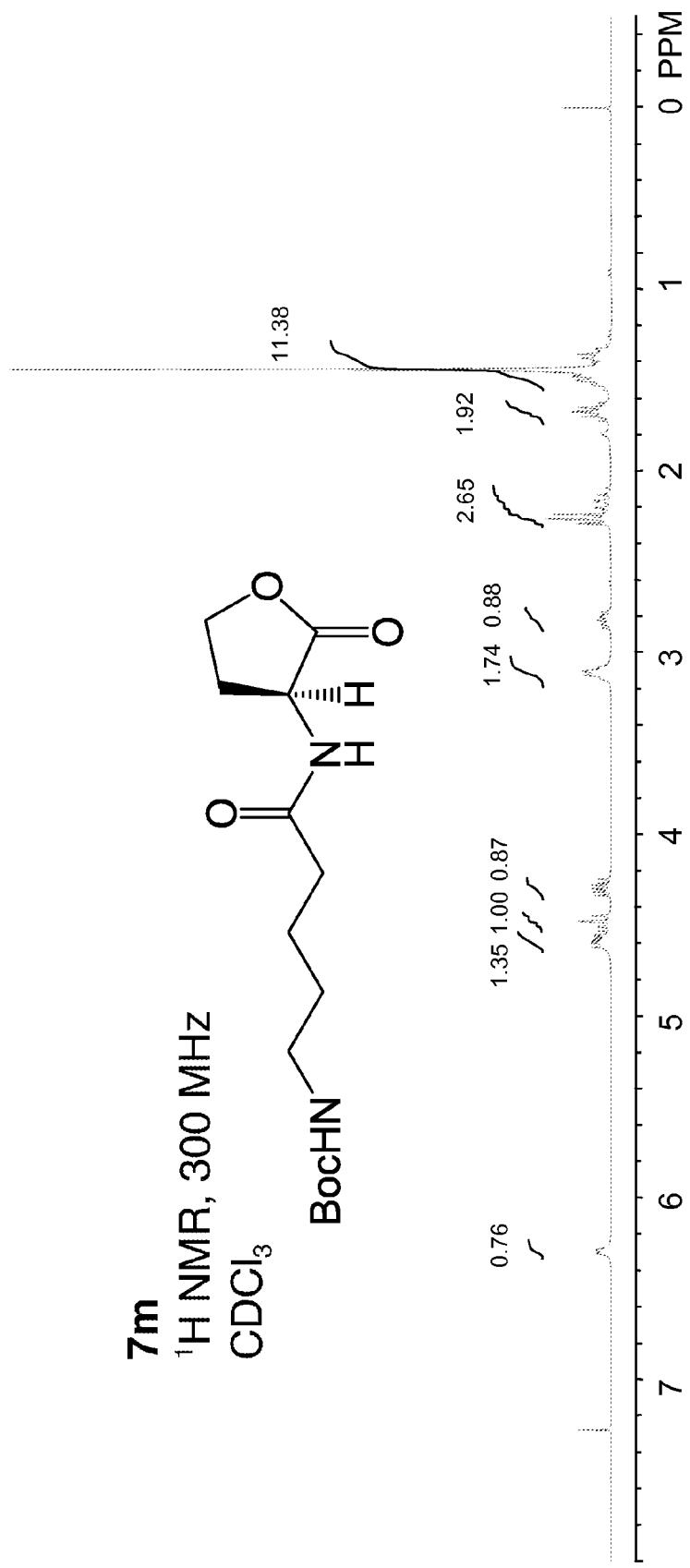


FIG. 14

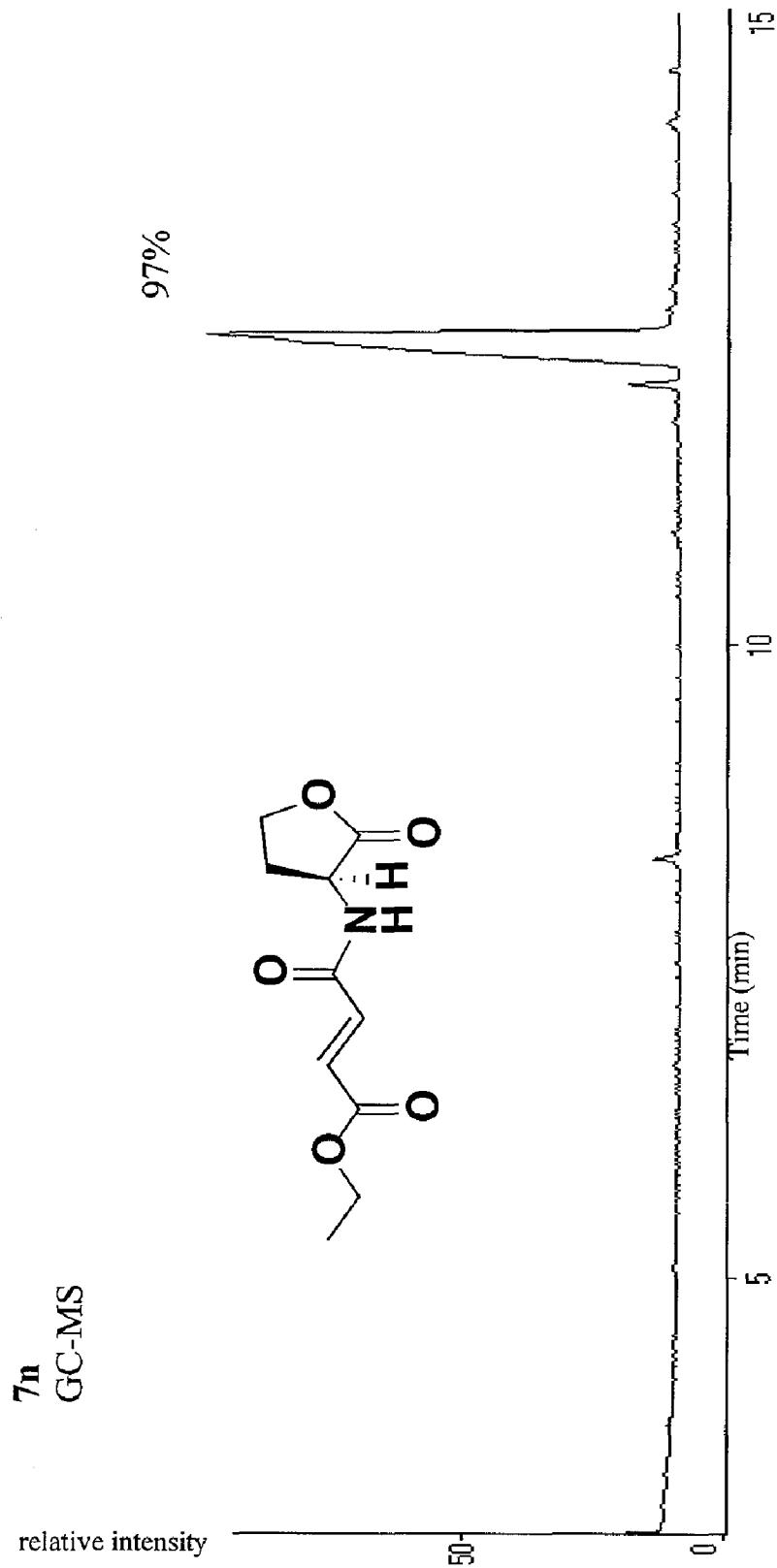


FIG. 15A

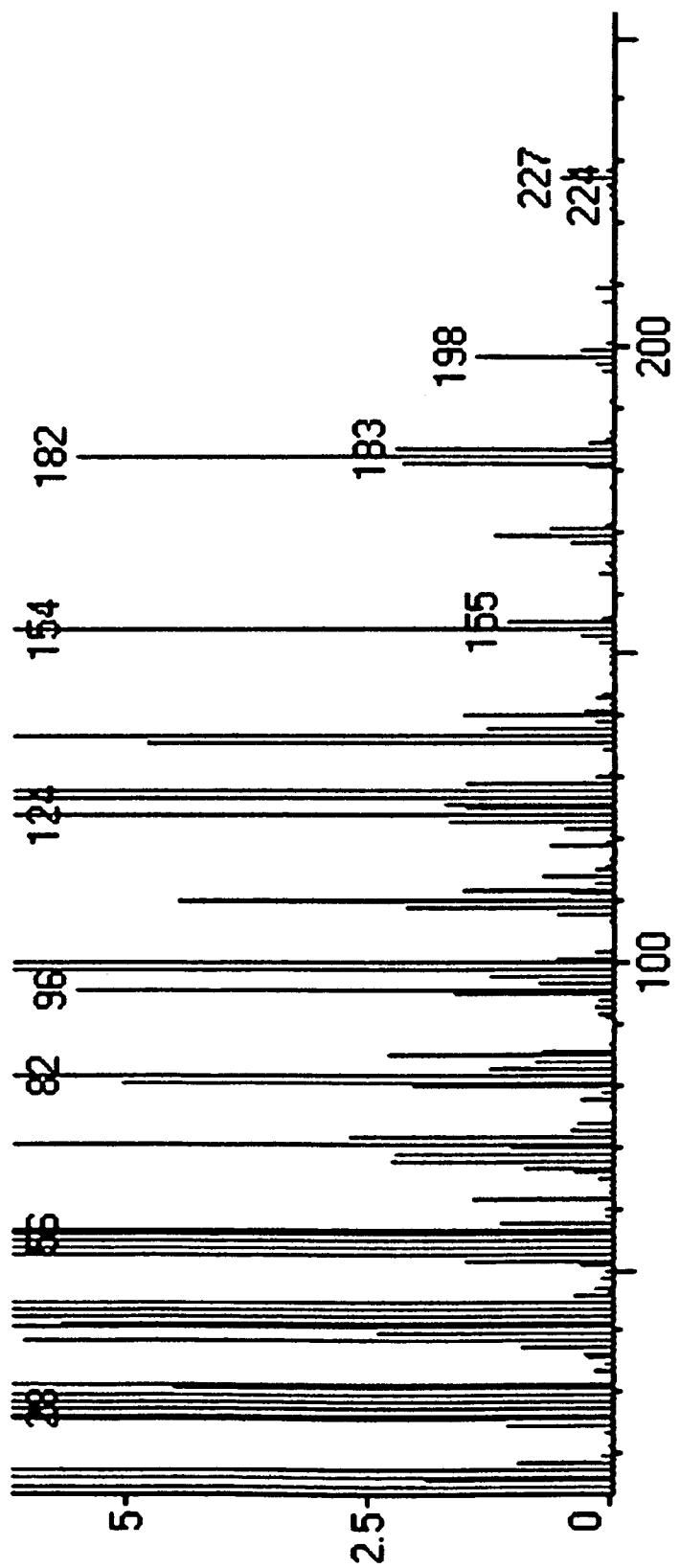


FIG. 15B

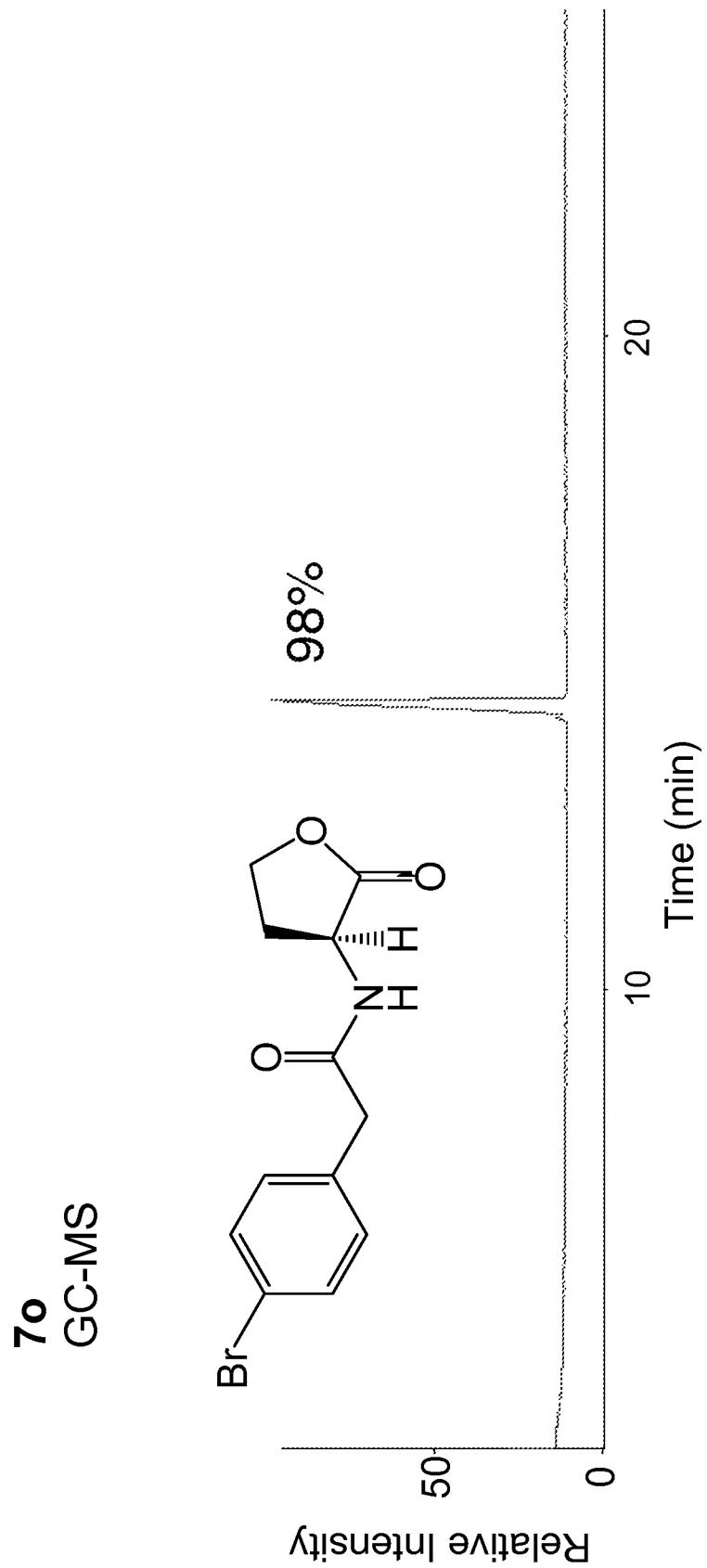


FIG. 16A

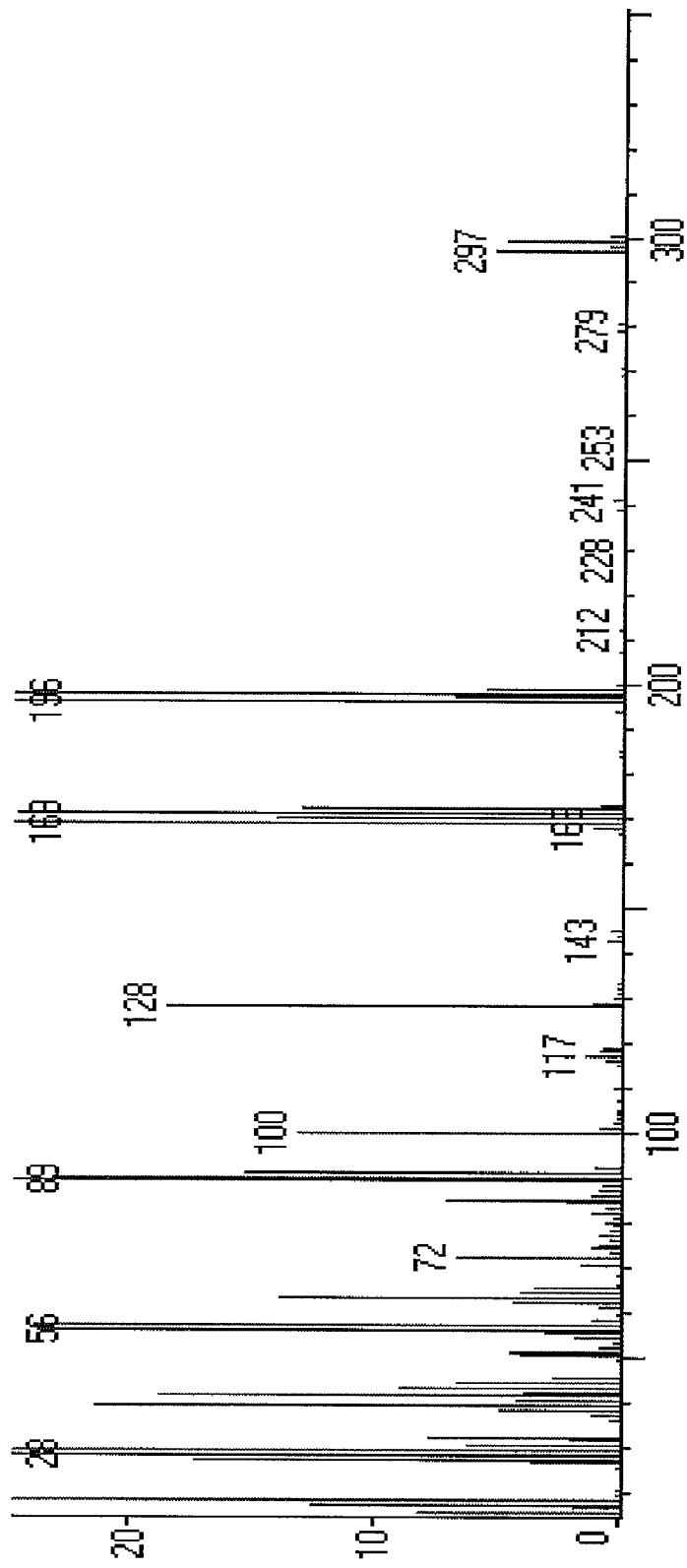


FIG. 16B

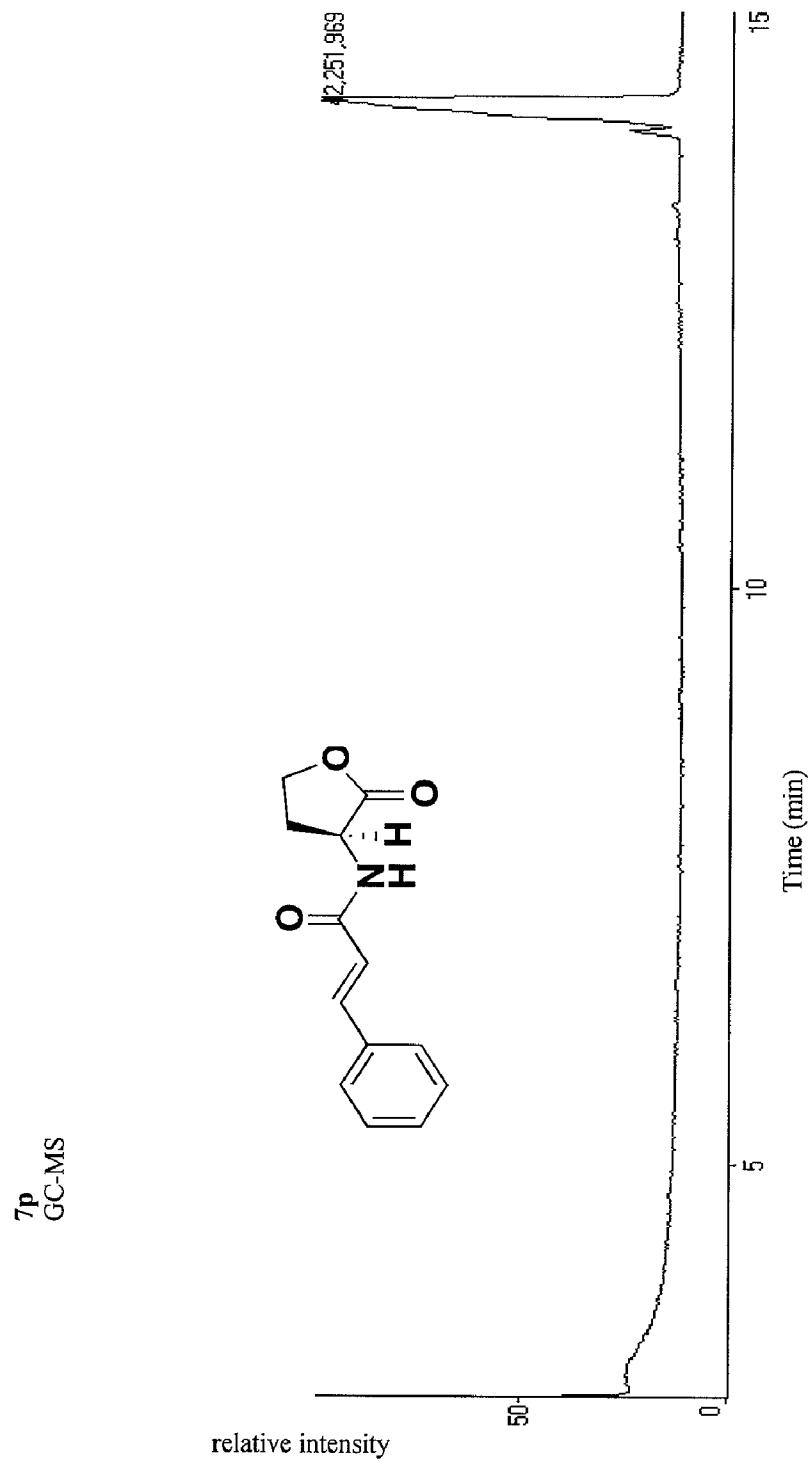


FIG. 17A

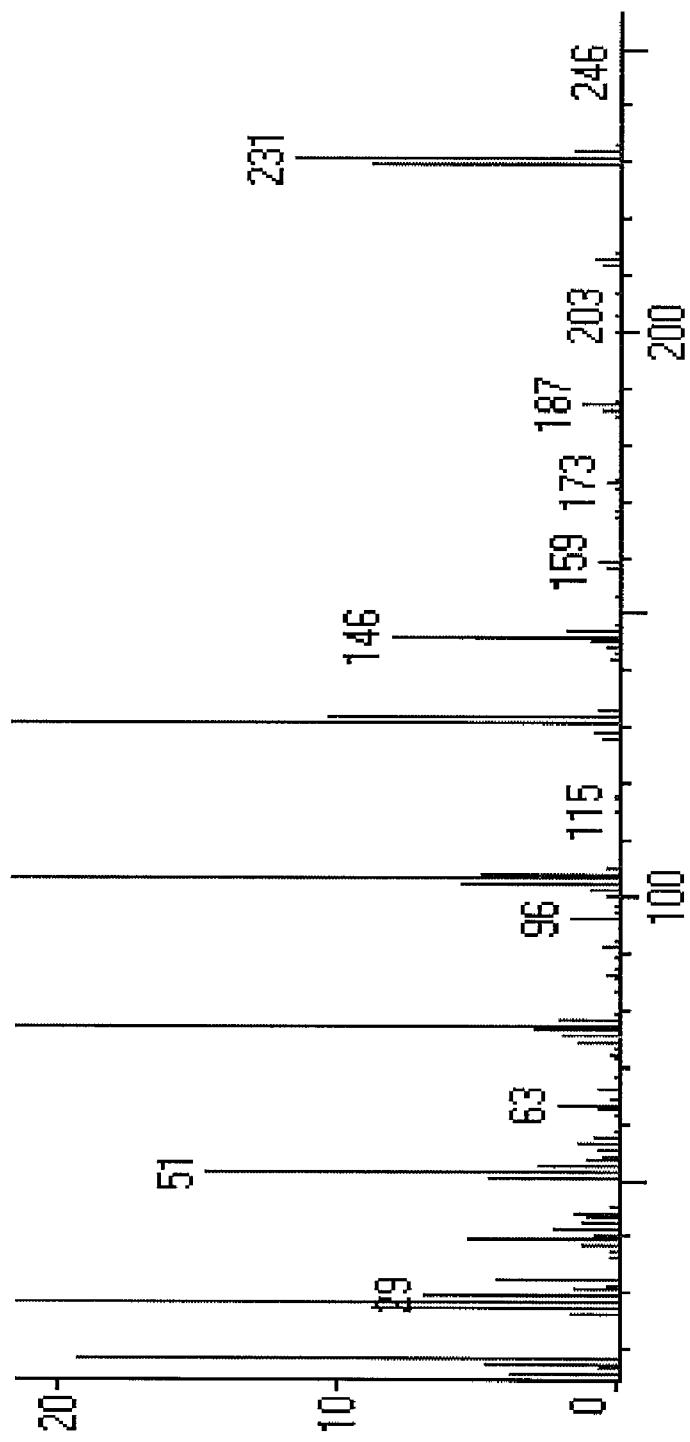


FIG. 17B

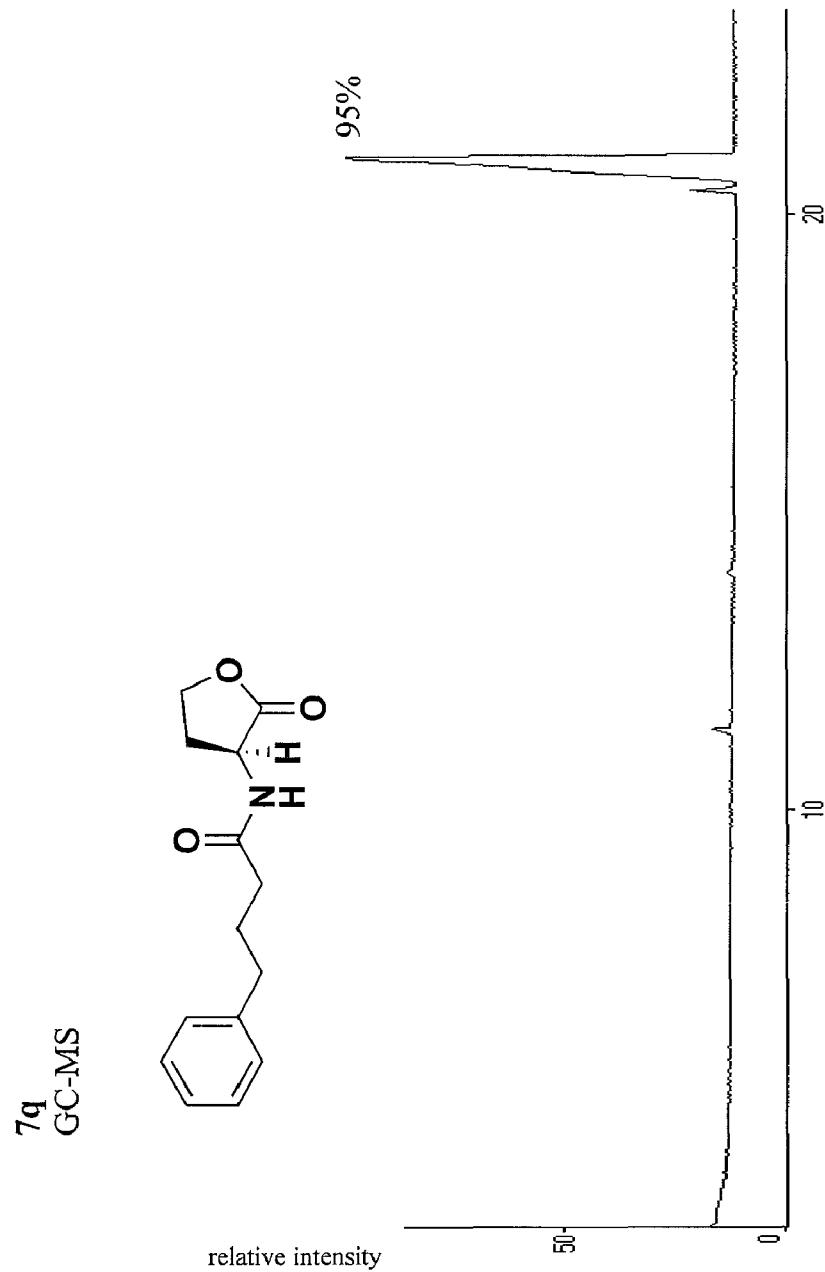


FIG. 18A

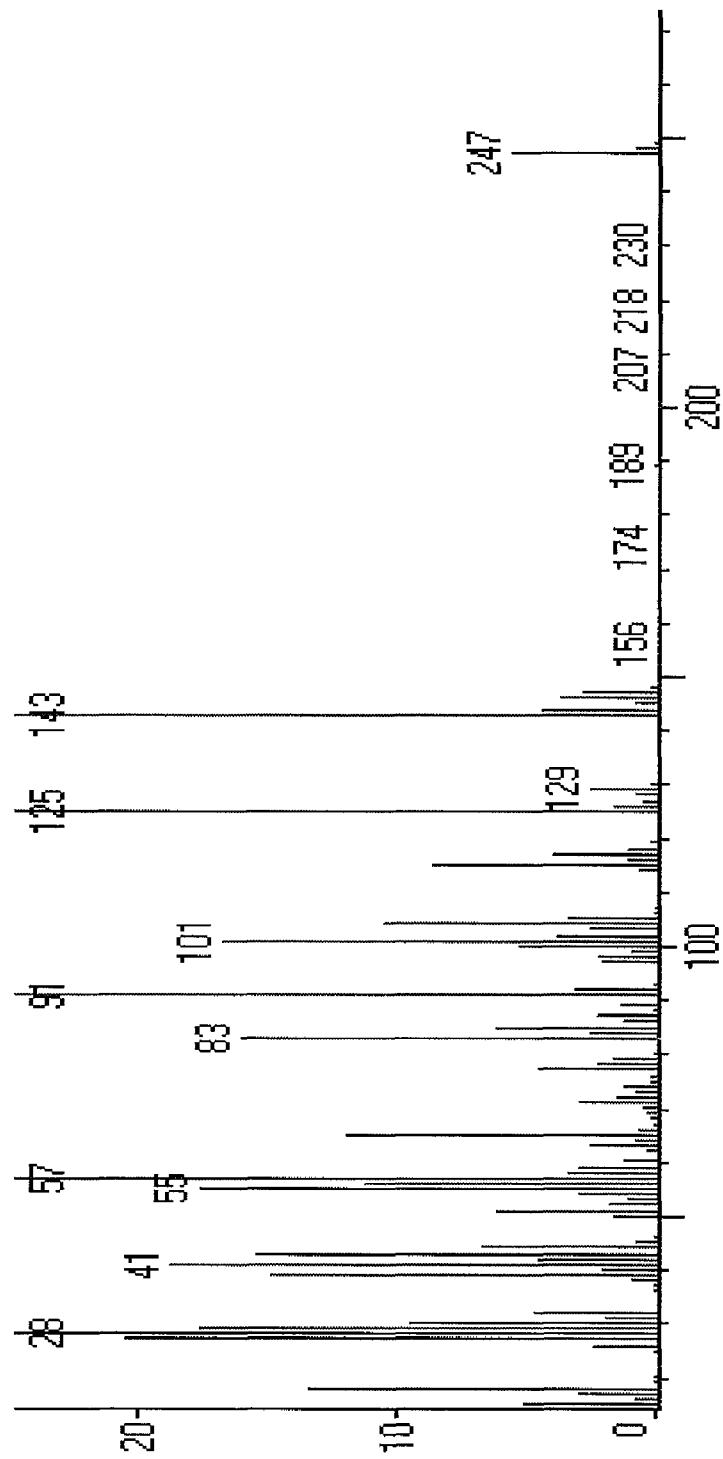


FIG. 18B

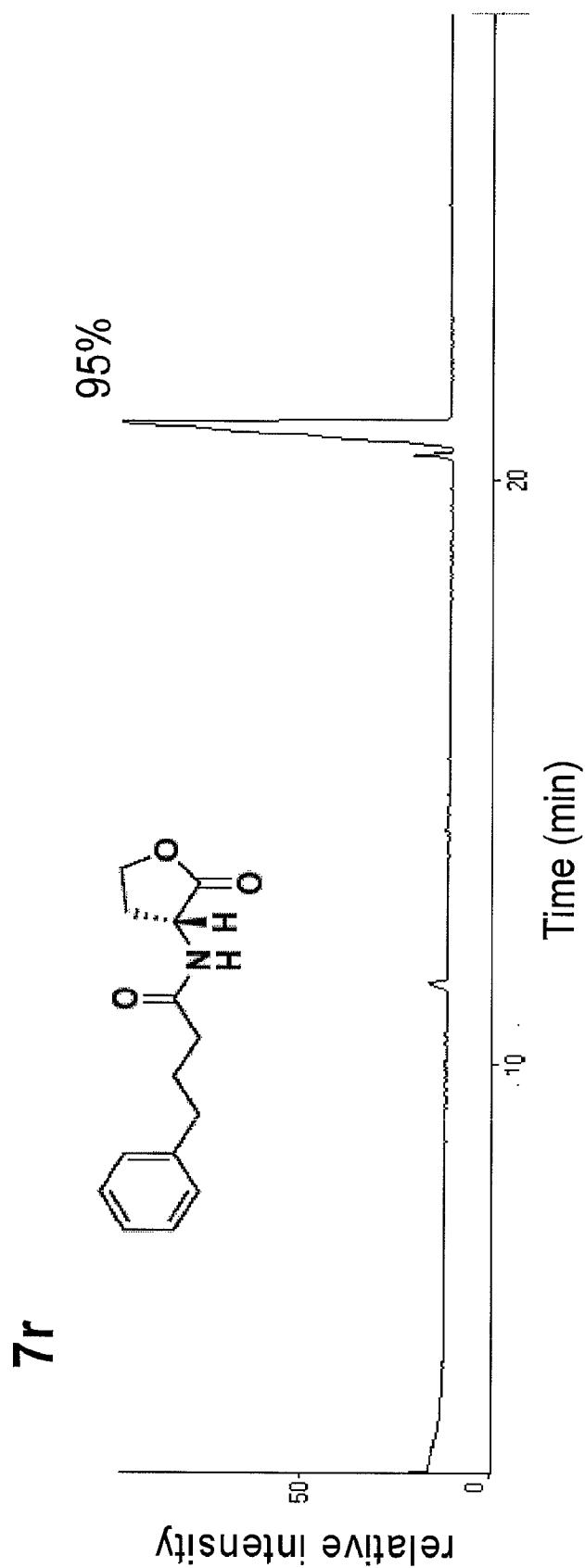


FIG. 19A

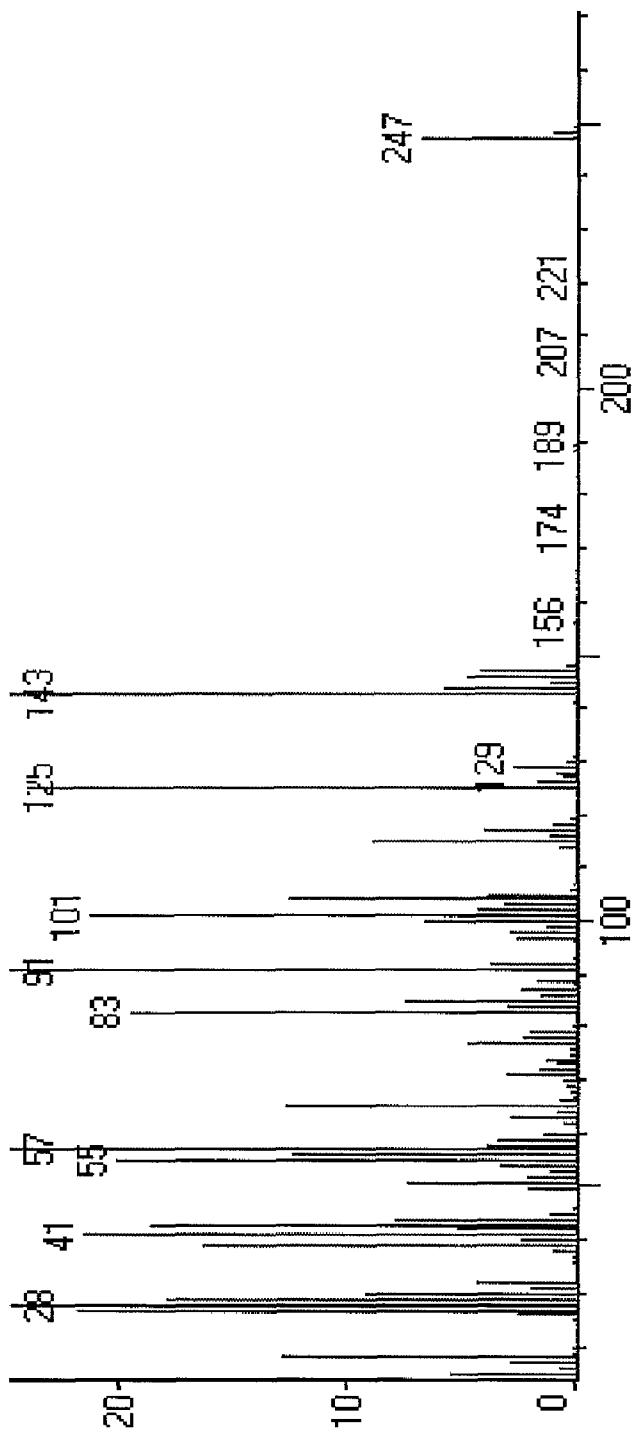


FIG. 19B

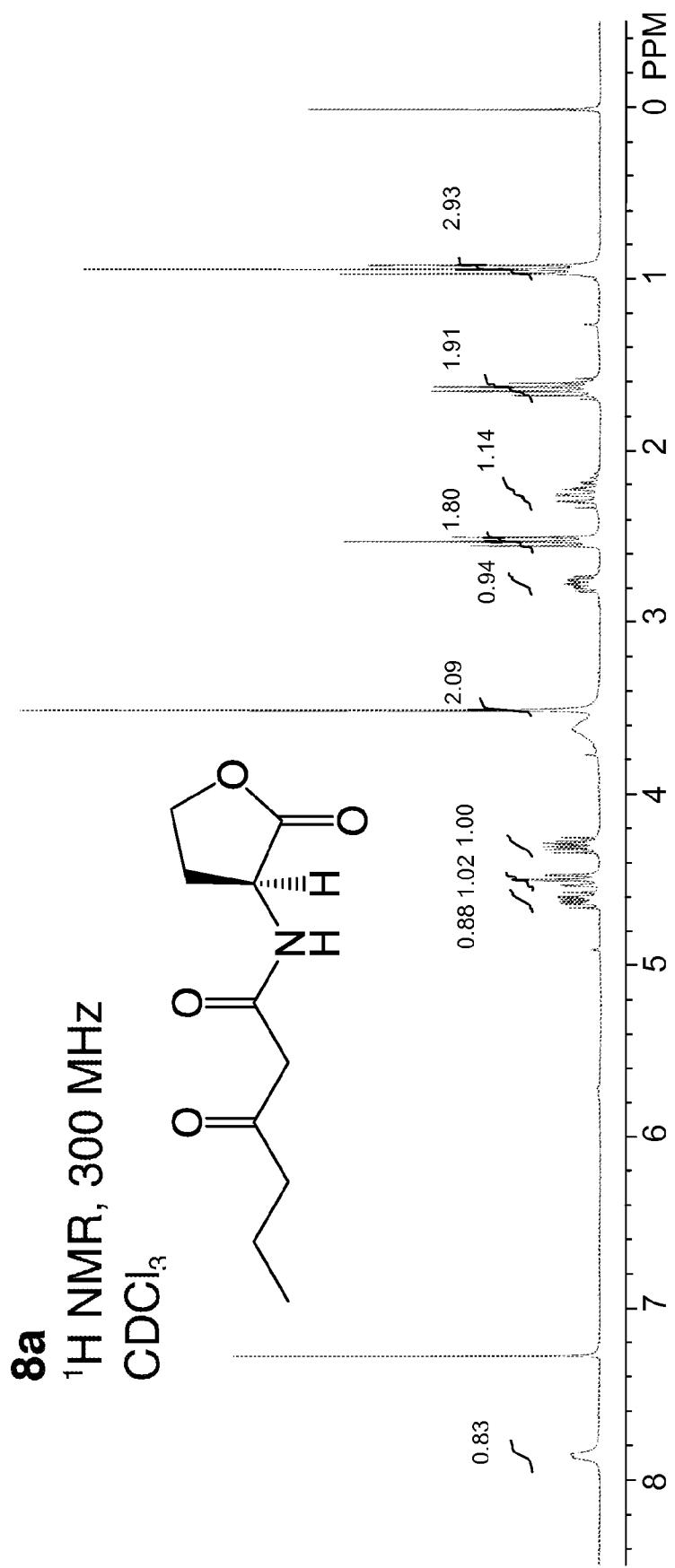


FIG. 20

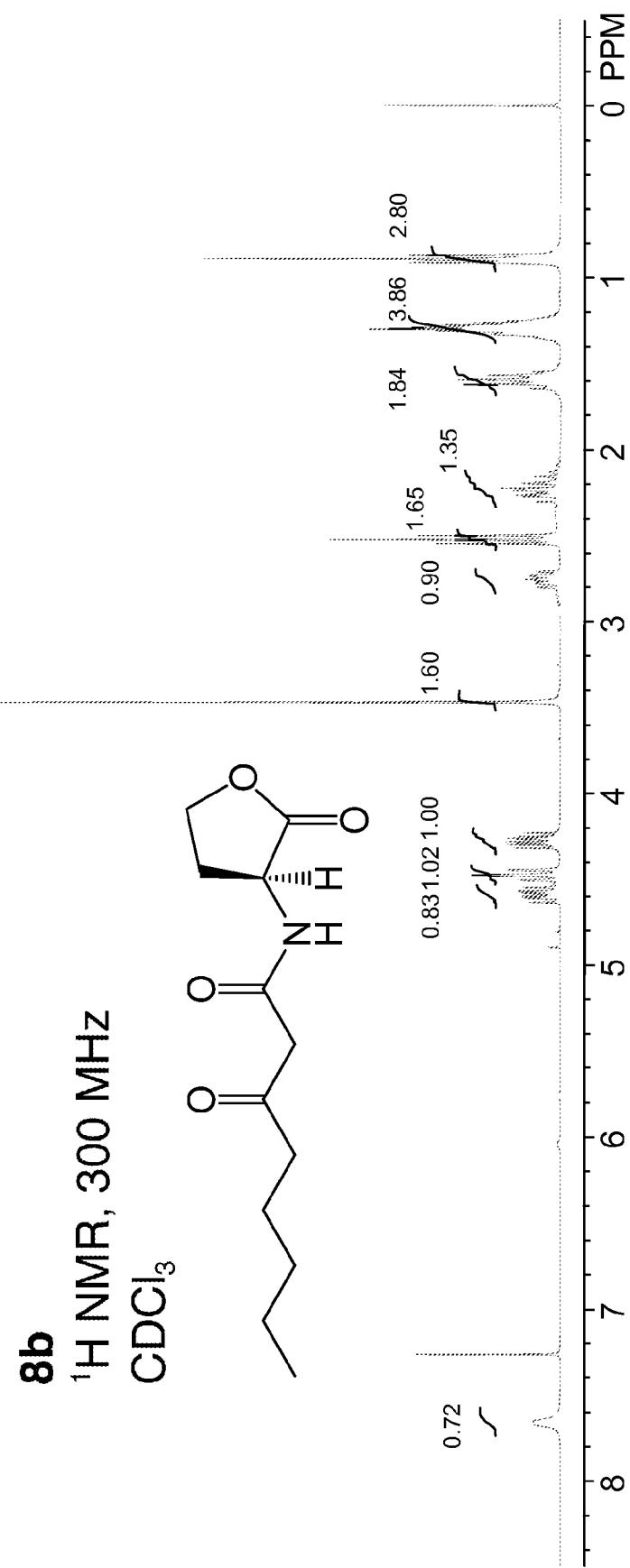


FIG. 21

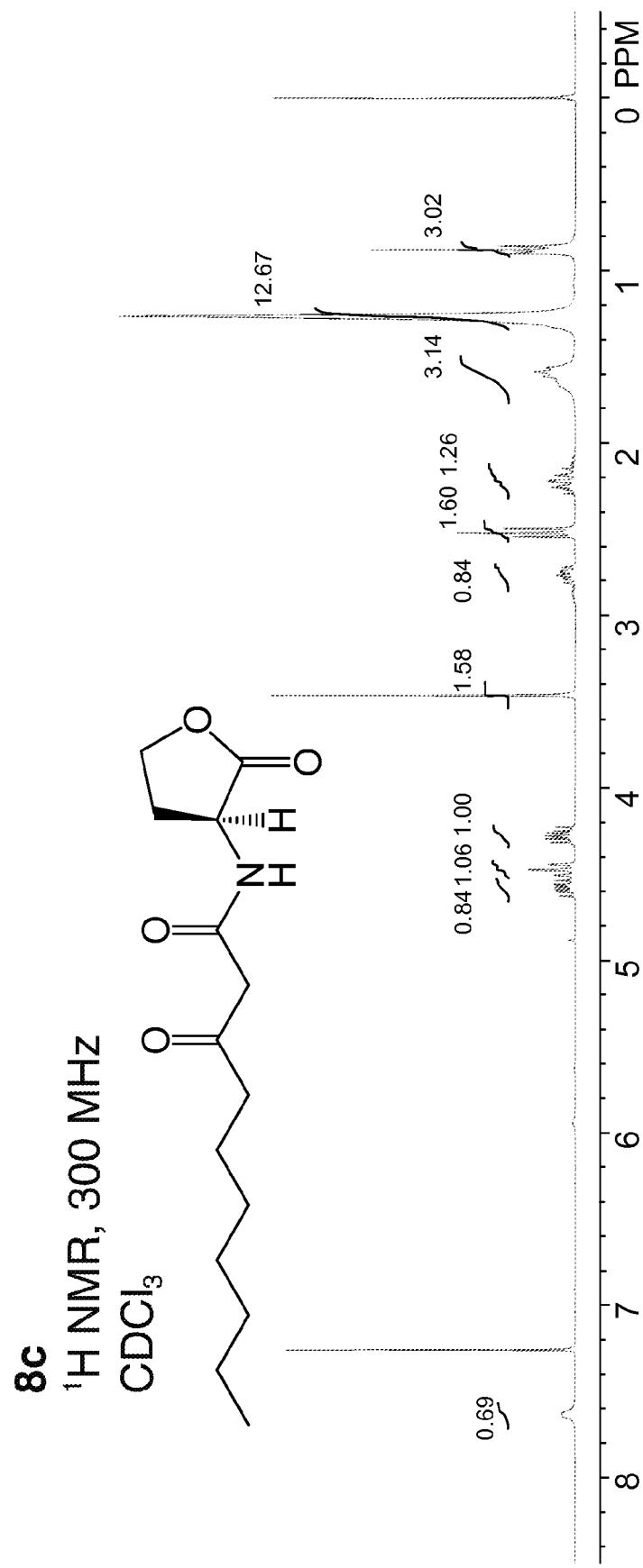


FIG. 22

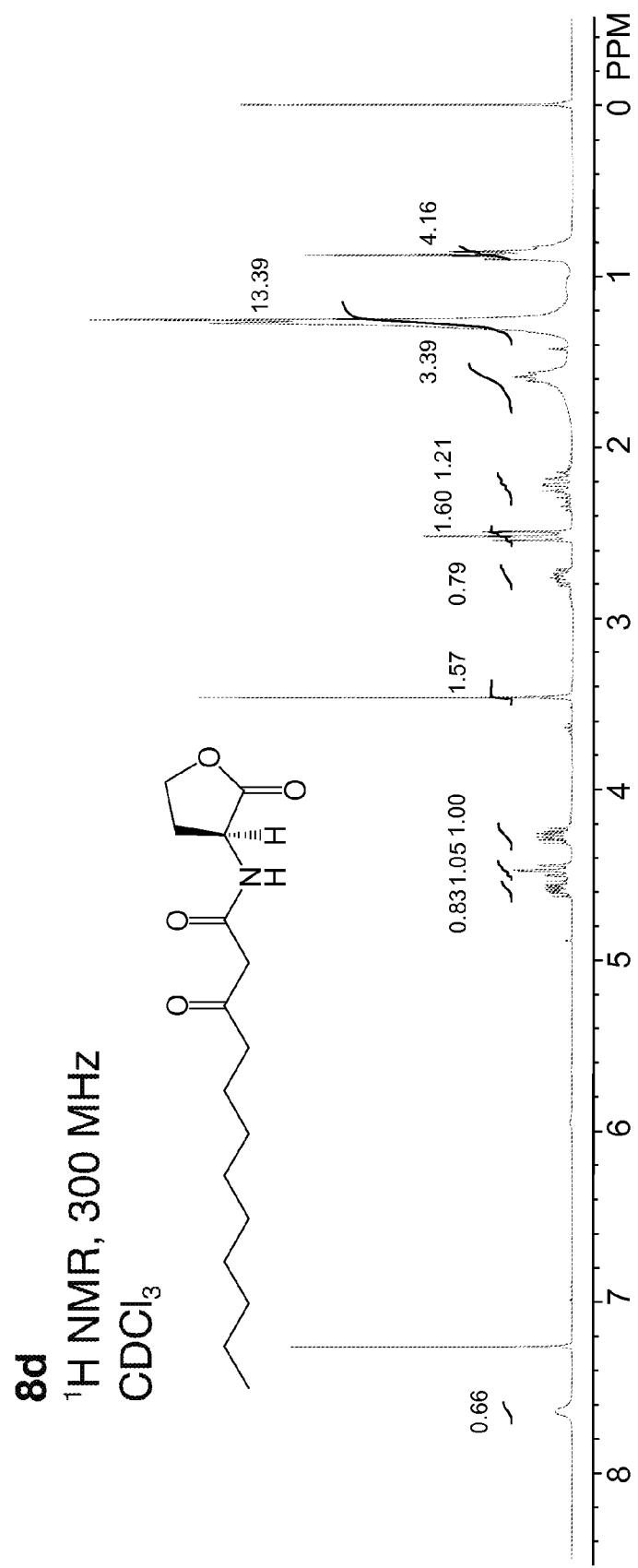


FIG. 23

8e
 ^1H NMR, 300 MHz
 CDCl_3

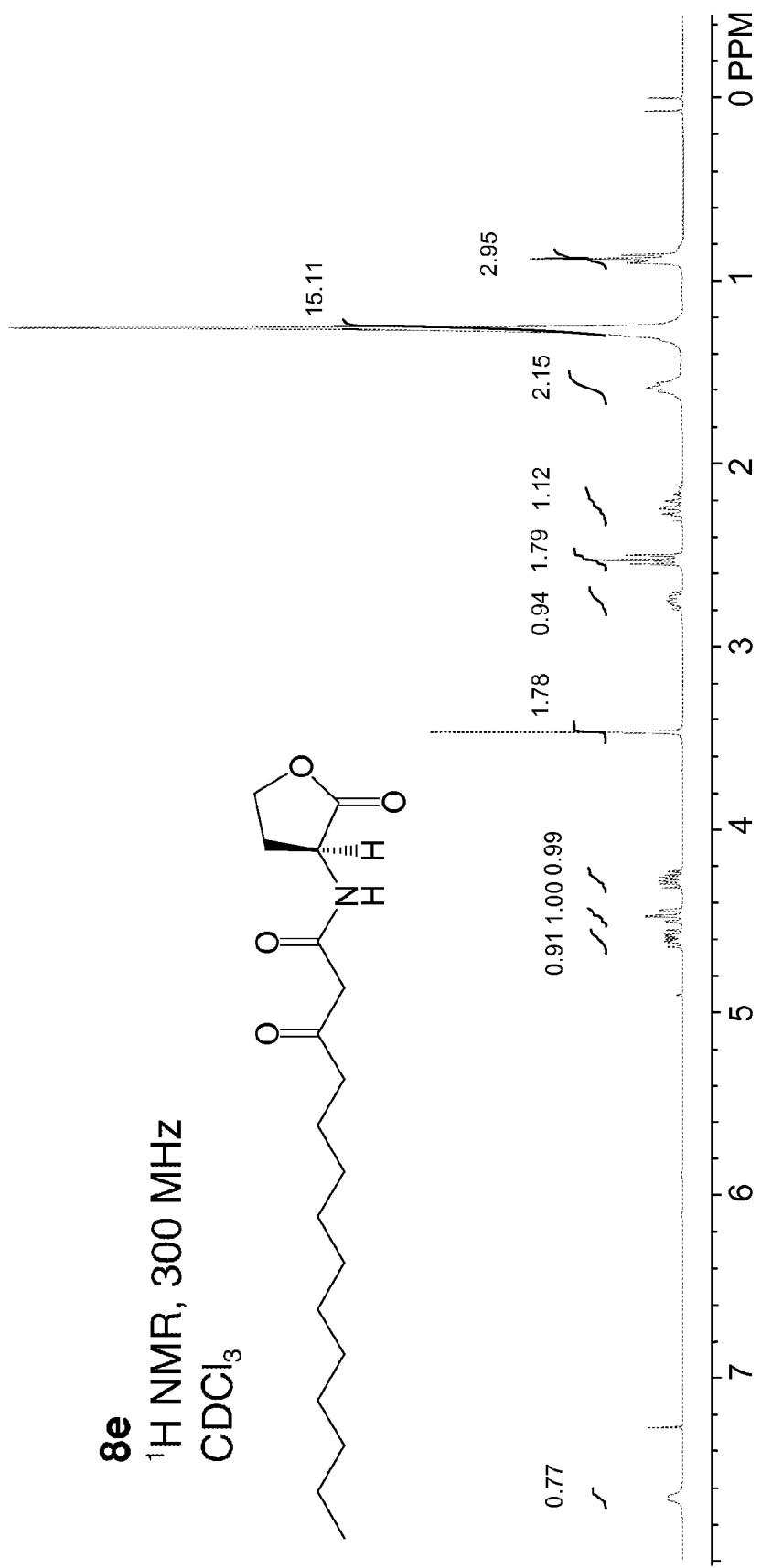
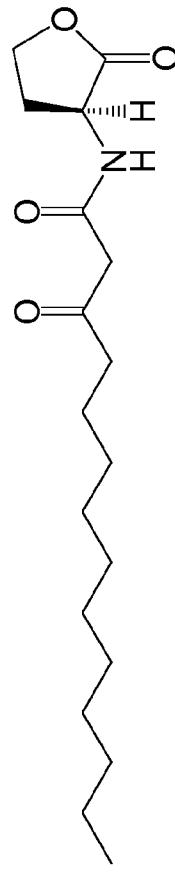


FIG. 24

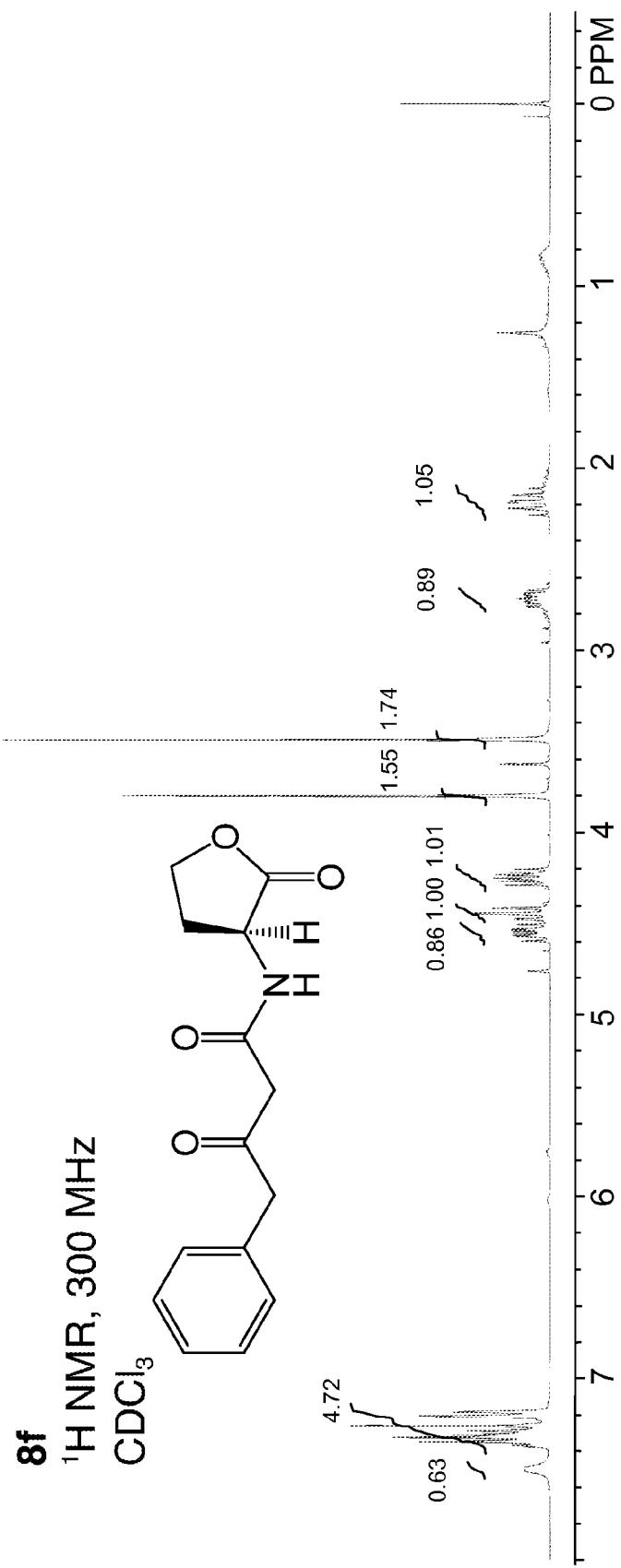


FIG. 25

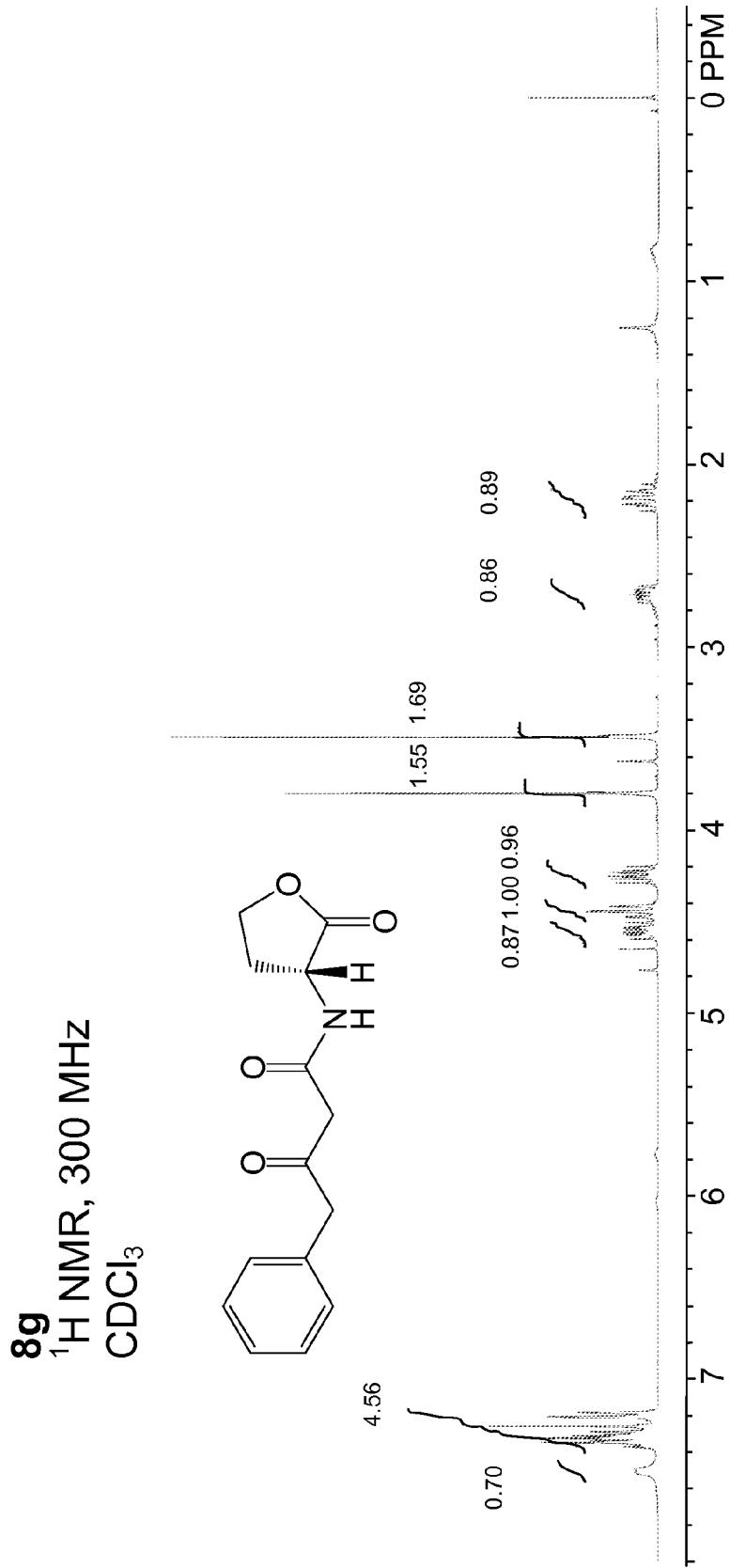


FIG. 26

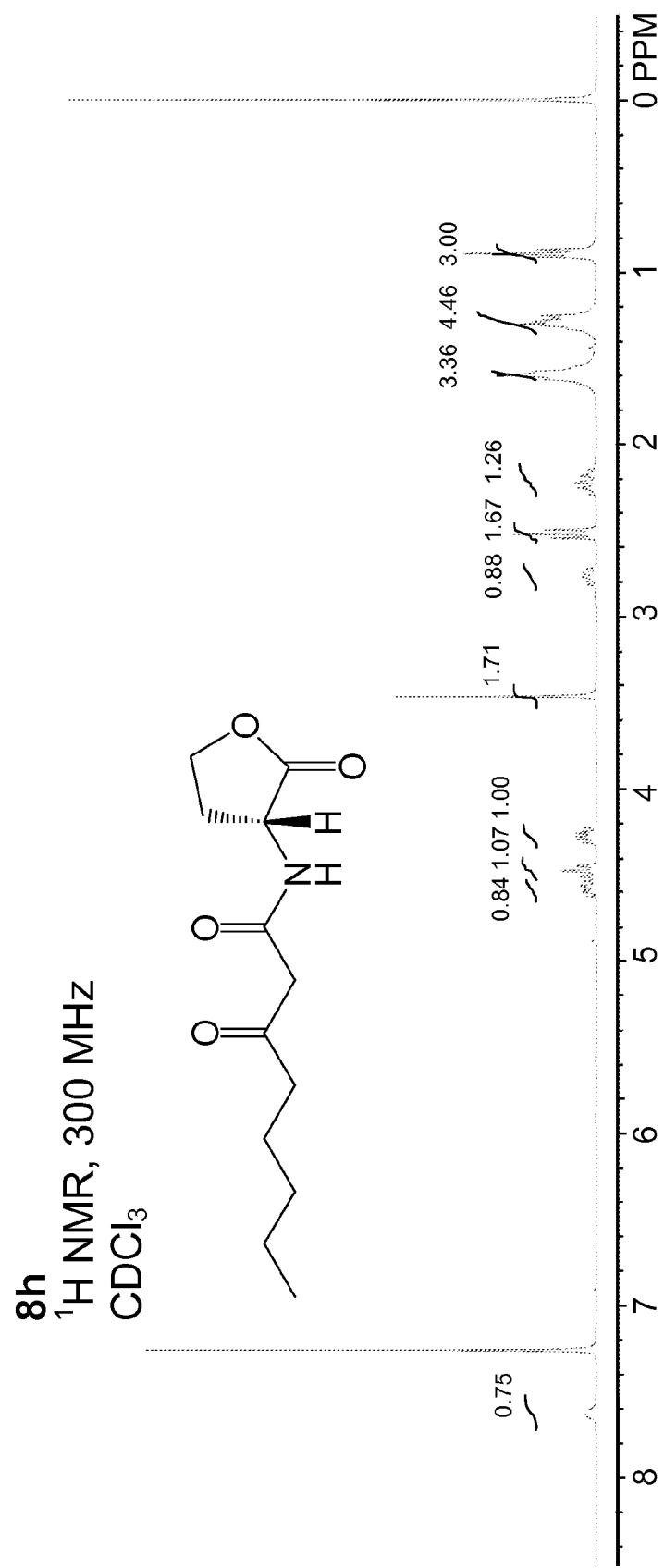


FIG. 27

a (R)-(+)-Methoxy- α -trifluoromethylphenylacetanoyl-D-homoserine lactone

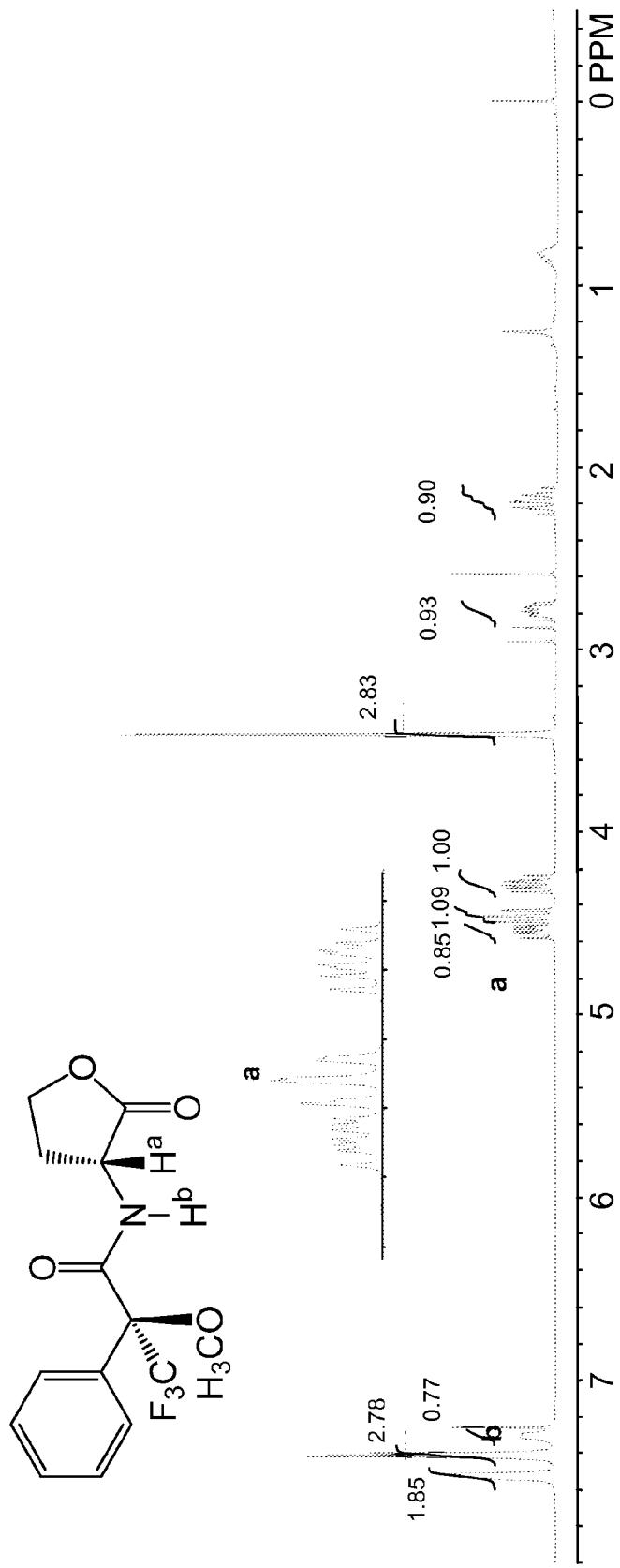


FIG. 28A

b (R)-(+)- α -Methoxy- α -trifluoromethylphenylacetanoyl-L-homoserine lactone

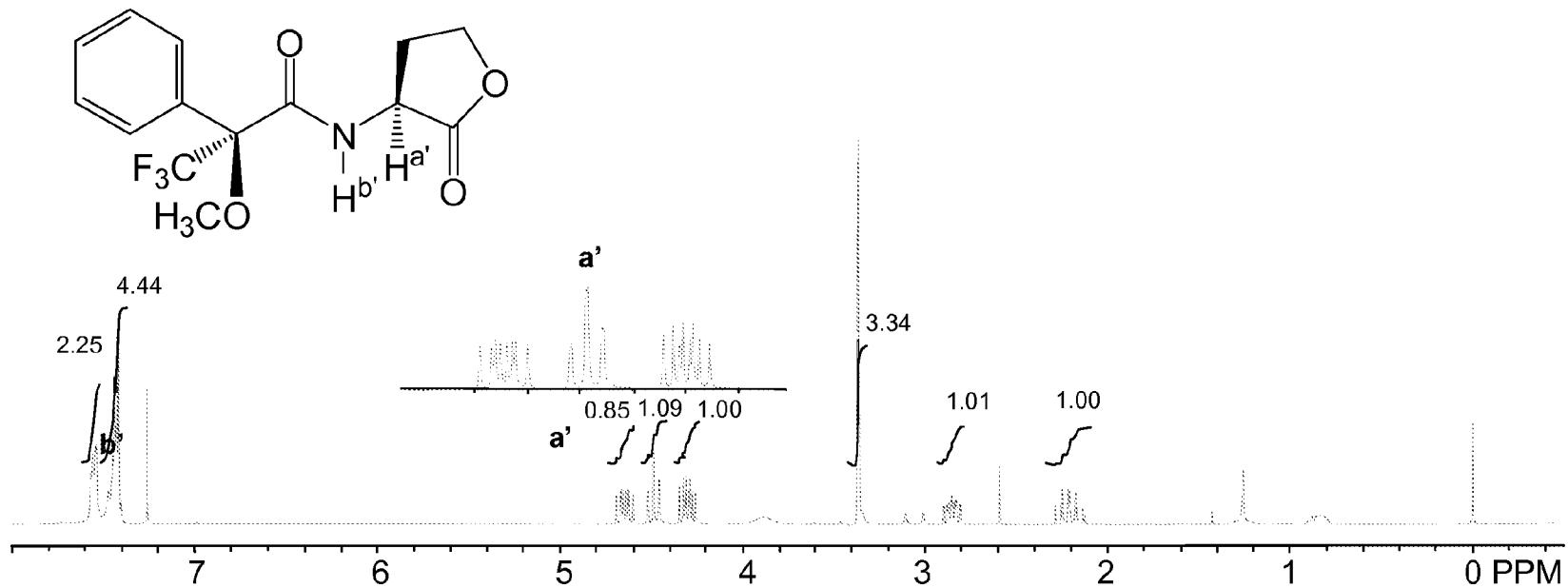


FIG. 28B

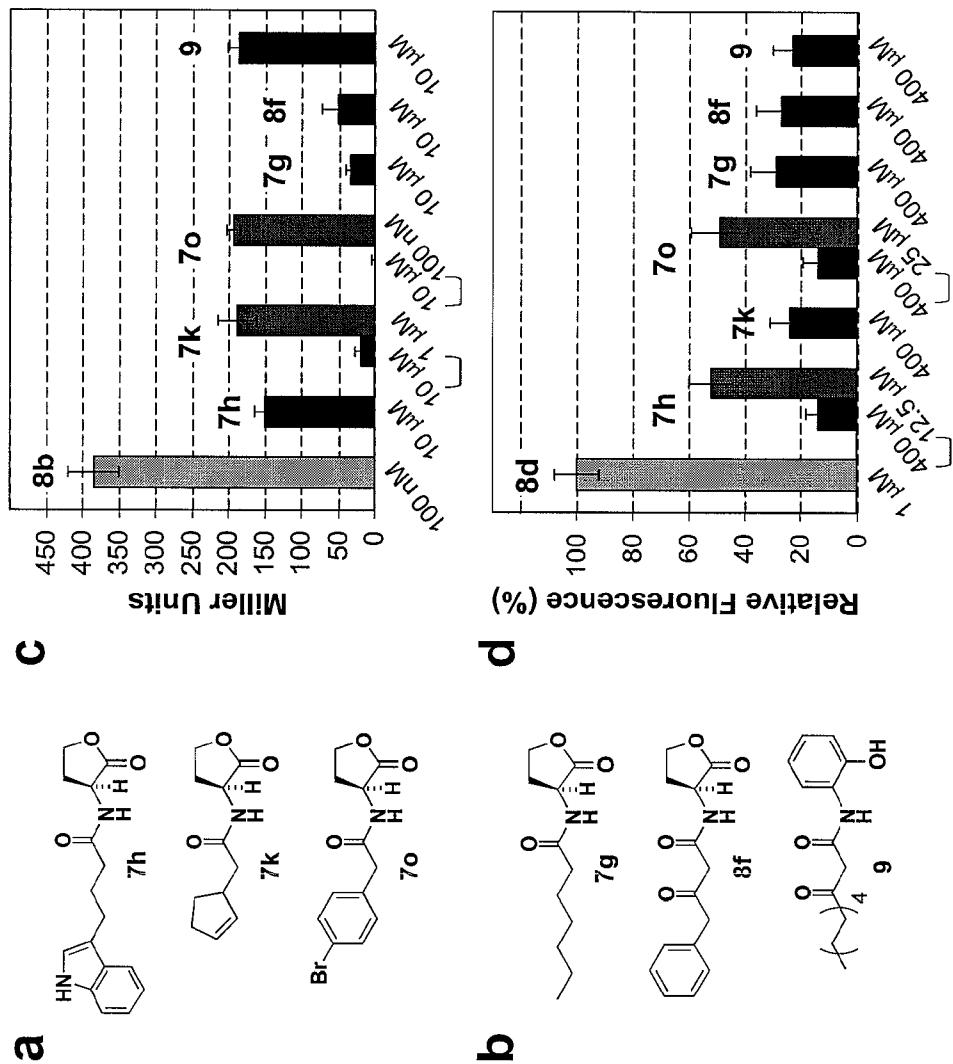


FIG. 29

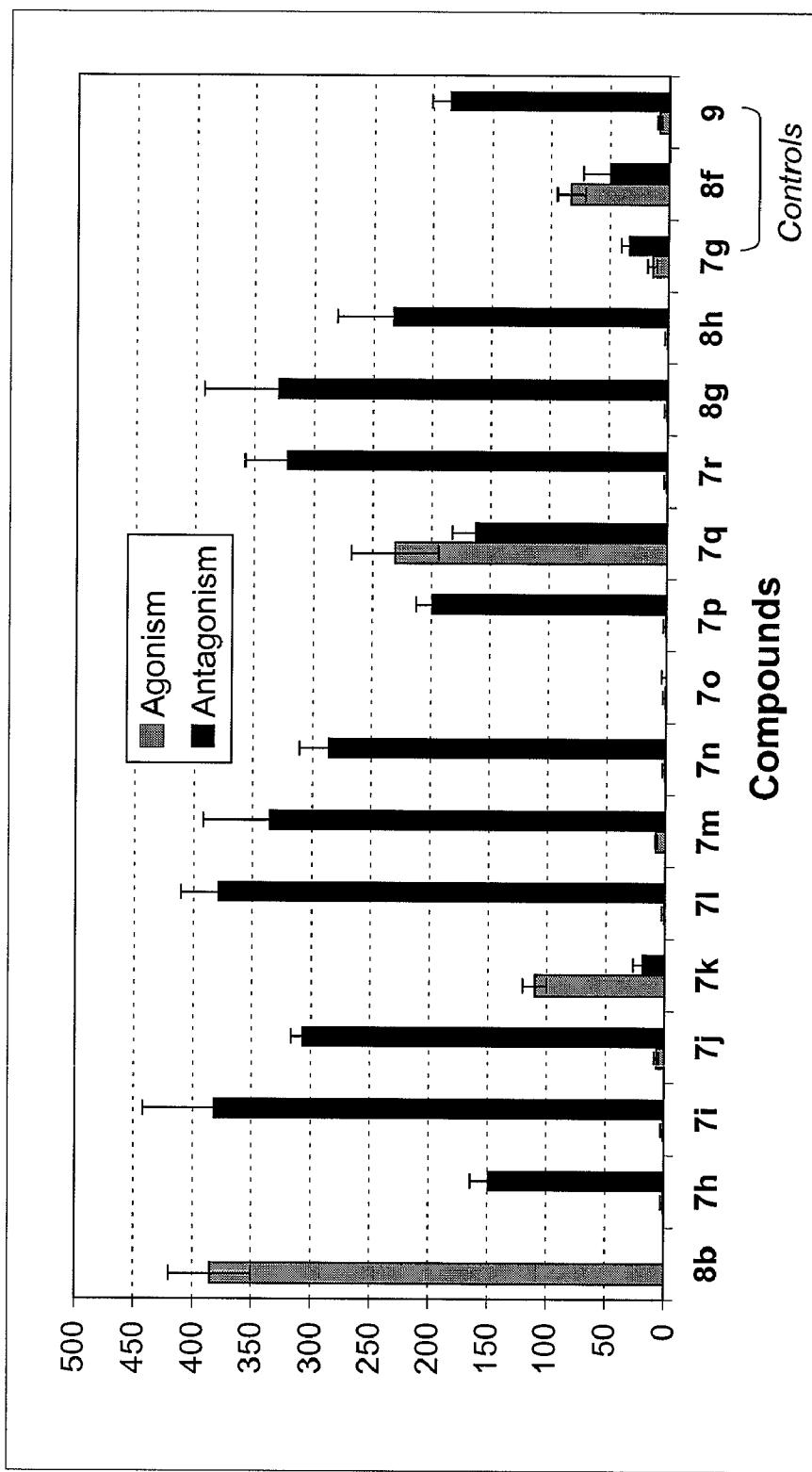


FIG. 30

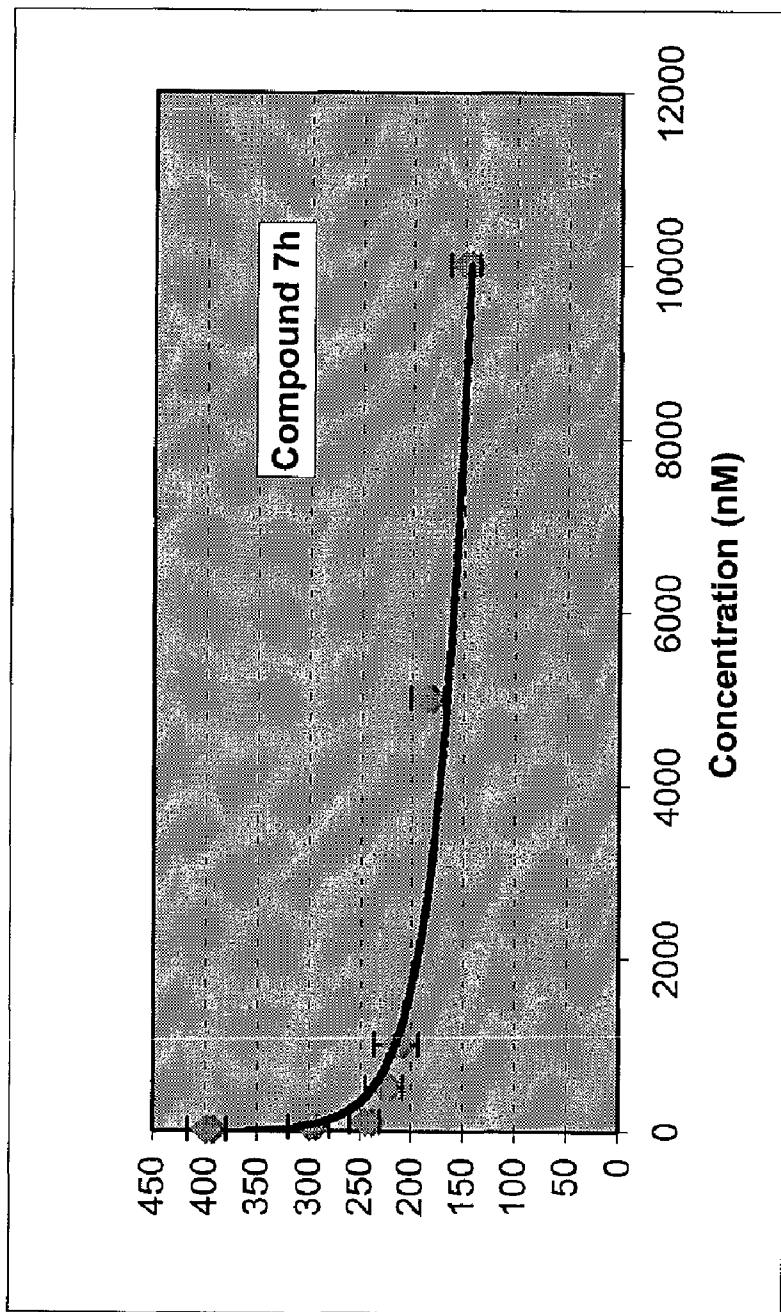


FIG. 31A

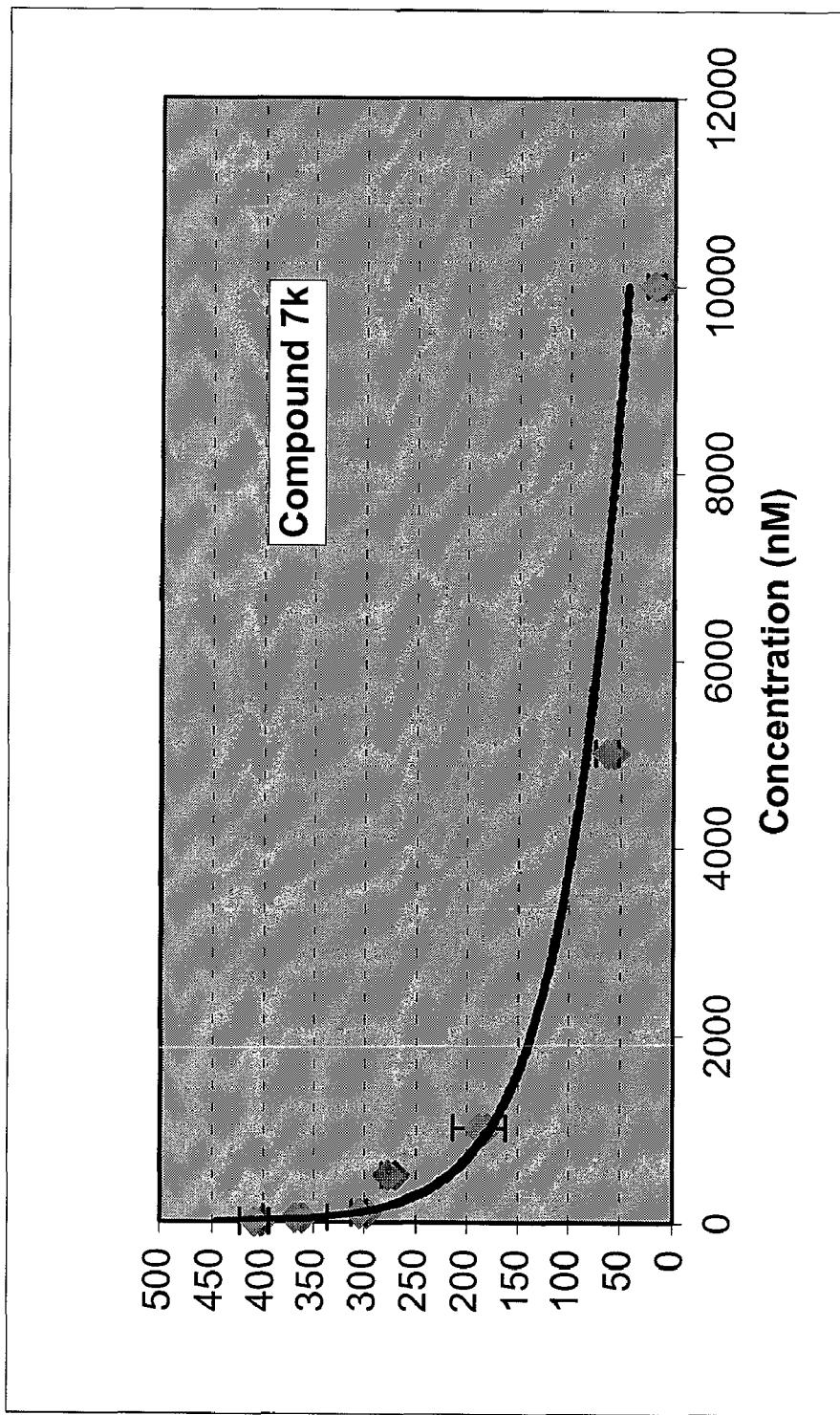


FIG. 31B

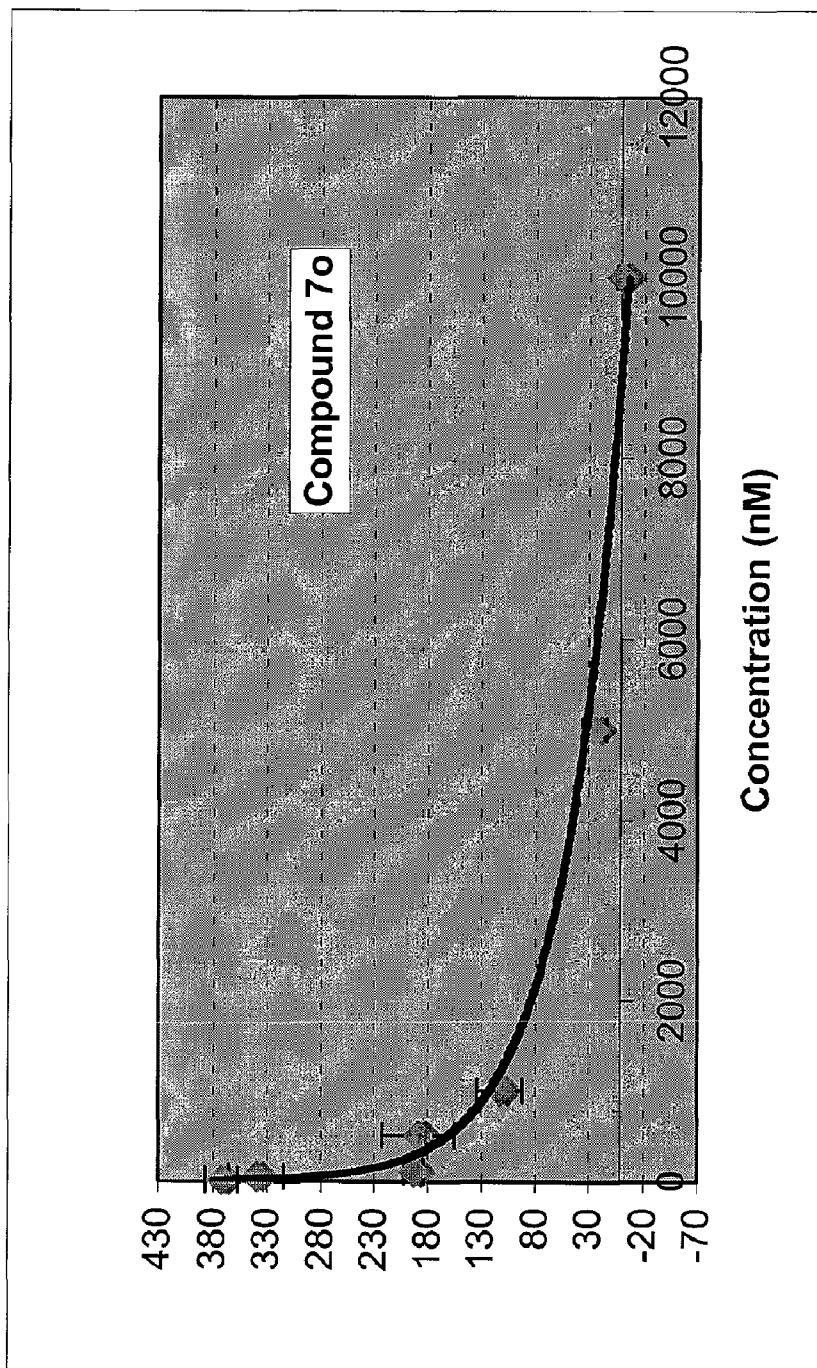


FIG. 31C

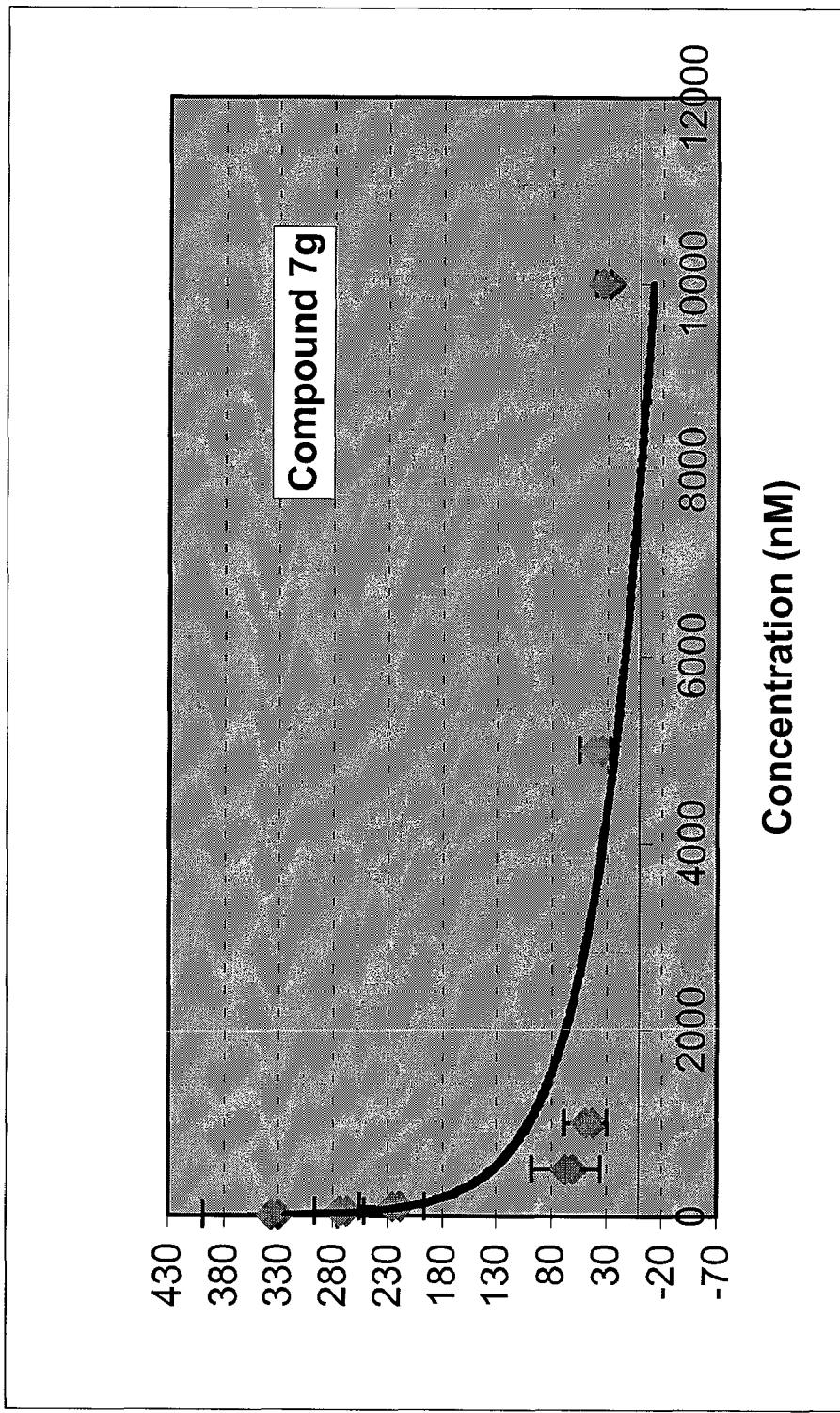


FIG. 31D

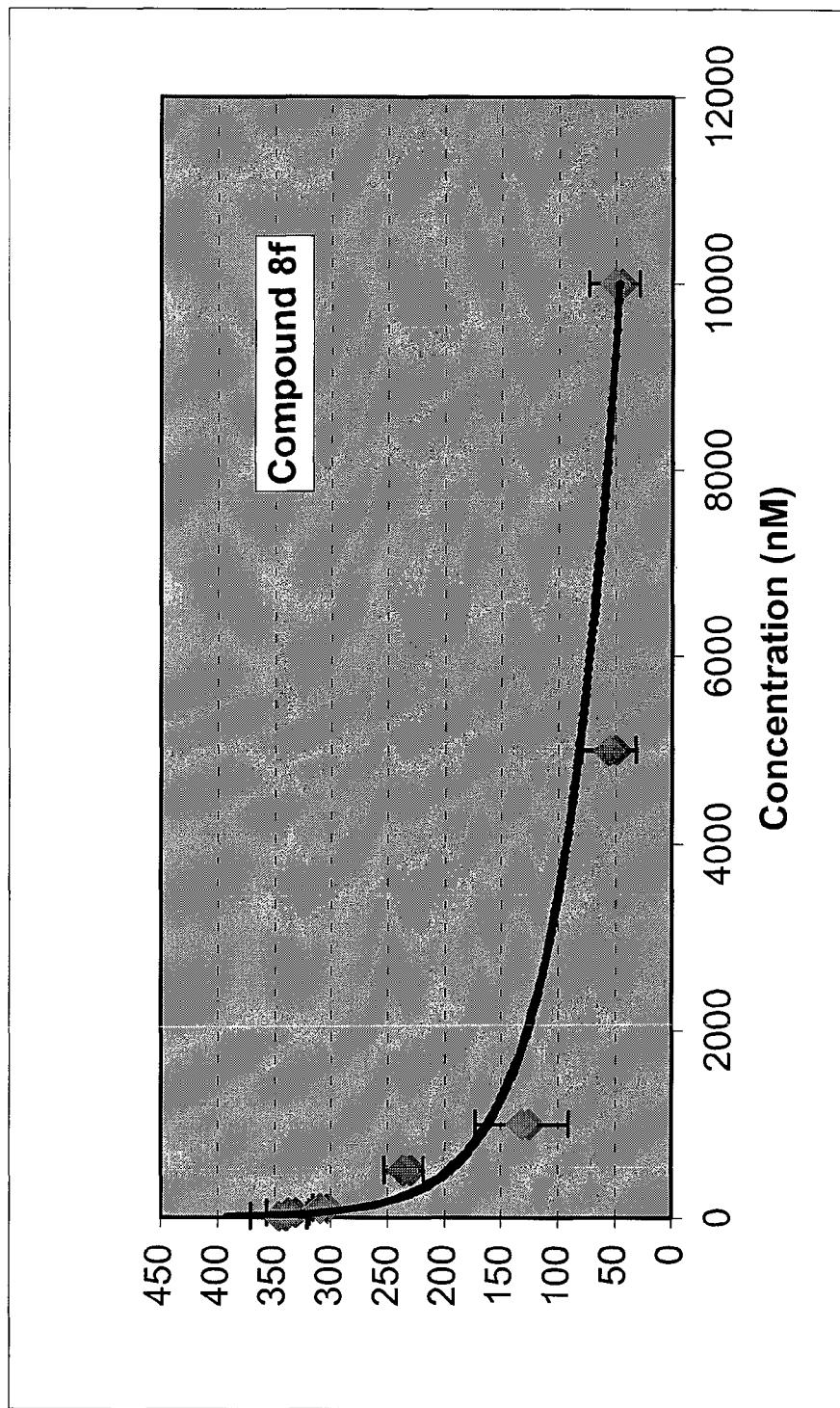


FIG. 31E

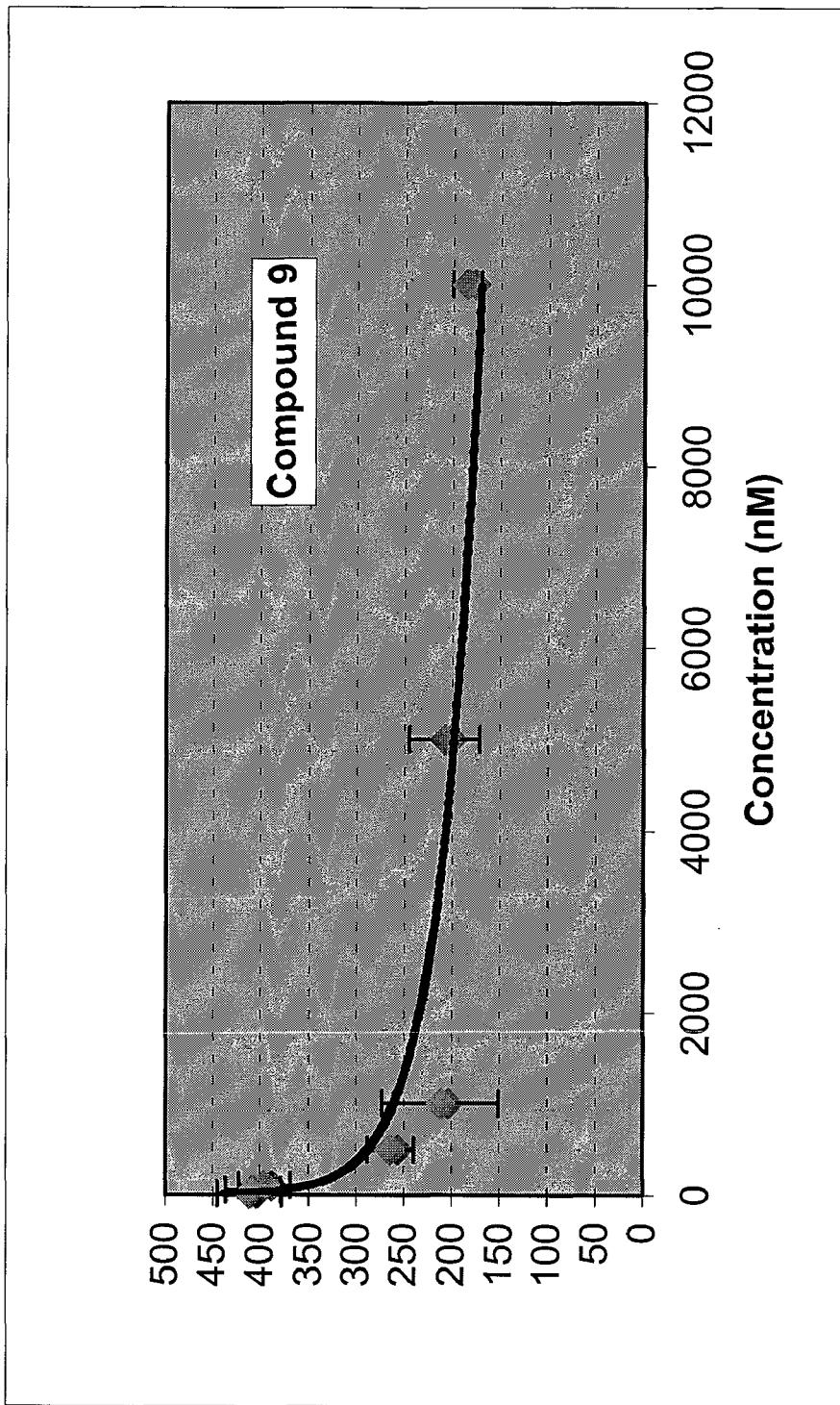


FIG. 31F

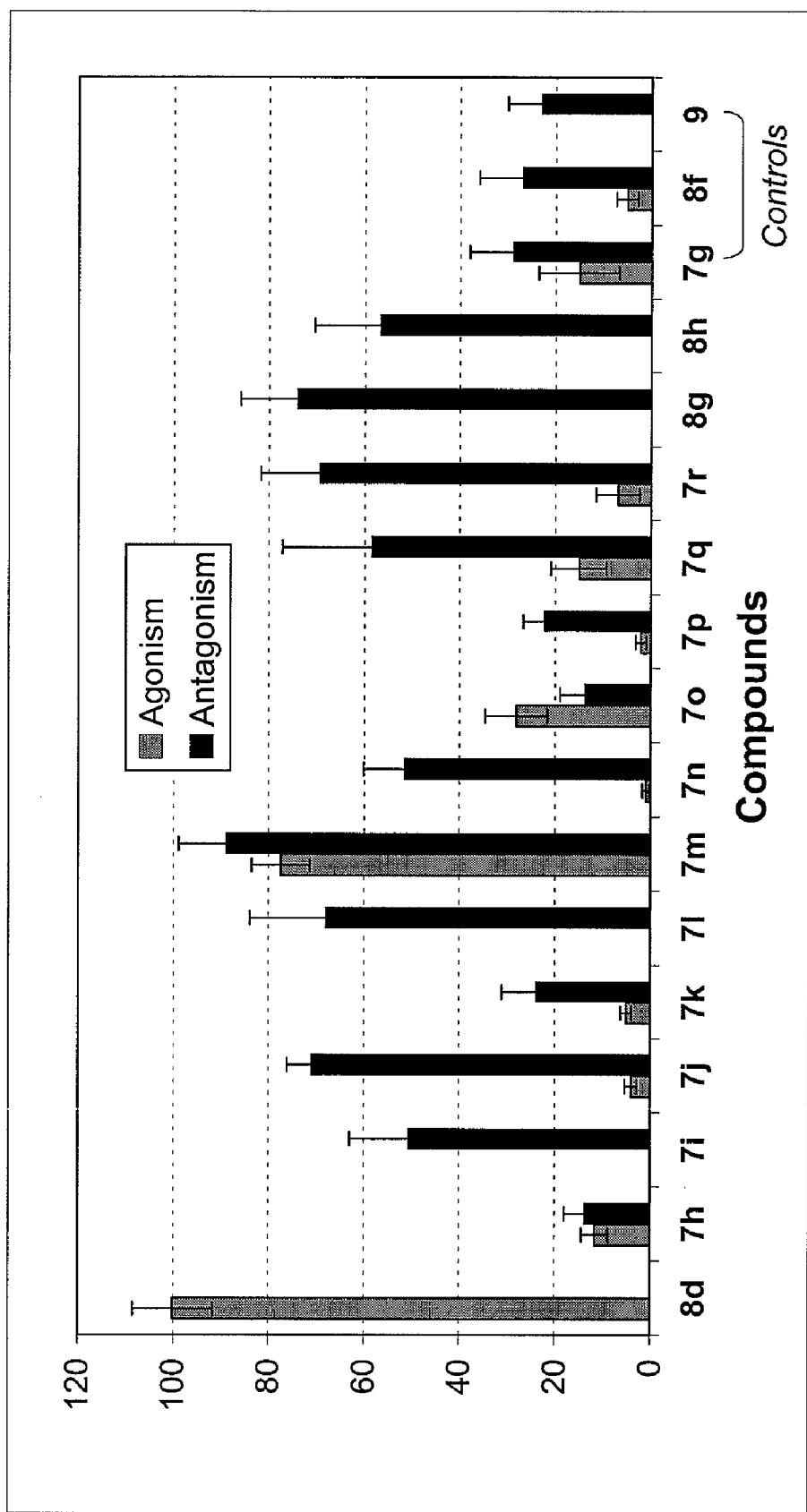


FIG. 32

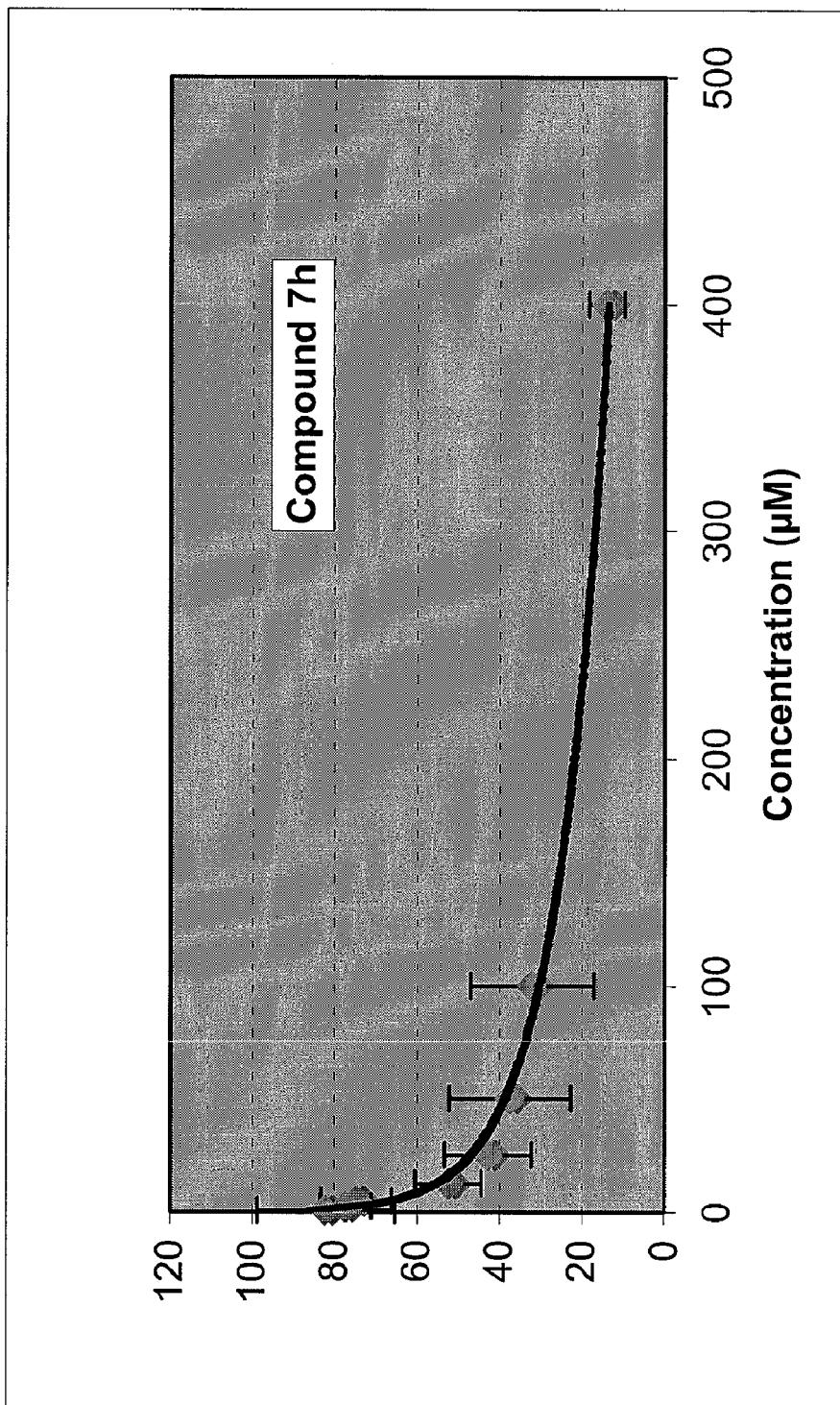


FIG. 33A

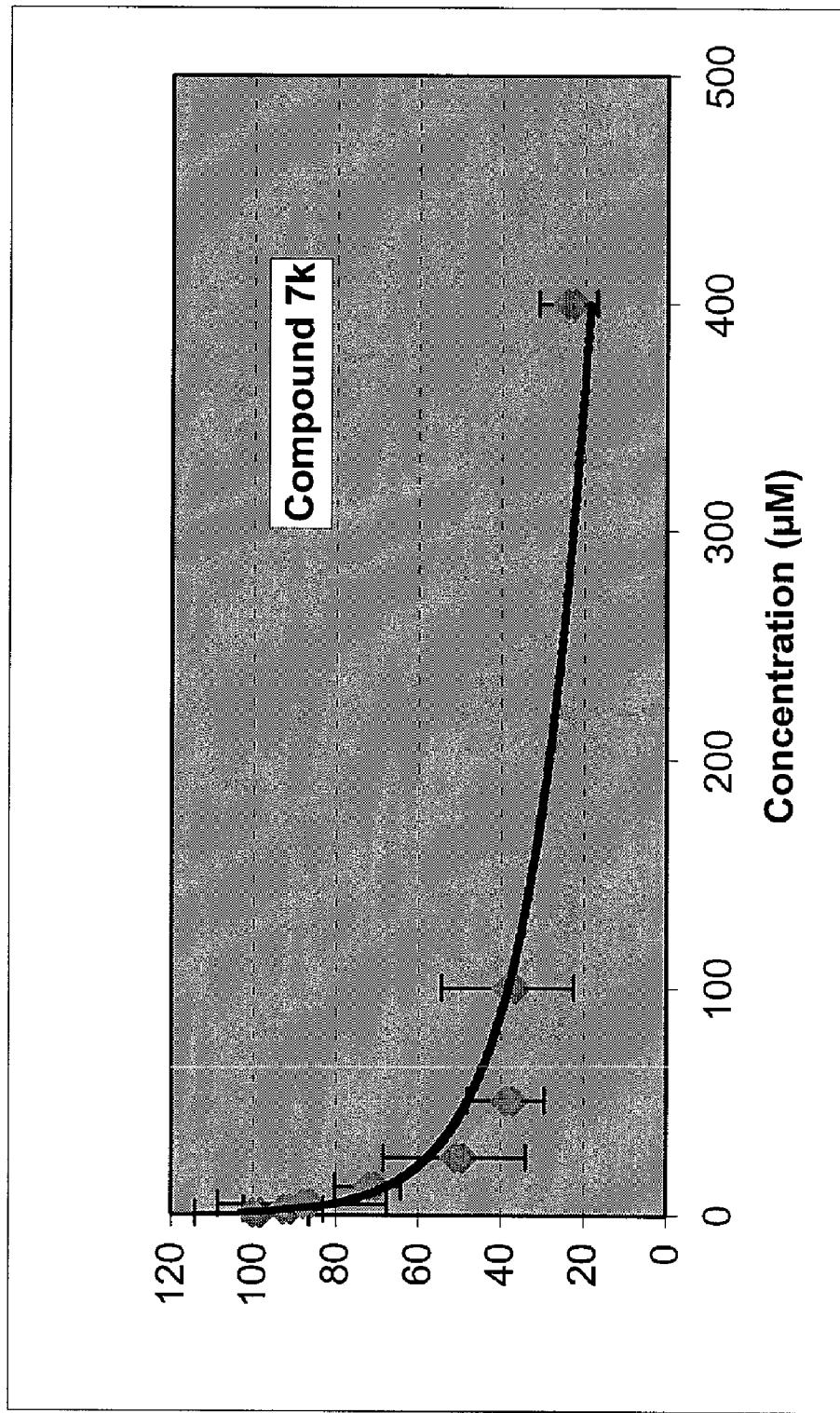


FIG. 33B

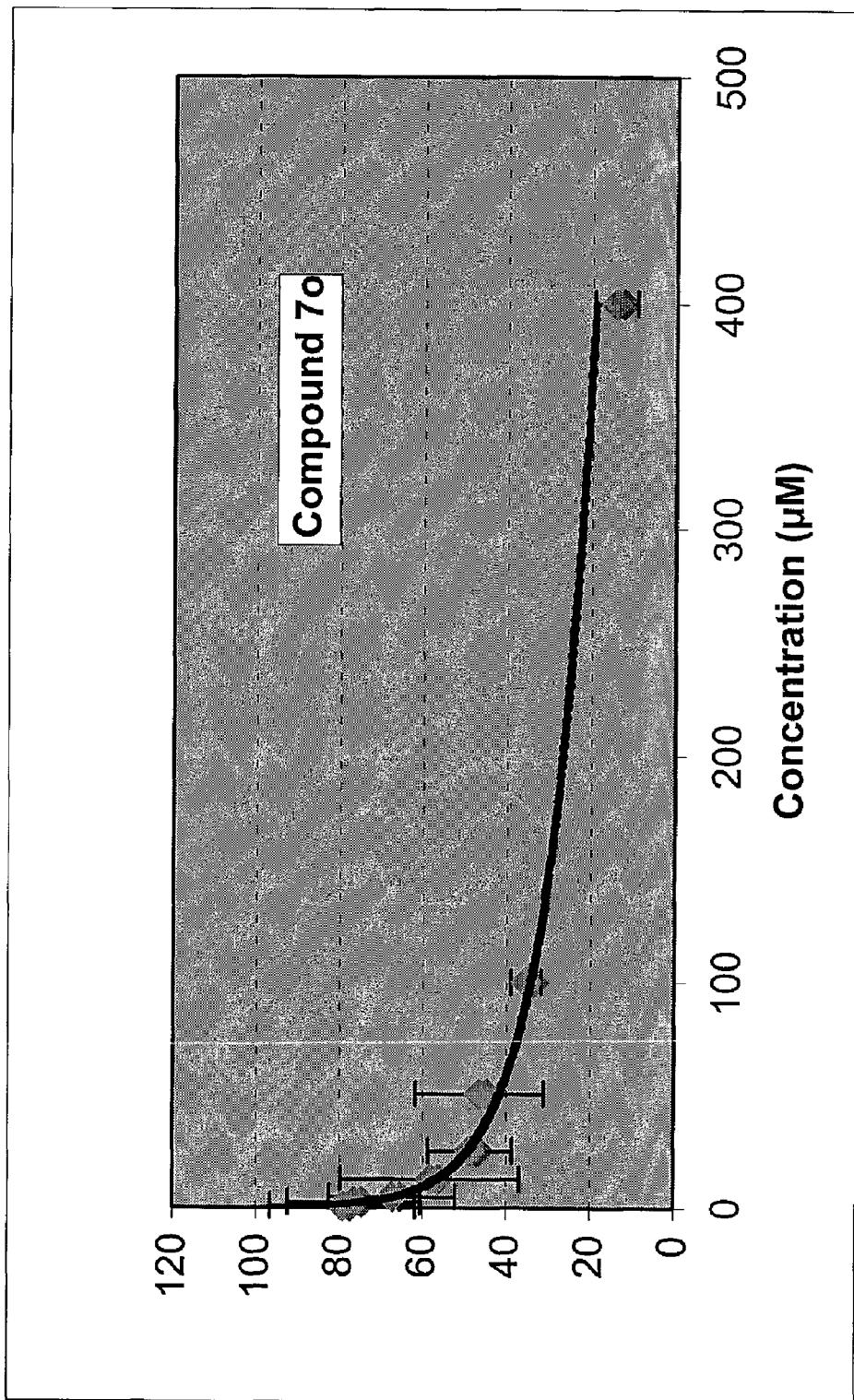


FIG. 33C

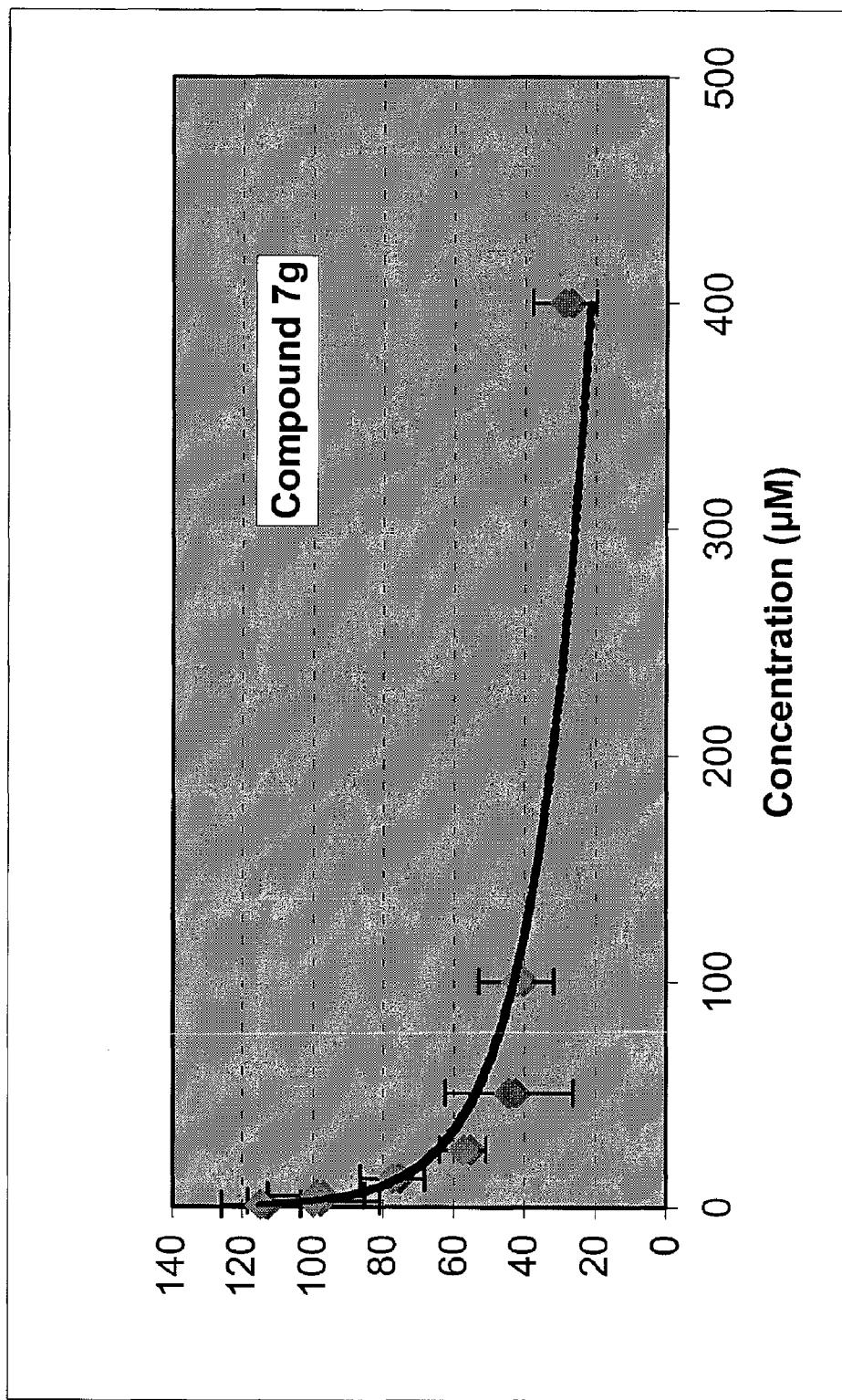


FIG. 33D

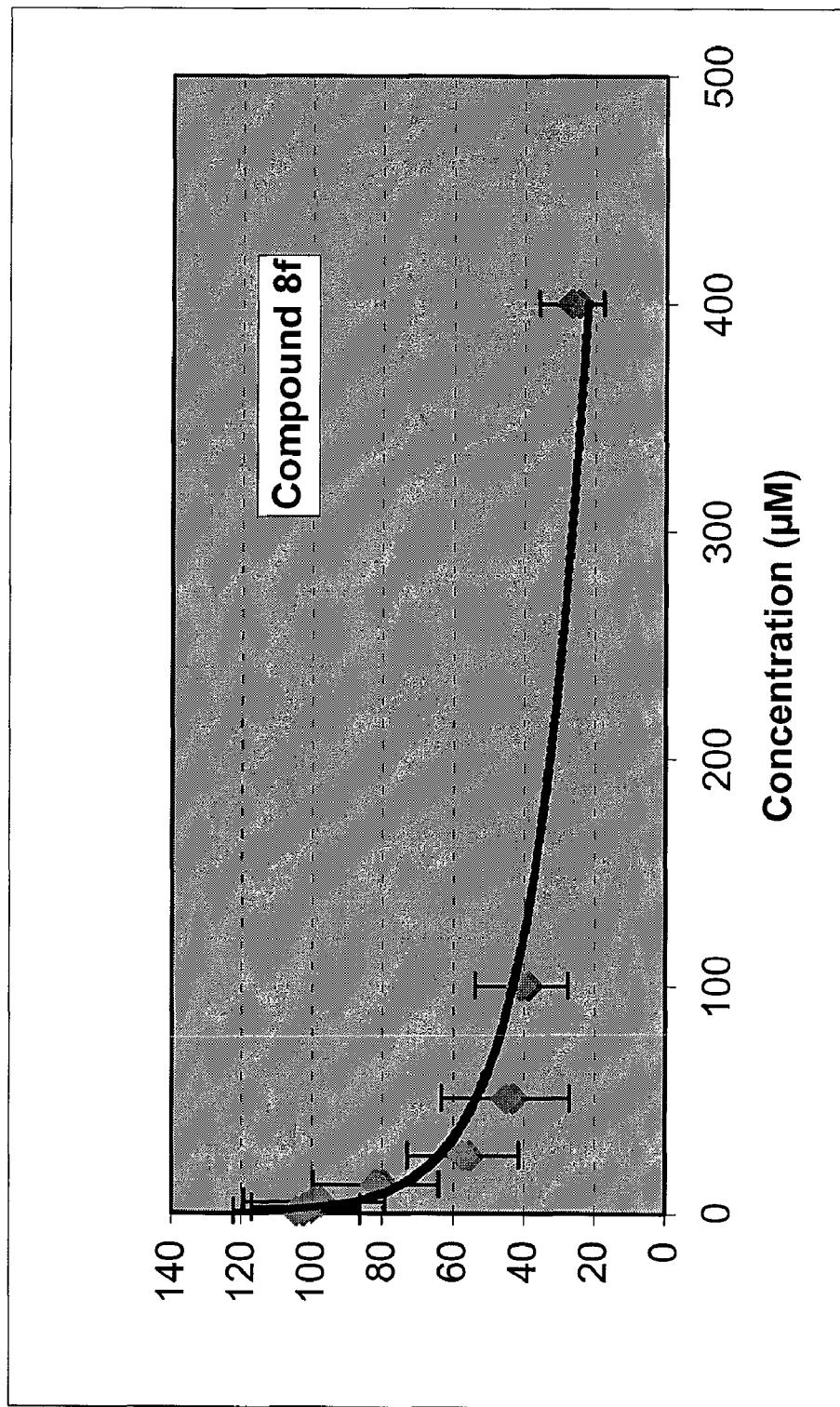


FIG. 33E

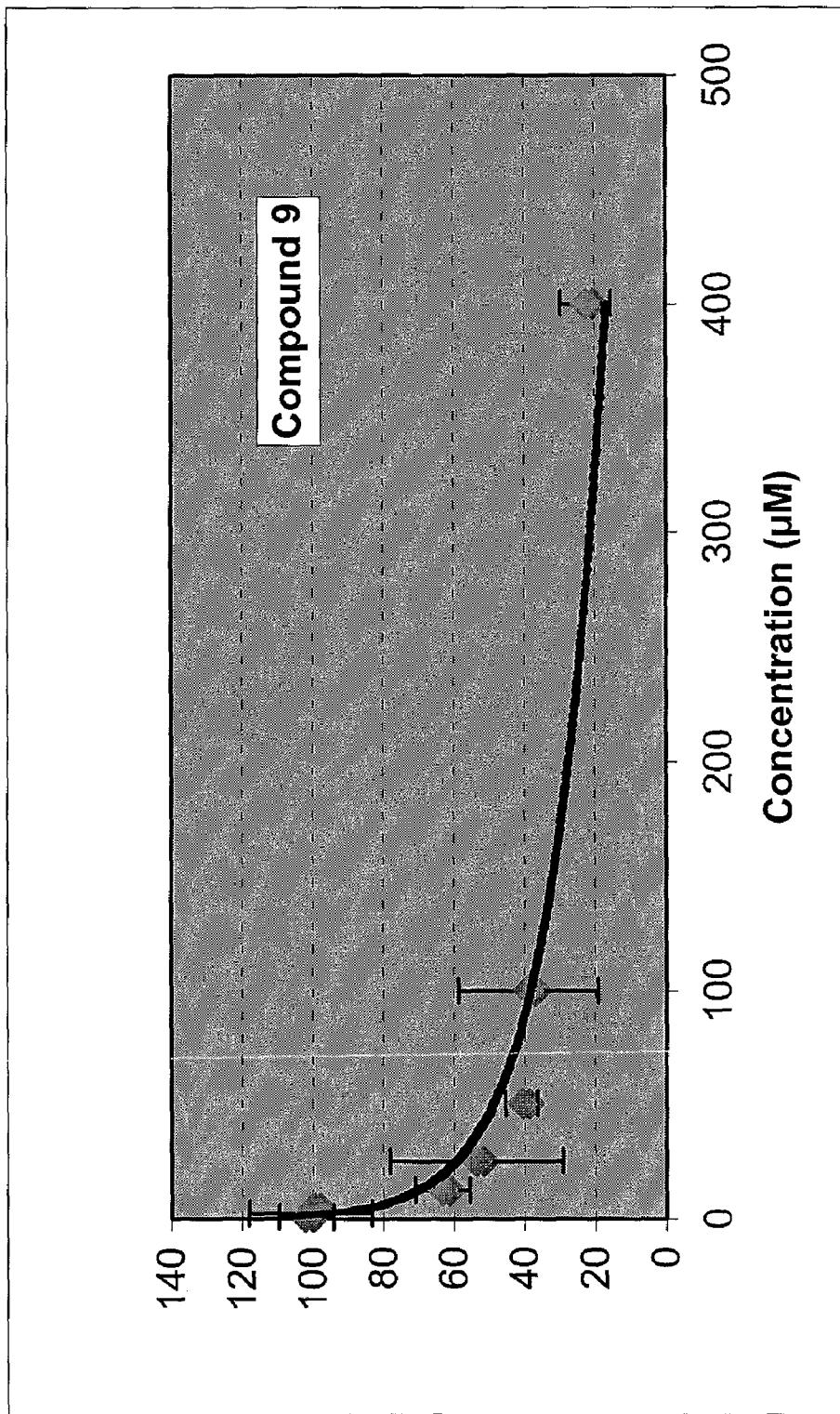


FIG. 33F

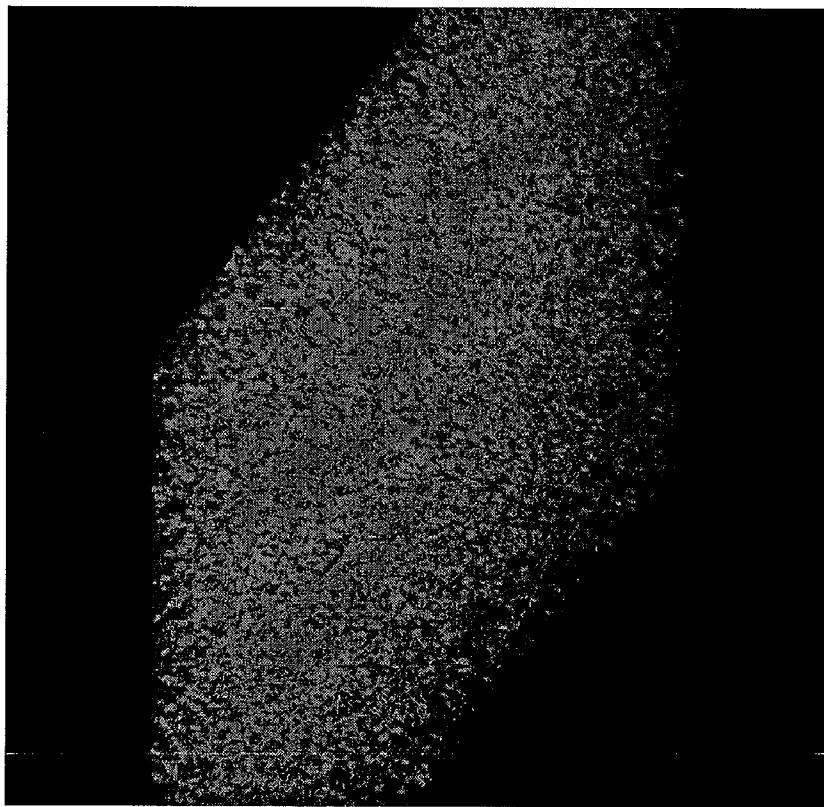


FIG. 34A

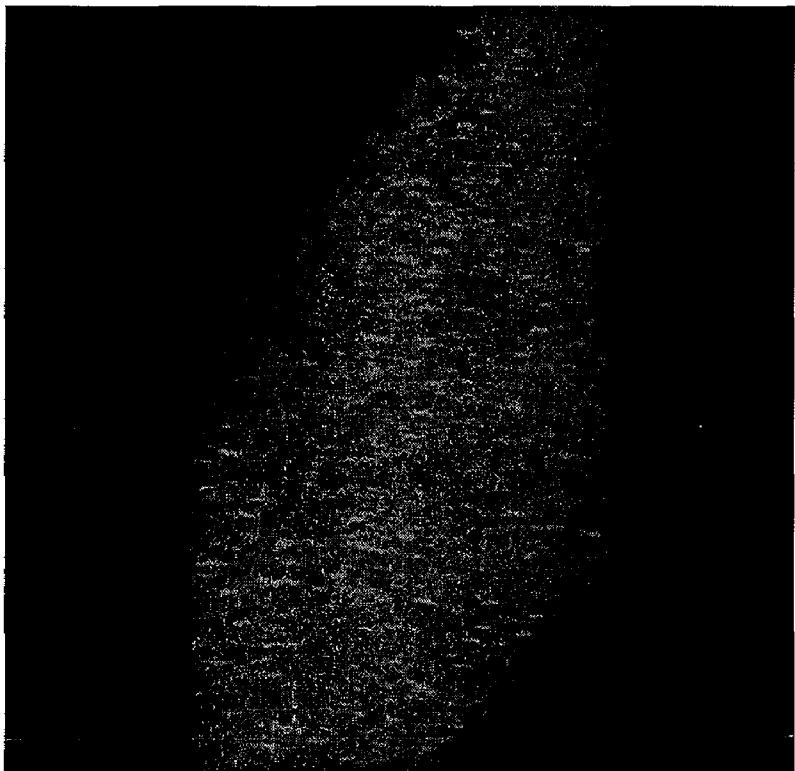


FIG. 34B

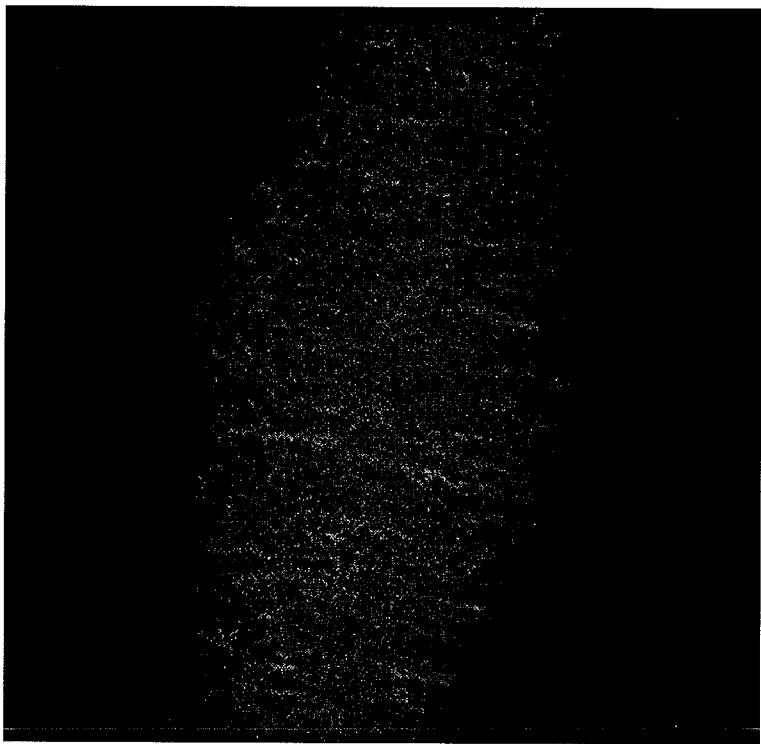


FIG. 34C

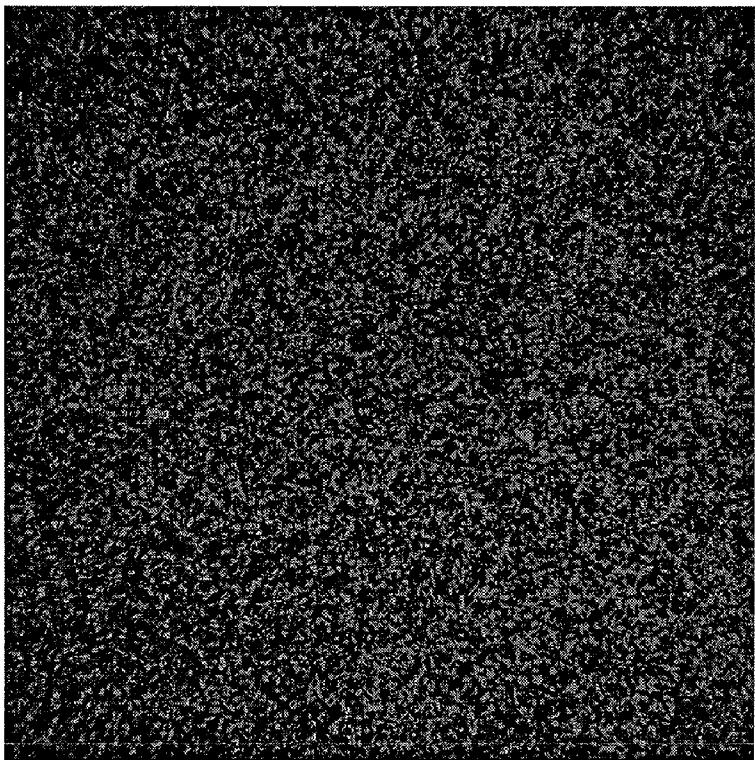


FIG. 35A

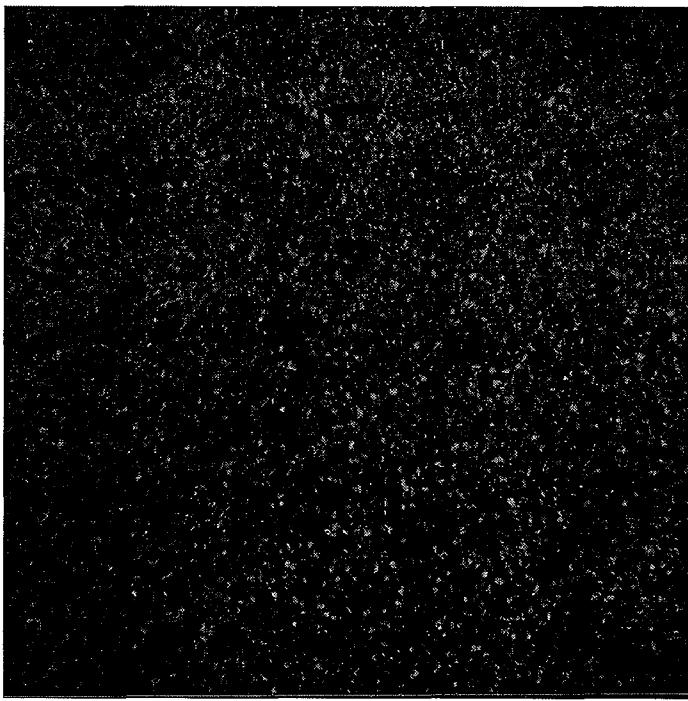


FIG. 35B

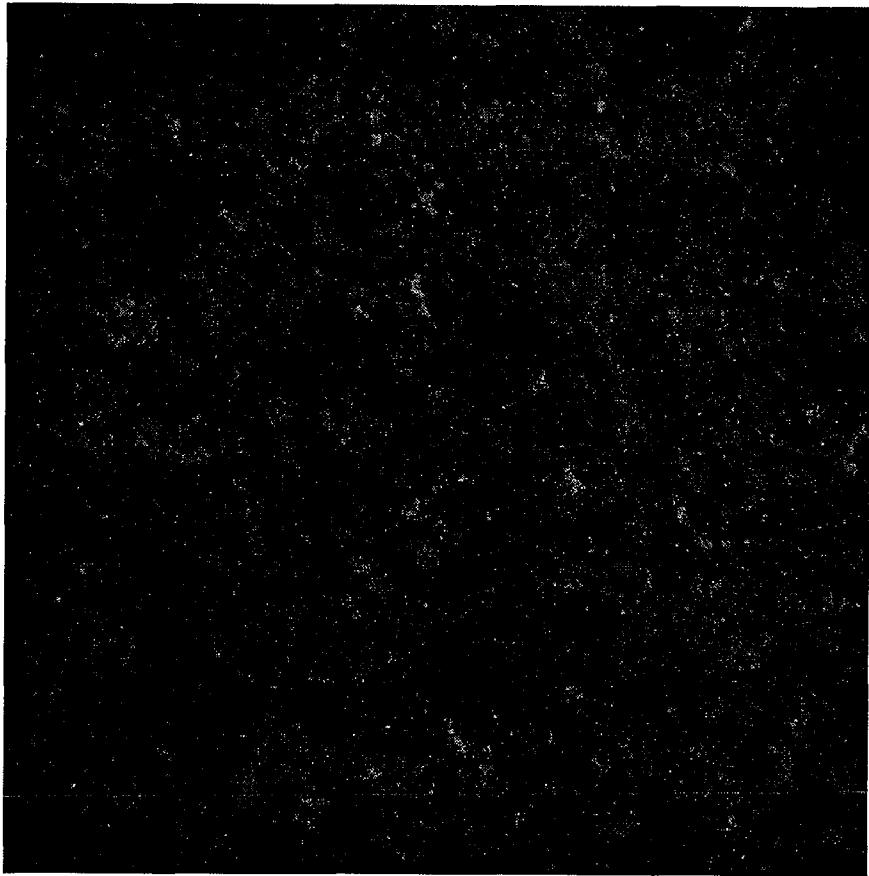


FIG. 35C

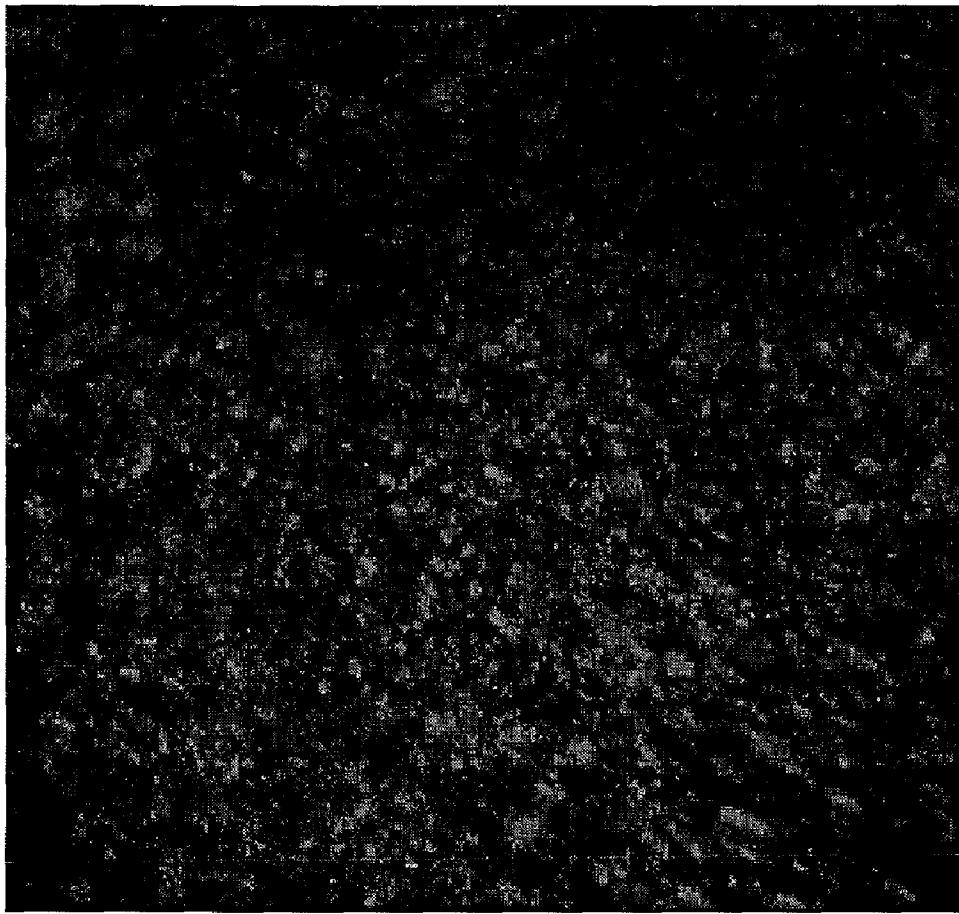


FIG. 36A

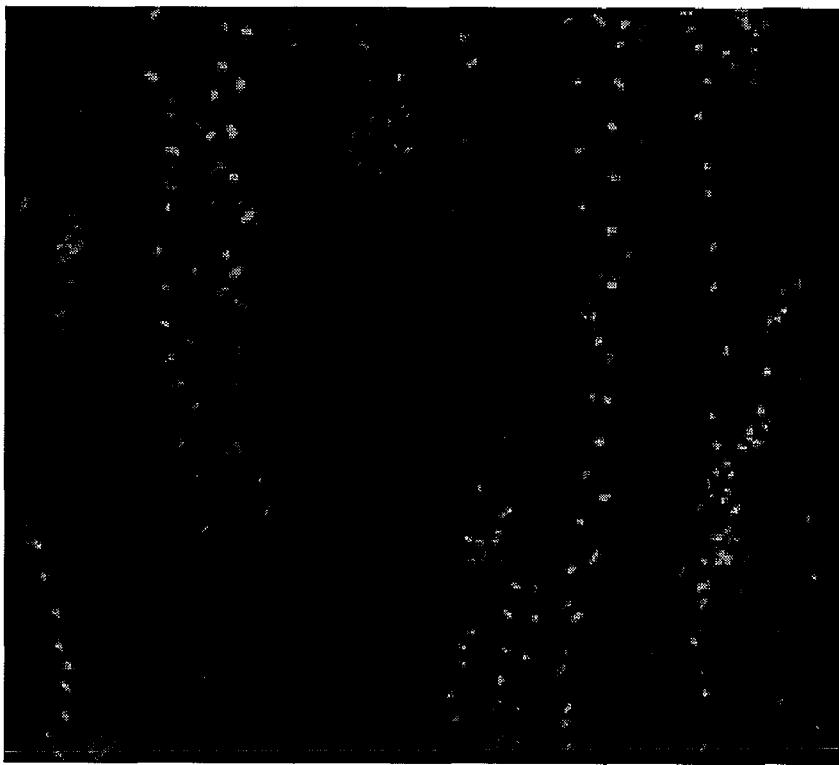


FIG. 36B

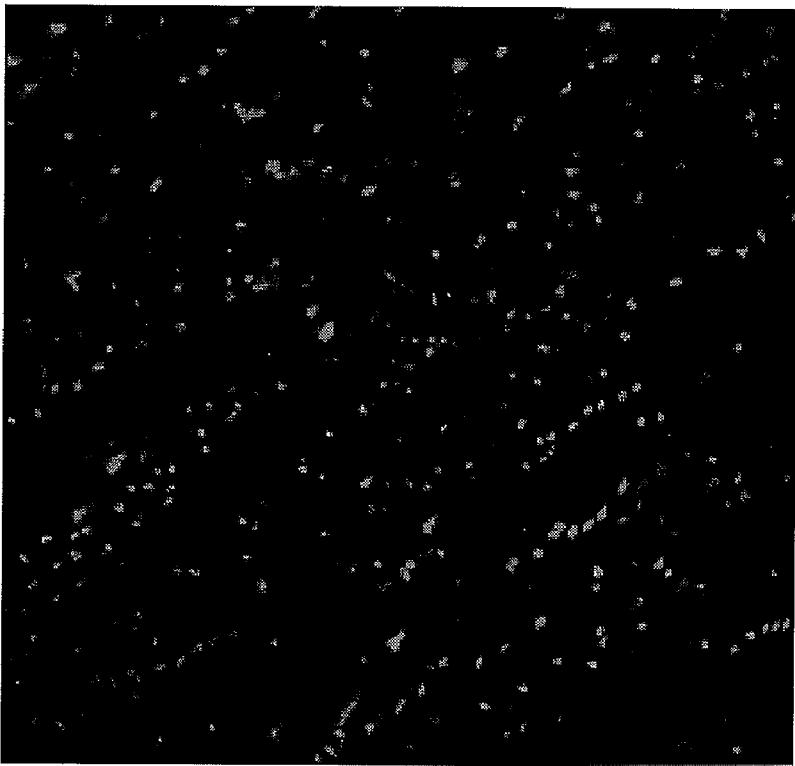


FIG. 36C

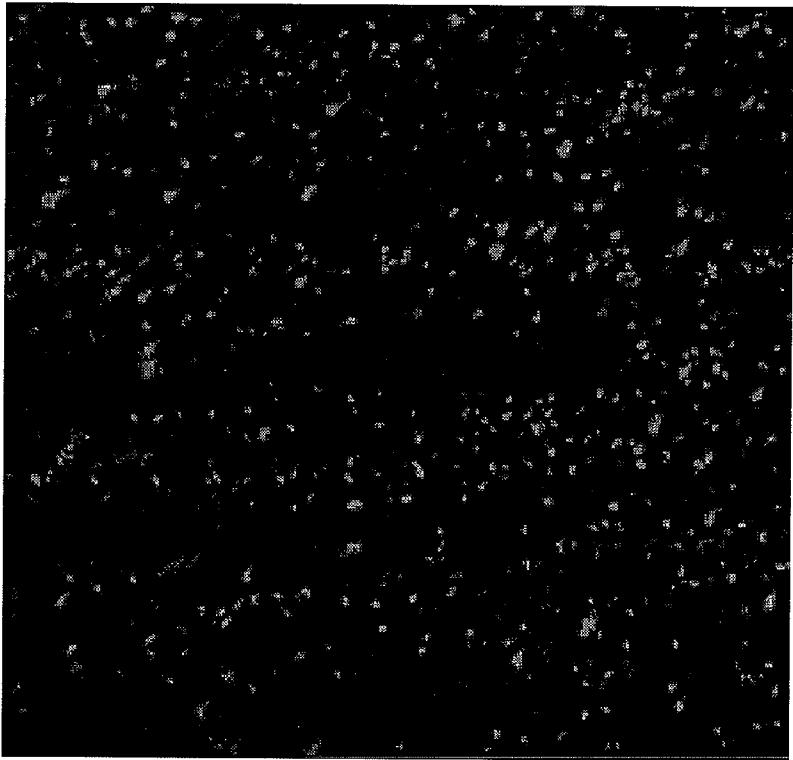


FIG. 36D

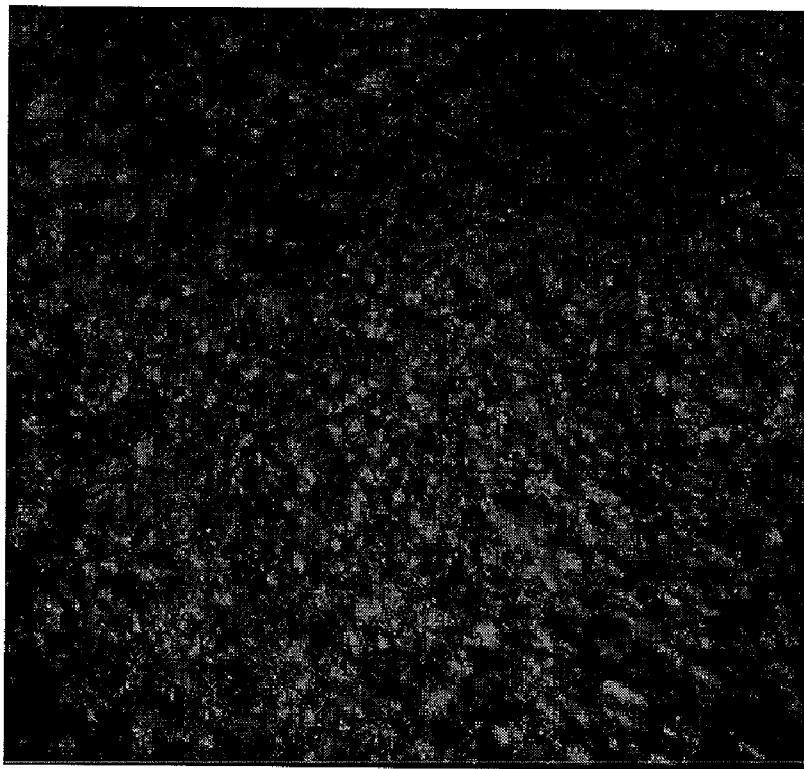


FIG. 37A

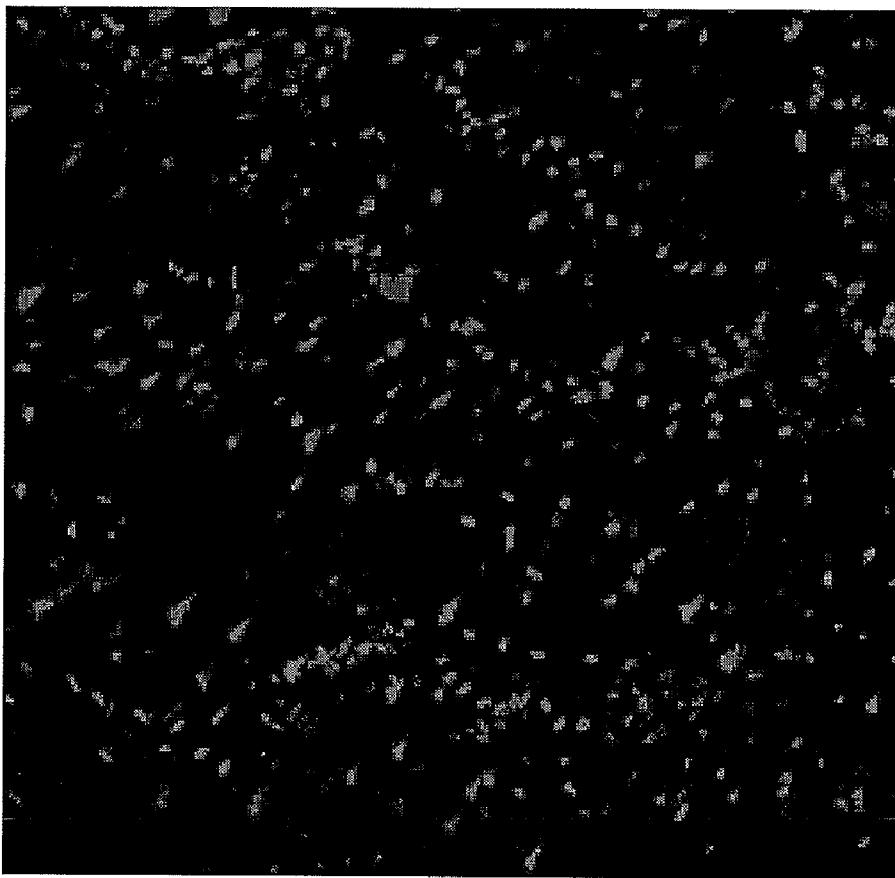


FIG. 37B

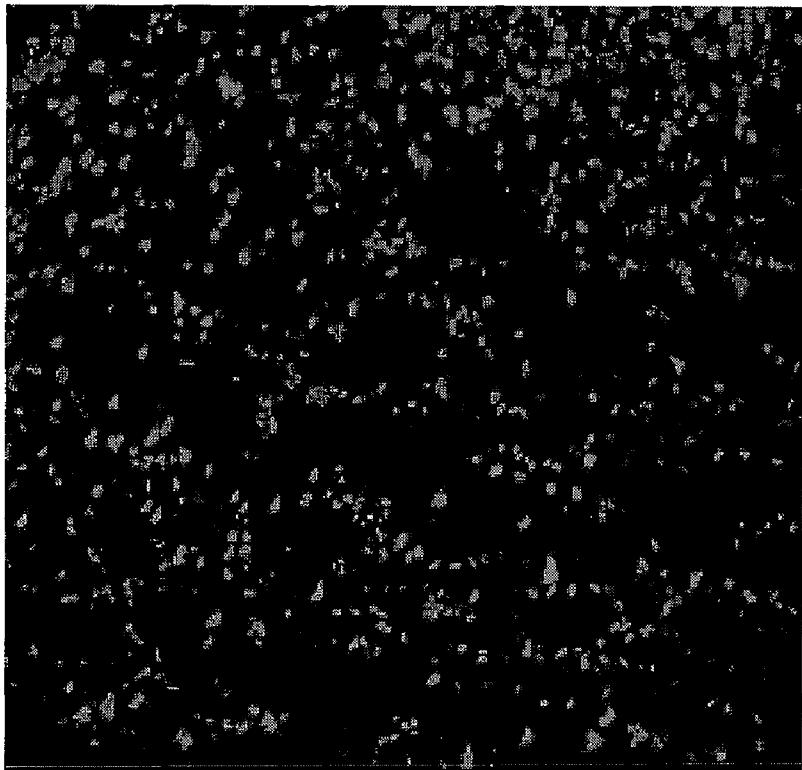


FIG. 37C

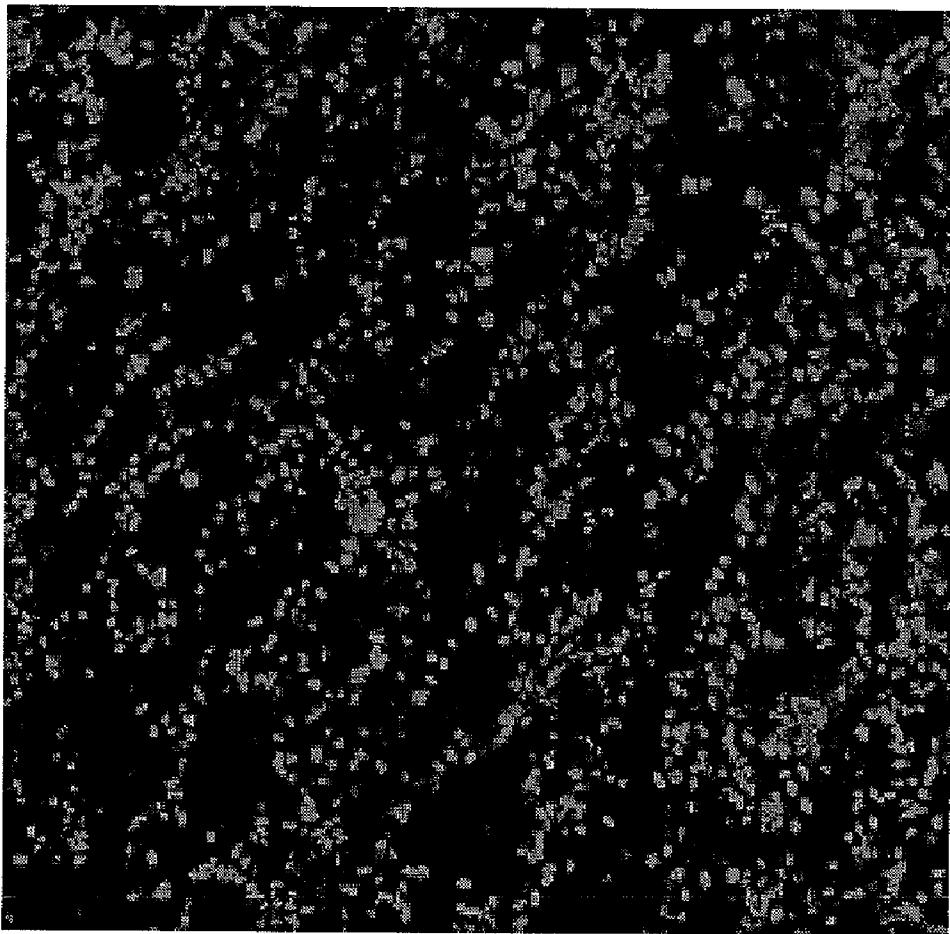


FIG. 37D

1

**COMPOUNDS AND METHODS FOR
MODULATING COMMUNICATION AND
VIRULENCE IN QUORUM SENSING
BACTERIA**

**CROSS-REFERENCE TO RELATED
APPLICATIONS**

This application is a continuation of U.S. application Ser. No. 11/275,896, filed Feb. 2, 2006 now allowed which claims the benefit of U.S. provisional application 60/593,681, filed Feb. 4, 2005 and U.S. provisional application 60/710,620, filed Aug. 23, 2005, all of which are incorporated by reference herein for all purposes.

FIELD OF THE INVENTION

This invention relates generally to molecules and methods for modulating quorum sensing in bacteria.

BACKGROUND OF THE INVENTION

Many microbial pathogens cause tremendous damage worldwide, in humans as well as in animals and crop plants. The continuing emergence of multiple-drug-resistant pathogen strains has necessitated finding new compounds that can be used in antimicrobial treatment. In general, two strategies exist for controlling pathogens, either kill the pathogen or attenuate its virulence such that it does not damage the host.

The strategy of attenuating bacterial virulence has the advantage of not creating selective pressure in favor of drug resistant strains. Antimicrobial compounds having virulence-attenuating but not cell-killing effects are expected to remain effective for longer periods of time than conventional antibiotics because of the lack of development of drug resistance. This approach has, however, suffered from a lack of specific targets for rational drug design.

Many bacteria use autoinducer ligands to monitor their population densities in a phenomenon called quorum sensing. At high cell densities, bacteria use this chemical signaling process to switch from a nomadic existence to that of multicellular community. This lifestyle switch is significant, as numerous pathogenic bacteria use quorum sensing to turn on virulence pathways and form drug-impervious communities called biofilms that are the basis of myriad chronic infections. Over 80% of bacterial infections in humans involve the formation of biofilms, as exemplified in lung infections by *Pseudomonas aeruginosa*, which is the primary cause of morbidity in cystic fibrosis patients. The treatment of infections by pathogens that form biofilms costs over \$1 billion/year in the US alone.

The control of gene expression in response to cell density was first described in the marine luminous bacteria *Vibrio fischeri* and *Vibrio harveyi*. Quorum sensing bacteria synthesize, release, and respond to specific acyl-homoserine lactone ("AHL" or "HSL") signaling molecules called autoinducers ("AI") to control gene expression as a function of cell density. The classical quorum-sensing pathway comprises at least three components: a membrane associated receptor/transcription factor; a diffusible signal, the autoinducer; and a recognition site in the promoter region of the target gene. The autoinducer binds to the receptor causing the receptor/AI complex to be internalized. This, in turn, allows the receptor or receptor/AI complex to bind to the promoter region of the target gene or genes altering transcription and down-regulat-

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ing or up-regulating gene expression. In most cases, this includes increased AI expression, thereby resulting in a cascade effect.

In recent years it has become apparent that many Gram-negative bacteria employ one or more quorum sensing systems. The quorum-sensing system is an attractive antibacterial target because it is not found in humans and is critical for high level bacterial virulence. Bacterial quorum sensing systems comprise AHL derivatives with different acyl side chains to regulate, in a cell-density dependent manner, a wide variety of physiological processes unique to the life-cycle of each microbe. These processes include: swarming, motility, biofilm formation, conjugation, bioluminescence and/or production of pigments, antibiotics and enzymes. For example, in *P. aeruginosa* quorum sensing pathways affect the expression of various exoenzymes, biofilm formation and cell-cell spacing. Other bacteria react to quorum sensing stimulation by expressing proteases and pectinases, expressing pili, entering stationary phase, emerging from lag phase and initiating cell division.

Biofilms are dense extracellular polymeric matrices in which the bacteria embed themselves. Biofilms allow bacteria to create a microenvironment that attaches the bacteria to the host surface and contains excreted enzymes and other factors allowing the bacteria to evade host immune responses including antibodies and cellular immune responses. Such biofilms can also exclude antibiotics. Further, biofilms can be extremely resistant to removal and disinfectant. For individuals suffering from cystic fibrosis, the formation of biofilms by *P. aeruginosa* is eventually fatal. Other bacteria also respond to quorum sensing signals by producing biofilms. Biofilms are inherent in dental plaques, and are found on surgical instruments, food processing and agriculture equipment and water treatment and power generating machinery and equipment.

Because of the virulence factors it triggers, the bacterial quorum-sensing system offers a novel target for use in modulating the virulence of pathogenic bacteria. All acyl-homoserine lactone quorum-sensing systems described to date, except that of *V. harveyi*, utilize AI synthases encoded by a gene homologous to luxI of *V. fischeri*. The response to the autoinducer is mediated by a transcriptional activator protein encoded by a gene homologous to luxR of *V. fischeri* (Bassler and Silverman, in Two Component Signal Transduction, Hoch et al., eds., Am. Soc. Microbiol. Washington D.C., pp. 431-435, 1995). Thus, the AHL quorum sensing system is present in a broad spectrum of pathogenic bacteria.

Gram-negative bacteria represent numerous relevant pathogens using quorum-sensing pathways. Besides *P. aeruginosa*, other quorum sensing bacteria include: *Aeromonas hydrophila*, *A. salmonicida*, *Agrobacterium tumefaciens*, *Burkholderia cepacia*, *Chromobacterium violaceum*, *Enterobacter agglomerans*, *Erwinia carotovora*, *E. chrysanthemi*, *Escherichia coli*, *Nitrosomas europaea*, *Obesumbacterium proteus*, *Pantoea stewartii*, *Pseudomonas aureofaciens*, *P. syringae*, *Ralstonia solanacearum*, *Rhizobium etli*, *R. leguminosarum*, *Rhodobacter sphaeroides*, *Serratia liquefaciens*, *S. marcescens*, *Vibrio anguillarum*, *V. fischeri*, *V. cholerae*, *Xenorhabdus nematophilus*, *Yersinia enterocolitica*, *Y. pestis*, *Y. pseudotuberculosis*, *Y. mediterranea*, and *Y. ruckeri*. Studies on the above listed bacteria indicate that, while the AI is generally an AHL compound, the genes affected as well as the phenotypes resulting from induction of the promoter differs according to the particular life cycle of each bacterium. Further, quorum sensing stimulation typically results in altered expression of multiple genes.

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In addition to affecting multiple genes, some bacteria have multiple stages of quorum sensing response. In these bacteria, the different stages of quorum sensing may be induced by different ligand/receptor pairs and result the expression of different sets of genes with similarly distinct phenotypes. For example, *V. harveyi* has two independent density sensing systems (Signaling Systems 1 and 2), and each is composed of a sensor-autoinducer pair. Signaling System 1 is composed of Sensor 1 and autoinducer 1 (AI-1), which is N43-hydroxybutanoyl-L-homoserine lactone (see Bassler et al., Mol. Microbiol. 9: 773-786, 1993). Signaling System 2 is composed of Sensor 2 and autoinducer 2 (AI-2) (Bassler et al., Mol. Microbiol. 13: 273-286, 1994). The structure of AI-2 heretofore has not been determined, nor have the gene(s) involved in biosynthesis of AI-2 been identified. Signaling System 1 is a highly specific system proposed to be used for intra-species communication and Signaling System 2 appears to be less species-selective, and is hypothesized to be for inter-species communication (Bassler et al., J. Bacteriol. 179: 4043-4045, 1997). Other research indicates that *V. cholerae* also has two stages of quorum-sensing response. The first, limits biofilm production, so that the microbe can escape the biofilm once it has passed through harsh environments such as hosts stomach. The second stage initiates swarming once the bacterium have escaped the biofilm and multiplied in the gut; allowing the bacteria to leave the host and start the cycle again.

Because of the diversity of quorum sensing ligands and phenotypes, having a large number of quorum sensing compounds with which to probe diverse quorum sensing responses will allow clinicians to identify ways to modulate or attenuate such responses. Further, if synthetic quorum sensing analogs were used, a greater diversity of responses may be identified other than those resulting from the native ligand. In addition, developing a synthetic route to quorum sensing compounds would provide a quick, more efficient way of producing analogs that would not rely on time-consuming techniques of molecular biology and would not per se be based on backbone of a native ligand. In addition, this strategy of attacking pathogenic bacteria via their quorum sensing pathways provides methods of controlling bacterial virulence without resorting to antibiotics. This will allow treatment of bacterial infections without inducing antibiotic resistance and the concomitant breeding of "superbugs".

Recent studies *in vivo* have shown that the virulence of *P. aeruginosa* lacking one or more genes responsible for quorum sensing is attenuated in its ability to colonize and spread within the host. Similarly, elimination of the AHL synthase in several plant pathogenic bacteria has led to complete loss of infectivity (Beck von Bodman, 1998, Proc. Natl. Acad. Sci. USA 95:7687-7692; Whitehead et al., 2001, Microbiol. Rev. 25:365-404). Transgenic plant systems engineered to express of AHL synthases ectopically, to produce inducing levels of AHLs, have shifted the balance of host-microbe interactions in favor of disease resistance (Fray et al., 1999, Nat. Biotechnol. 17:1017-1020; Mae et al., 2001, Mol. Plant Microbe Interact. 14:1035-1042). It is thought that the production of endogenous AHL compounds by plants is the basis of varying degrees of disease resistance and susceptibility (Teplitski et al., 2000, Mol. Plant-Microbe Interact. 13:637-648). The halogenated furanones produced by some marine algae are known to have a pronounced effect on suppressing marine biofouling. Some furanones have also been shown to effect on *V. cholerae* by eliminating its ability to express genes associated with their virulence phase.

The current understanding is that, at some threshold AHL concentration (and related cell density), the AHL ligand (AI)

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will bind its cognate receptor, a LuxR-type protein, and activate the transcription of target genes involved in group behavior. Fuqua, C.; Greenberg, E. P. *Nat. Rev. Mol. Cell Biol.* 2002, 3, 685-695. Blocking the binding of the endogenous AHL to its receptor with a non-native AHL is an attractive strategy for quorum sensing control.

In addition to their pathogenic costs, quorum sensing bacteria also have significant economic impact in industries other than health care. For example, in agriculture, various species of the genera *Rhizobium*, *Bradyrhizobium* and *Sinorhizobium* are important plant symbionts helping legumes to fix nitrogen, while, species of the genera *Erwinia*, *Xanthomonas* and *Pseudomonas* are responsible for significant food-spoilage of. Other industries, such as power generation, paper making and water treatment are subject to biofouling by many types of slime forming bacteria such as *Deinococcus geothermalis*.

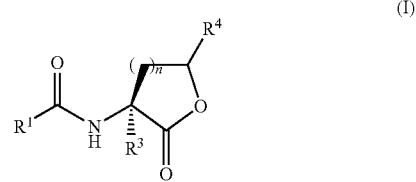
Nevertheless, the pace of AHL analog discovery has been slow as the majority of AHLs synthesized to date have been generated in poor yields and low purities and screened on an ad hoc basis (Eberhard, A.; Schineller, J. B. *Methods Enzymol.* 2000, 305, 301-315; Reverchon, S.; Chantegrel, B.; Deshayes, C.; Douthieu, A.; Cotte-Pattat, N. *Bioorg. Med. Chem. Lett.* 2002, 12, 1153-1157; Zhu, J.; Beaber, J. W.; More, M. I.; Fuqua, C.; Eberhard, A.; Winans, S. C. *J. Bacteriol.* 1998, 180, 5398-5405). Currently there are no antibacterial compounds that target the bacterial quorum sensing system to reduce bacterial virulence and increase susceptibility to bactericidal antibiotics. Therefore, new synthetic approaches are required for the generation of AHL analogs and the systematic evaluation of the effects of AHL ligand structure on quorum sensing. In addition, non-native AHL-analogs may provide significant benefits in their ability to stimulate quorum pathways without resulting increased virulence and pathogenicity.

SUMMARY OF THE INVENTION

The invention disclosed herein provides newly identified novel compounds that are autoinducer analogs. In addition, the novel compounds disclosed may act as either antagonists or agonists in various quorum sensing pathways. In various embodiments the invention also provides novel methods of synthesis to produce autoinducer analogs that include both natural and non-natural AHL analogs. Various AHL analogs produced as described herein can be used to modulate and/or regulate the infectivity and pathogenicity of quorum sensing bacteria and allow treatment of bacterial infections or blooms without resort to or in addition to antibiotics.

In various exemplary embodiments this invention discloses novel quorum sensing AHL analogs. In these exemplary embodiments the novel quorum sensing analogs disclosed herein are both AHL agonists and antagonists.

In various other exemplary embodiments the AHL analogs disclosed herein can be synthesized in high purity and with excellent yields.



In various exemplary embodiments, the present invention provides compounds of Formula I where n is 1, 2 or 3, wherein if n=1, the ring C bound to the substituted amino can be a chiral center, R¹ is selected from —H, —(CH₂)_aCH₃, —(CH₂)_aCOR₂, —(CH₂)_aCHOHR², —(CH₂)_aR⁶, —O—(CH₂)_aCH₃, —(CH₂)_aHC=CH, —HC=CH(CH₂)_aCH₃, —(CH₂)_aHC=CH(CH₂)_bCH₃, —R⁶HC=CHR⁷, —R⁶C=CR⁷, substituted and unsubstituted C₃-C₈ cycloalkyl, substituted and unsubstituted C₃-C₈ aryl, three to eight member substituted and unsubstituted heterocyclic rings, where the heteroatom is at least one of O, S or N, R² is selected from —H, —(CH₂)_aCH₃, substituted and unsubstituted C₃-C₈ cycloalkyl, substituted and unsubstituted C₃-C₈

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aryl, three to eight member substituted and unsubstituted heterocyclic rings, where the heteroatom is at least one of O, S or N, R³ is selected from —H, —CH₂CH₃, —CH₃, R⁴ is selected from —H, —CH₂—, R⁵ is —H, R⁶ and R⁷ are the same or different, selected from —H, substituted and unsubstituted C₃-C₈ cycloalkyl, substituted and unsubstituted C₃-C₈ aryl, three to eight member substituted and unsubstituted heterocyclic rings, where the heteroatom is at least one of O, S or N, and a and b are independently integers from 0 to 15.

In certain preferred embodiments, compounds are selected from the compounds of Table 1.

TABLE 1

Compounds of Formula I

Cmpd	R ¹	R ²	R ³	R ⁴	R ⁵	n
7a	—(CH ₂) ₂ CH ₃	—	H	H	—	1
7b	—(CH ₂) ₄ CH ₃	—	H	H	—	1
7c	—(CH ₂) ₆ CH ₃	—	H	H	—	1
7d	—(CH ₂) ₈ CH ₃	—	H	H	—	1
7e	—(CH ₂) ₁₀ CH ₃	—	H	H	—	1
7f	—(CH ₂) ₁₂ CH ₃	—	H	H	—	1
8a	—(CH ₂)COR ²	—(CH ₂) ₂ CH ₃	H	H	—	1
8b	—(CH ₂)COR ²	—(CH ₂) ₄ CH ₃				
8c	—(CH ₂)COR ²	—(CH ₂) ₆ CH ₃				
8d	—(CH ₂)COR ²	—(CH ₂) ₈ CH ₃				
8e	—(CH ₂)COR ²	—(CH ₂) ₁₀ CH ₃				
	—(CH ₂) _a CHOHR ²					
	—(CH ₂) _a R ⁶					
	—O—(CH ₂) _a CH ₃					
	—(CH ₂) _a HC=CH					
	—HC=CH(CH ₂) _a CH ₃					
	—(CH ₂) _a HC=CH(CH ₂) _b CH ₃					
29 (L)	—(CH ₂) ₅ HC=CH(CH ₂) ₅ CH ₃	—	H	H	—	1
22 (L)	—(CH ₂) ₂ HC=CH ₂	—	H	H	—	1
	—R ⁶ HC=CHR ⁷	—				
	—R ⁶ C≡CR ⁷					
30 (L)	—(CH ₂) _a SH					
7h (L)		—	H	H	—	1
7i (D)						
7j (L)		—	H	H	—	1
7k (L)		—	H	H	—	1
7l (D)						

TABLE 1-continued

Compounds of Formula I						
	R ¹	R ²	R ³	R ⁴	R ⁵	n
7m (L)		—	H	H	—	1
7n (L)		—	H	H	—	1
		—				
A = H, F, Cl, Br, I						
7o (L)		—	H	H	—	1
7p (L)		—	H	H	—	1
7q (L)		—	H	H	—	1
7r (D)						
8f (L)	—(CH ₂)COR ²					
8g (D)		H	H	—		
14 (L)		—	H	H	—	1
50 (L)	—CH(CH ₃) ₂ CH ₂ CH ₃	—	—CH ₂ CH ₃	—CH ₂ —	I	1
51 (D)	—(CH ₂) ₂ C ₆ F ₆	—	H	H	—	1

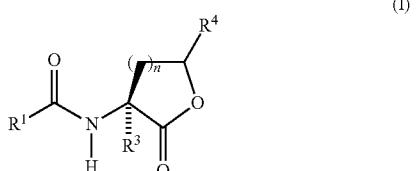
TABLE 1-continued

Compounds of Formula I						
Cmpd	R ¹	R ²	R ³	R ⁴	R ⁵	n
53		—	H	—CH ₂ —		2
54 (L)	—(CH ₂) ₃ CH ₃	—	—CH ₃	—CH ₂ —	I	1
55	—(CH ₂)COR ²	—(CH ₂) ₂ CH ₃	H	—CH ₂ —	—N(CH ₃) ₂	2
56	—(CH ₂) ₂ HC=CH(CH ₂) ₃ CH ₃	—	—CH ₃	—CH ₂ —		2
57 (L)	—CH ₂ CH(CH ₃) ₂	—	H	—CH ₂ —	—N(CH ₂ HC=CH ₂) ₂	1
58 (L)		—	H	H	—	1
59 (L)	—CH ₂ (CF ₂) ₄ F ₃	—	H	H	—	1

In other preferred embodiments, the present invention provides combinatorial libraries of autoinducer analogs. In certain embodiments, the invention provides a combinatorial library of two or more compounds of Formula (I).

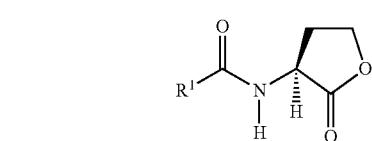
—CH₃, R⁴ is selected from —H, —CH₂—, R⁵ is —H, R⁶ and R⁷ are the same or different, selected from —H, substituted and unsubstituted C₃-C₈ cycloalkyl, substituted and unsubstituted C₃-C₈ aryl, three to eight member substituted and unsubstituted heterocyclic rings, where the heteroatom is at least one of O, S or N, and a and b are independently integers from 0 to 15.

In certain embodiments, the compound has the formula:

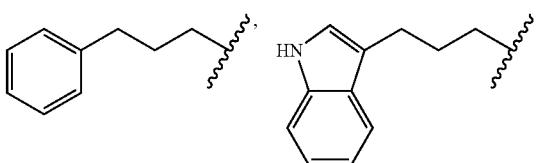


where n is 1, 2 or 3, wherein if n=1, the ring C bound to the substituted amino can be a chiral center, R¹ is selected from —H, —(CH₂)_aCH₃, —(CH₂)_aCOR², —(CH₂)_aCHOHR², —(CH₂)_aR⁶, —O—(CH₂)_aCH₃, —(CH₂)_aHC=CH, —HC=CH(CH₂)_aCH₃, —(CH₂)_aHC=CH(CH₂)_bCH₃, —R⁶HC=CHR⁷, —R⁶C≡CR⁷, substituted and unsubstituted C₃-C₈ cycloalkyl, substituted and unsubstituted C₃-C₈ aryl, three to eight member substituted and unsubstituted heterocyclic rings, where the heteroatom is at least one of O, S or N, R² is selected from —H, —(CH₂)_aCH₃, substituted and unsubstituted C₃-C₈ cycloalkyl, substituted and unsubstituted C₃-C₈ aryl, three to eight member substituted and unsubstituted heterocyclic rings, where the heteroatom is at least one of O, S or N, R³ is selected from —H, —CH₂CH₃,

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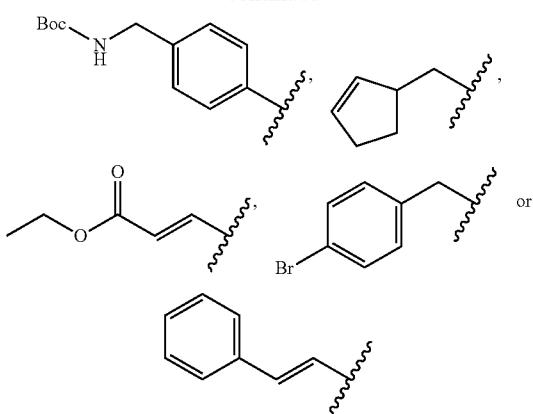


where R¹ is —CH₂COR², C₆H₁₃,

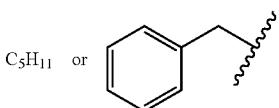


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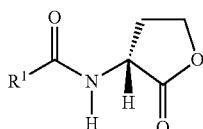
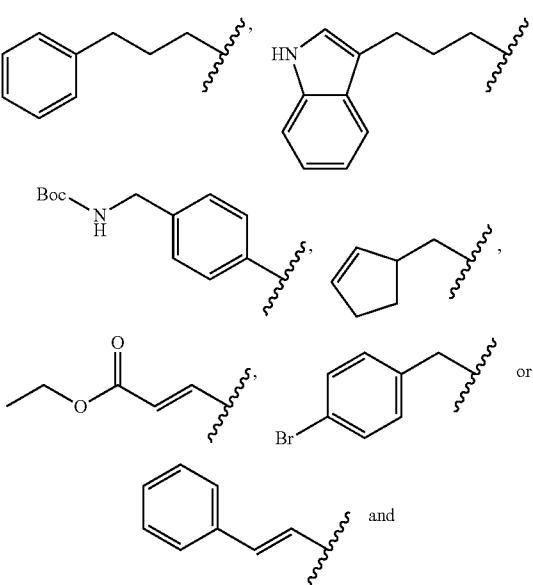
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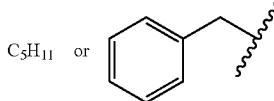
and

R² is

In other embodiments, the compound has the formula:

where R¹ is —CH₂COR² or**12**R² is

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10 In various exemplary embodiments, this invention separately provides methods for the synthesis of AHL analogs that are solid phase syntheses as described in Schemes I-III.

15 In some other exemplary embodiments, this invention provides methods for the efficient synthesis of AHL analogs that are produced with high purity and yield. In various exemplary embodiments, AHL analogs synthesized by the methods described herein are synthesized in a short period of time.

This invention separately provides a method for the synthesis of both naturally occurring and non-naturally occurring AHL analogs. In various other exemplary embodiments, the 20 invention provides methods for the efficient synthesis of AHL ligand libraries. In various exemplary embodiments, the AHL analog library is a combinatorial library. In still other exemplary embodiments, the AHL analogs produced by the methods disclosed herein can separately include both agonists and antagonists.

This invention separately provides methods for the efficient screening of AHL analogs. In various exemplary embodiments the screening methods described herein include reporter gene assays and biofilm production assays.

30 This invention separately provides compositions and methods for the manipulation and perturbation of the bacterial quorum-sensing pathway. In various exemplary embodiments AHL analogs described herein may compete with native AHL ligands inhibiting biofilm formation. In various 35 other exemplary embodiments the AHL prematurely stimulate biofilm production.

In various exemplary embodiments, the invention provides methods for inhibiting and/or attenuating the virulence of 40 quorum sensing bacteria by interfering with the quorum sensing pathway and reducing and/or inhibiting the production of biofilms, virulence factors and/or enzymes by quorum sensing bacteria.

In various other exemplary embodiments, this invention provides compounds and methods for use in agriculture to 45 inhibit/modulate and/or encourage the growth and infectivity bacteria.

In yet other exemplary embodiments, this invention separately provides compounds and methods for their use where biofouling has an economic impact such as in paper making, 50 water treatment and power generation.

Other objects, features and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples, while indicating exemplary embodiments of the invention, are given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

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BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1A is a schematic diagram illustrating Scheme I for the synthetic production of AHLs, where a=DIC, HOBT, 65 CHCl₃/DMF, microwave 50° C., 2×10 minutes; b=DMF, microwave 150° C., 7 minutes; c=CNBr, TFA, CHCl₃/H₂O, microwave 60° C., 30 minutes.

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FIG. 1B is a schematic diagram illustrating Scheme II for the synthetic production of quorum sensing compounds.

FIG. 1C is a schematic diagram illustrating Scheme III for the synthetic production of quorum sensing compounds.

FIG. 2A shows a GC-MS spectrum and FIG. 2B shows an NMR spectrum for N-butanoyl-L-homoserine lactone (7a) produced by the scheme shown in FIG. 1A.

FIG. 3A shows a GC-MS spectrum and FIG. 3B shows an NMR spectrum for N-hexanoyl-L-homoserine lactone (7b) produced by the scheme shown in FIG. 1A.

FIG. 4A shows a GC-MS spectrum and FIG. 4B shows an NMR spectrum for N-octanoyl-L-homoserine lactone (7c) produced by the scheme shown in FIG. 1A.

FIG. 5A shows a GC-MS spectrum and FIG. 5B shows an NMR spectrum for N-decanoyl-L-homoserine lactone (7d) produced by the scheme shown in FIG. 1A.

FIG. 6A shows a GC-MS spectrum and FIG. 6B shows an NMR spectrum for N-dodecanoyl-L-homoserine lactone (7e) produced by the scheme shown in FIG. 1A.

FIG. 7A shows a GC-MS spectrum and FIG. 7B shows an NMR spectrum for N-hexadecanoyl-L-homoserine lactone (7f) produced by the scheme shown in FIG. 1A.

FIG. 8A shows a GC-MS spectrum and FIG. 8B shows an NMR spectrum for N-heptanoyl-L-homoserine lactone (7g) produced by the scheme shown in FIG. 1A.

FIG. 9A shows a GC-MS spectrum and FIG. 9B shows an NMR spectrum for N-(indole-3-butanoyl)-L-homoserine lactone (7h) produced by the scheme shown in FIG. 1A.

FIG. 10A shows a GC-MS spectrum and FIG. 10B shows an NMR spectrum for N-(indole-3-butanoyl)-D-homoserine lactone (7i) produced by the scheme shown in FIG. 1A.

FIG. 11 shows an NMR spectrum for N-Boc-(4-aminomethyl)-N-benzoyl-L-homoserine lactone (7j) produced by the scheme shown in FIG. 1A.

FIG. 12A shows a GC-MS spectrum and FIG. 12B shows an NMR spectrum for N-(2-cyclopentene-1-acetanoyl)-L-homoserine lactone (7k) produced by the scheme shown in FIG. 1A.

FIG. 13A shows a GC-MS spectrum and FIG. 13B shows an NMR spectrum for N-(2-cyclopentene-1-acetanoyl)-D-homoserine lactone (7l) produced by the scheme shown in FIG. 1A.

FIG. 14 shows an NMR spectrum for N-Boc-aminocaproyl-L-homoserine lactone (7m) produced by the scheme shown in FIG. 1A.

FIG. 15A shows a GC-MS spectrum and FIG. 15B shows an NMR spectrum for N-monoethyl fumaroyl-L-homoserine lactone (7n) produced by the scheme shown in FIG. 1A.

FIG. 16A shows a GC-MS spectrum and FIG. 16B shows an NMR spectrum for N-(4-bromophenylacetanoyl)-L-homoserine lactone (7o) produced by the scheme shown in FIG. 1A.

FIG. 17A shows a GC-MS spectrum and FIG. 17B shows an NMR spectrum for N-(trans-cinamoyl)-L-homoserine lactone (7p) produced by the scheme shown in FIG. 1A.

FIG. 18A shows a GC-MS spectrum and FIG. 18B shows an NMR spectrum for N-(4-phenylbutanoyl)-L-homoserine lactone (7q) produced by the scheme shown in FIG. 1A.

FIG. 19A shows a GC-MS spectrum and FIG. 19B shows an NMR spectrum for N-(4-phenylbutanoyl)-D-homoserine lactone (7r) produced by the scheme shown in FIG. 1A.

FIG. 20 shows an NMR spectrum for N-(3-oxo-hexanoyl)-L-homoserine lactone (8a) produced by the scheme shown in FIG. 1A.

FIG. 21 shows an NMR spectrum for N-(3-oxo-octanoyl)-L-homoserine lactone (8b) produced by the scheme shown in FIG. 1A.

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FIG. 22 shows an NMR spectrum for N-(3-oxo-decanoyl)-L-homoserine lactone (8c) produced by the scheme shown in FIG. 1A.

FIG. 23 shows an NMR spectrum for N-(3-oxo-dodecanoyl)-L-homoserine lactone (8d) produced by the scheme shown in FIG. 1A.

FIG. 24 shows an NMR spectrum for N-(3-oxo-tetradecanoyl)-L-homoserine lactone (8e) produced by the scheme shown in FIG. 1A.

FIG. 25 shows an NMR spectrum for N-(3-oxo-4-phenylbutanoyl)-L-homoserine lactone (8f) produced by the scheme shown in FIG. 1A.

FIG. 26 shows an NMR spectrum for N-(3-oxo-4-phenylbutanoyl)-D-homoserine lactone (8g) produced by the scheme shown in FIG. 1A.

FIG. 27 shows an NMR spectrum for N-(3-oxo-octanoyl)-D-homoserine lactone (8h) produced by the scheme shown in FIG. 1A.

FIGS. 28A-B show an NMR spectrum for R-Mosher amide AHL derivatives (300 MHz, CDCl₃) produced by the scheme shown in FIG. 1A.

FIGS. 29A-29D show the results of antagonism screens for (FIG. 29A) quorum sensing antagonists identified by the methods of the present invention and (FIG. 29B) known quorum sensing antagonists and quorum sensing antagonism activity of both sets of compounds in (FIG. 29C) *A. tumefaciens* and (FIG. 29D) *P. aeruginosa* reporter strains.

FIG. 30 is a graphical display of the results of *A. tumefaciens* reporter gene screening using AHL derivatives 7h-7r and 8f-8h.

FIGS. 31A-31F are graphical displays of the results of dose response antagonism screening data for AHL derivatives 7h (FIG. 31A), 7k (FIG. 31B) and 7o (FIG. 31C) and control compounds 7g (FIG. 31D), 8f (FIG. 31E) and 9 (FIG. 31F) in an *A. tumefaciens* reporter strain. Reporter strain: WCF47 (pCF372). Miller units report relative beta-galactosidase activity with ONPG as colometric substrate. Compounds screened at various concentrations against 100 nM native ligand 8b (OOHL). Error bars are ± one S.E.M., calculated from at least three replicate screens.

FIG. 32 is a graphical display of the results of *P. aeruginosa* reporter gene screening using AHL derivatives 7h-7r and 8f-8h.

FIGS. 33A-33F are graphical displays of the results of dose response antagonism screening data for AHL derivatives 7h (FIG. 33A), 7k (FIG. 33B) and 7o (FIG. 33C) and control compounds 7g (FIG. 33D), 8f (FIG. 33E) and 9 (FIG. 33F) in a *P. aeruginosa* reporter strain. Reporter strain: PAO-JP2 (plasI-LVAgfp). Fluorescence reported as percentage relative to native ligand 8d (ODHL). Compounds screened at various concentrations against 1 μM native ligand 8d (ODHL). Error bars are ± one S.E.M., calculated from at least three replicate screens.

FIGS. 34A-C show composite 3D micrographs of *P. aeruginosa* biofilms grown on glass slides after 48 hours in the presence of synthetic ligands (at 50 μM); (FIG. 34A) untreated, (FIG. 34B) compound 7h and (FIG. 34C) compound 7o.

FIGS. 35A-35C show composite stacked scanning confocal laser micrographs of *P. aeruginosa* (PAO1(pLVAgfp)) biofilms grown on glass slides after 48 hours in the presence of synthetic ligands (50 μM). Scale bar=50 μm. FIG. 35A. untreated. FIG. 35B. Compound 7h. FIG. 35C. Compound 7o.

FIGS. 36A-36D show composite stacked scanning confocal laser micrographs of *P. aeruginosa* (PAO1 (pLVAgfp)) biofilms grown on glass slides after 48 hours in the presence

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of AHL ligand 7o. Scale bar=10 μm . FIG. 36A. untreated. FIG. 36B. 50 μM compound 7o. FIG. 36C. 25 μM compound 7o. FIG. 36D. 12.5 μM compound 7o.

FIGS. 37A-37D show composite stacked scanning confocal laser micrographs of *P. aeruginosa* (PAO1(pLVAgfp)) biofilms grown on glass slides after 48 hours in the presence of AHL ligand 7h. Scale bar=10 μm . FIG. 37A. 50 μM compound 7h. FIG. 37B. 25 μM compound 7h. FIG. 37C. 12.5 μM compound 7h.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

Many bacterial phenotypic traits are modulated in response to bacterial density that is detected by quorum sensing. These phenotypes have important health consequences in pathogenic bacteria and include virulence, carbapenem antibiotic production, biofilm formation, enzyme synthesis and secondary metabolite synthesis. Modulation or interruption of these signaling pathways can alter the life-cycle of quorum-sensing bacteria and thereby alter their virulence. Other quorum sensing bacteria also have important economic impact in such industries as agriculture, water treatment and power generation.

The quorum sensing system can be manipulated at both the level of the autoinducer and its receptor. Studies have shown that both biofilm formation and expression of other virulence factors has been inhibited by mutating or deleting the receptor, as well as the AI. Therefore, by synthesizing non-native AIs, or AHL, analogs, a similar ability to delete or attenuate microbial virulence can be achieved. Moreover, an efficient method for producing such synthetic analogs provides the opportunity to create a library of autoinducers without using the time-consuming tools of molecular biology to mutate naturally occurring AHLs. Not only does such a method allow for a more rapid and efficient method to create such a library but it also allows the production of AHL analogs that are not per se based on a native model. Further, the use of non-naturally occurring AHL analogs may provide compounds that stimulate quorum sensing pathways in an attenuated manner, bind with different affinities and result in a steric hindrance for the receptor when binding to the promoter thereby eliciting different phenotypic responses.

Generally, there are at least three components to quorum sensing: (1) a receptor/transcription factor; (2) a diffusible signal, the autoinducer (AI); and (3) a recognition site in the promoter of the target gene. The general model for quorum sensing requires a membrane associated activator or transcription factor (LuxR), a cis-acting inverted repeat called the lux box in the promoter region of the gene and the AI, an N-acyl-homoserine lactone (AHL). In *V. fischeri*, the lux genes are responsible for bioluminescence and are transcribed at a low level. Small amounts of AHL diffuse out of the cell and collect in the environment. At high cell densities the AHL accumulates and binds to the receptor/activator presented on the cell membrane. This forms a complex that is internalized, allowing the activator/transcription factor to bind to the lux box, increasing and amplifying the production of the AI and resulting in a cascade effect and increased transcription of the AI.

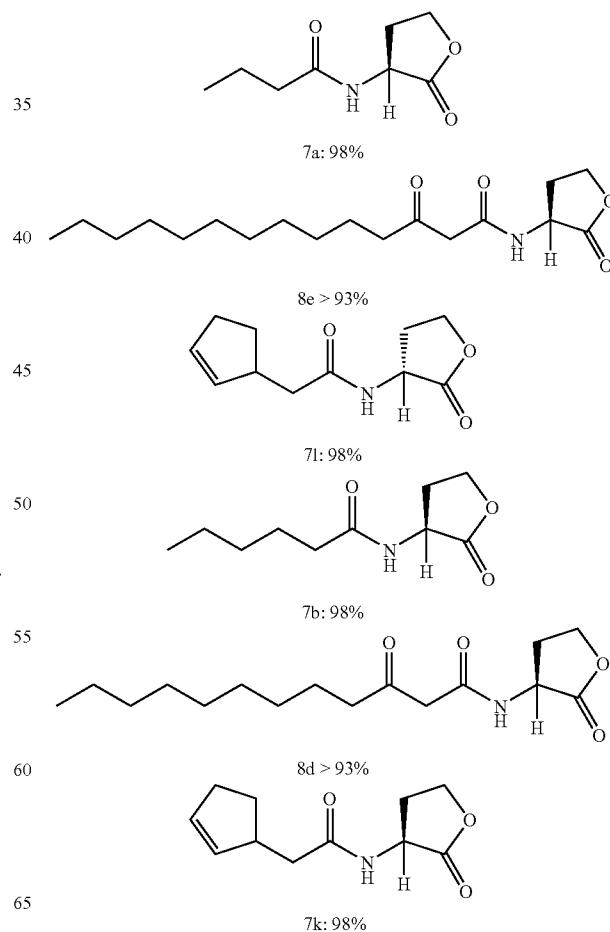
The ability to modulate or disrupt the quorum sensing system presents novel opportunities to affect the metabolic pathways during the bacterial life cycle and thus, the pathogenesis of the quorum-sensing bacteria. By compiling a library of AHL ligands the binding affinities of naturally occurring and non-natural AHL ligands can be explored. Various strategies used to exploiting such compounds include

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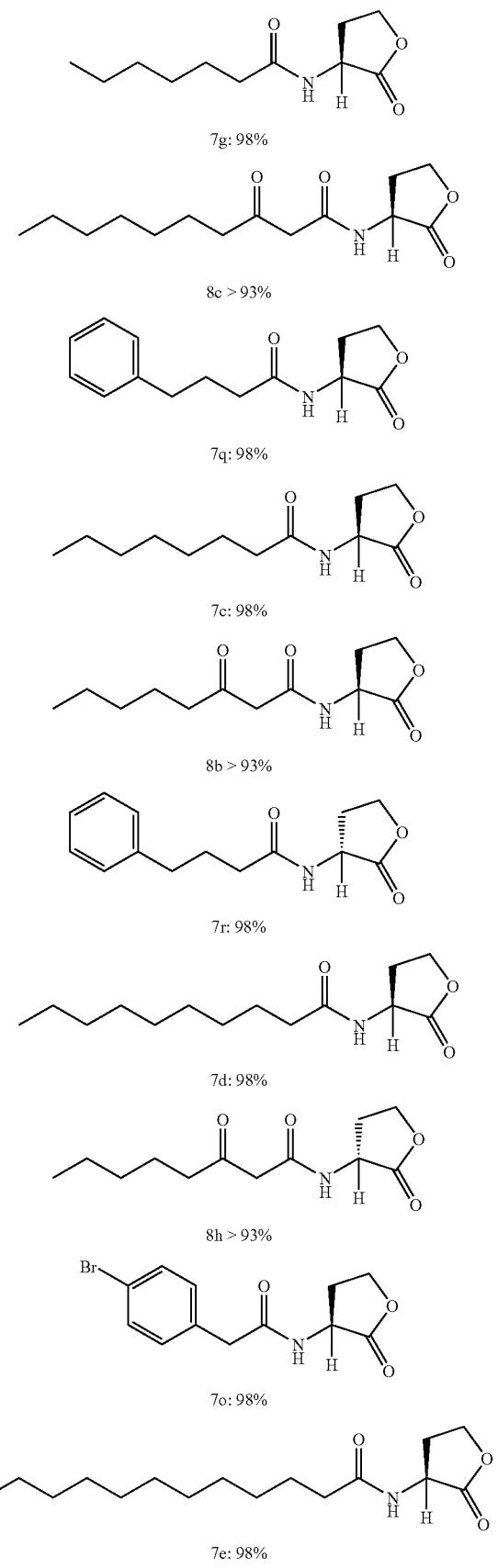
using AHL analogs that inhibit binding of the native ligand without stimulating quorum-sensing pathways; utilizing ligands that prematurely initiate internalization triggering quorum sensing pathways; and using ligands that initiate internalization but inhibit binding of the receptor to the promoter.

The pathogenicity of quorum sensing bacteria can be attenuated or eliminated by inhibiting or reducing the microbe's ability to respond to quorum sensing signals with virulence factors such biofilms. Virulence factors can be inhibited by adding synthetic AHL analogs, to a suspected bacterial bloom. In various exemplary embodiments the analog can be administered in an aqueous solution. By preventing the formation of biofilm formation antibodies, cells of the immune response and antibiotics are allowed to access to bacteria thereby increasing the ability of a host to fight infections. Other strategies include initiating the production of biofilm matrices, before the bacteria are dense enough to produce an exclusionary biofilm allowing the host to mount a more effective immune response while the bacteria remain accessible. In addition, identification of synthetic ligands that bind the receptor with greater or less affinity than the native ligand presents opportunities to inhibit binding of the receptor to the target promoter.

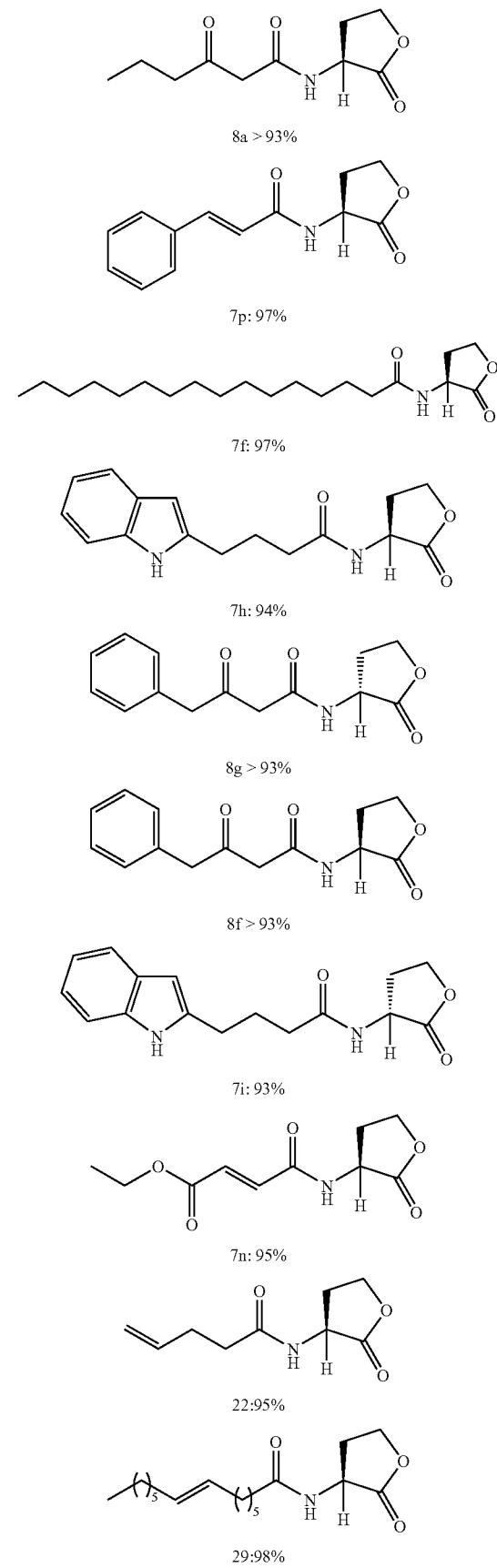
In one exemplary embodiment, the present invention provides quorum sensing compounds comprising naturally occurring AHL ligands and non-natural AHL analogues synthesized by the methods described herein and include:



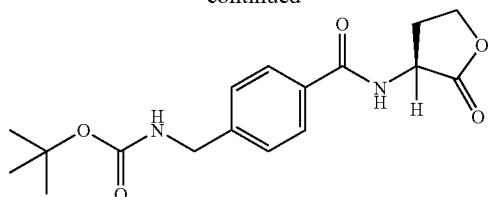
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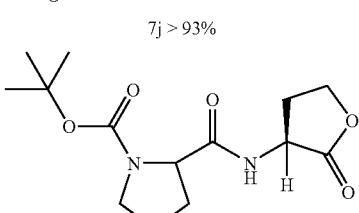
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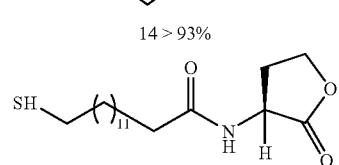
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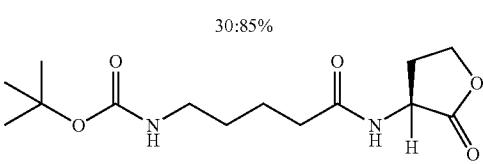
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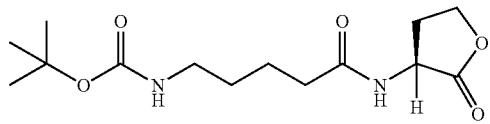


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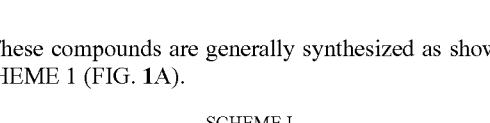
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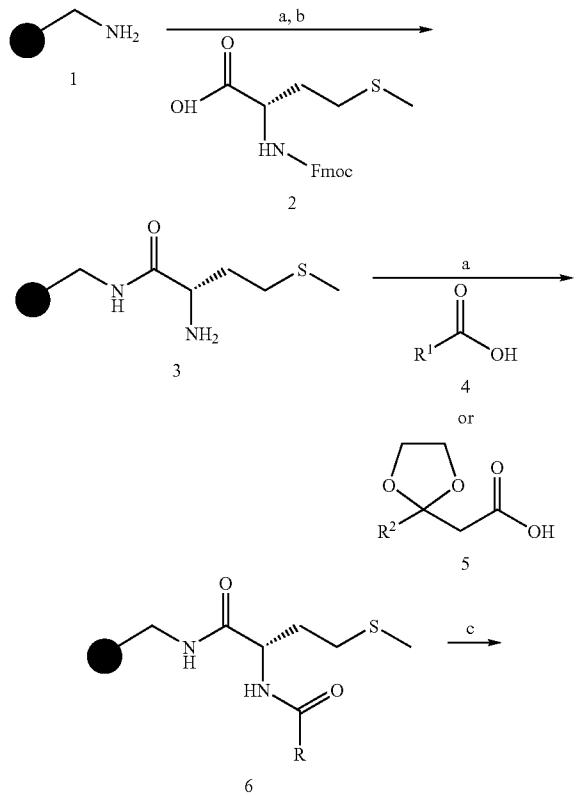
7m > 93%



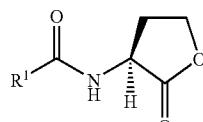
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These compounds are generally synthesized as shown in SCHEME 1 (FIG. 1A).

SCHEME I

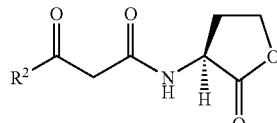


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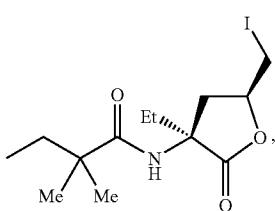
7a-q

or

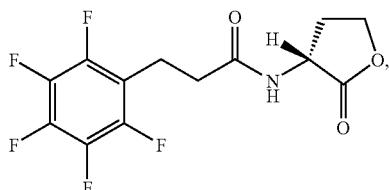


8a-h

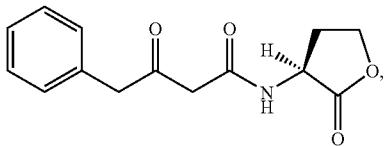
where a=DIC, HOBT, CHCl₃/DMF, microwave 50° C. (2×10 min); b=DMF, 5 microwave 150° C., 7 min; c=CNBr, TFA, CHCl₃/H₂O, microwave 60° C., 30 min. In another exemplary embodiment, the compounds disclosed herein include:



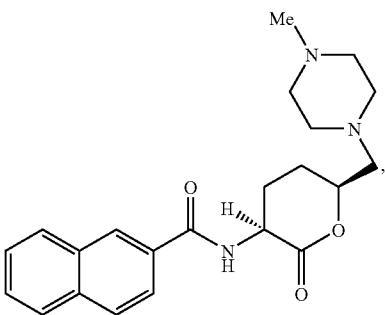
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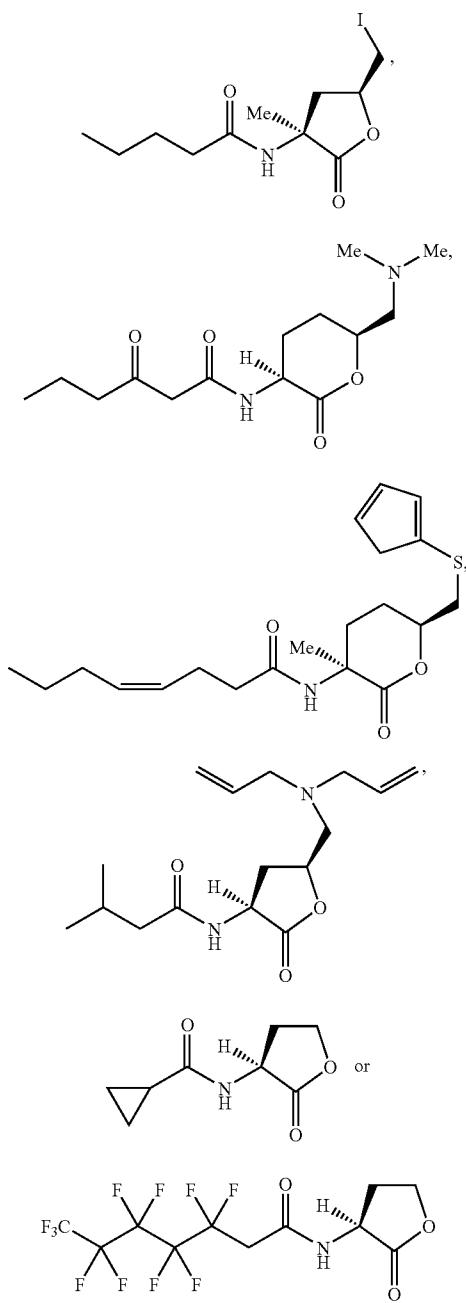


8f

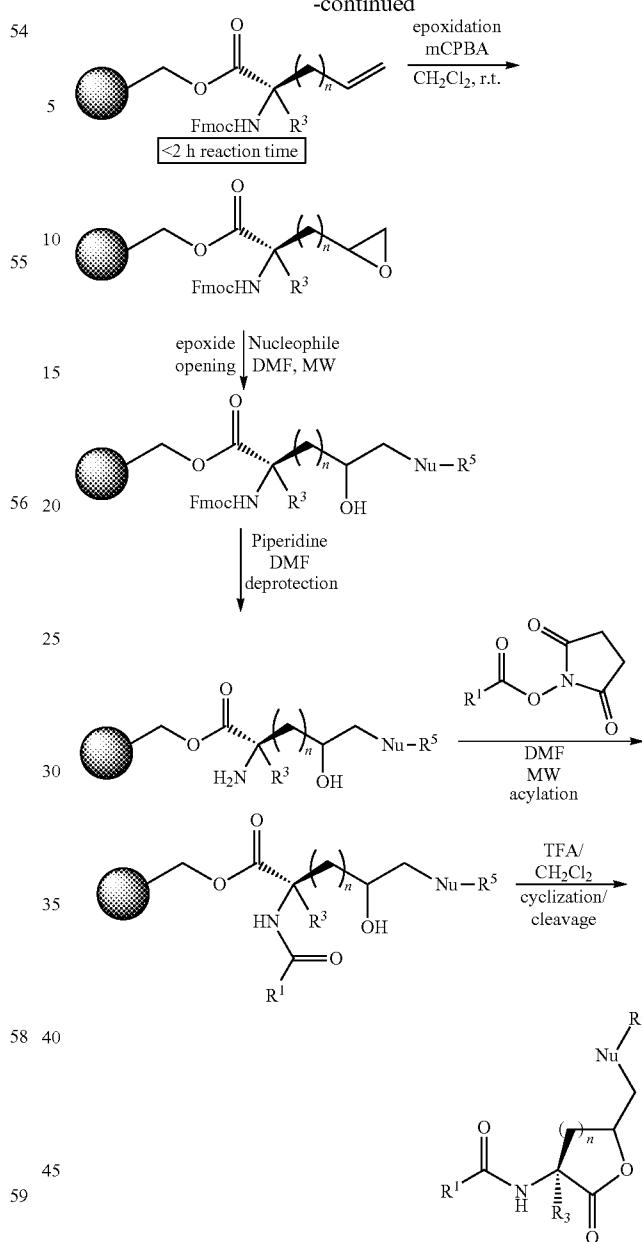


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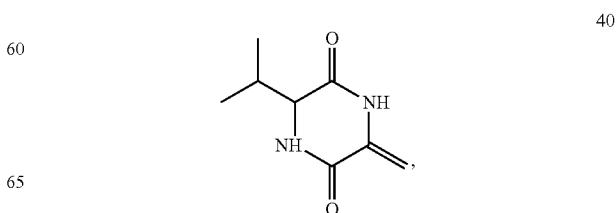
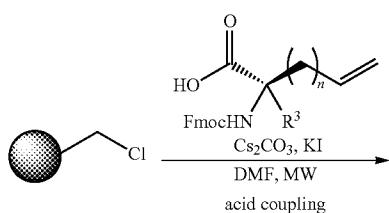


Such compounds are generally synthesized as shown in SCHEME II (FIG. 1B).

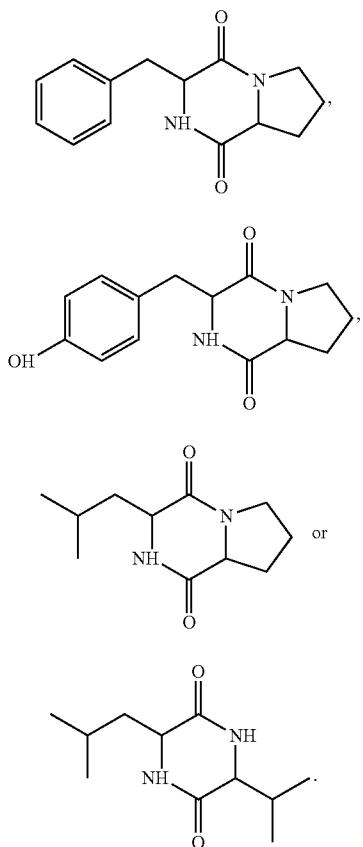
50 In preferred embodiments, the method illustrated by Scheme II comprises the steps of providing a halide derivatized substrate, preferably a chloride derivatized substrate, acid coupling, epoxidation, epoxide opening, deprotection, acylation, cyclization and deleavage from the substrate.

55 In yet another exemplary embodiment, the quorum sensing compounds disclosed herein include:

SCHEME II

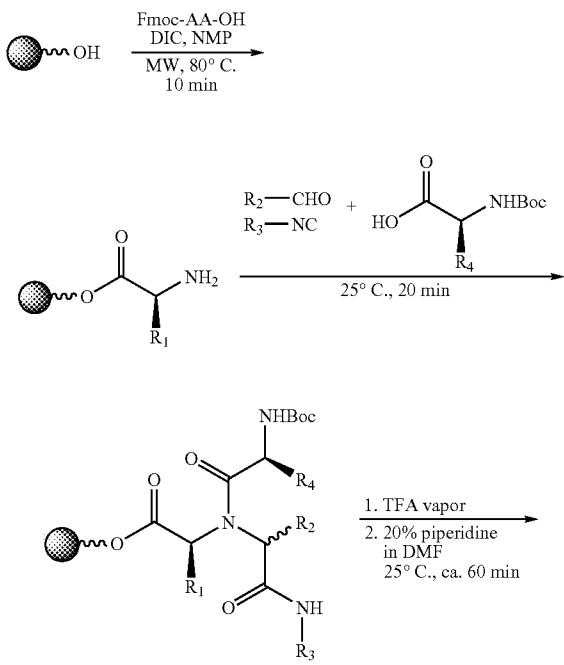


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These compounds are generally synthesized by the method shown in SCHEME III (FIG. 1C).

SCHEME III



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In various exemplary embodiments, the invention described herein provides a method of regulating microbial population density comprising the step of contacting a microbe with a quorum sensing compound as shown above.

15 Contacting may occur in numerous ways.

As defined herein, "contacting" means that the quorum sensing compound used in the present invention is introduced into a sample containing the receptor in a test tube, flask, tissue culture, chip, array, plate, microplate, capillary, or the like, and incubated at a temperature and time sufficient to permit binding of the quorum sensing compound to a receptor. Methods for contacting the samples with the quorum sensing compound or other specific binding components are known to those skilled in the art and may be selected depending on the type of assay protocol to be run. Incubation methods are also standard and are known to those skilled in the art.

43 20 25 30 35 In another embodiment, the term "contacting" means that the quorum sensing compound used in the present invention is introduced into a subject receiving treatment, and the compound is allowed to come in contact *in vivo*.

As used herein, the term "treating" includes preventative as well as disorder remittent treatment. As used herein, the terms "reducing", "suppressing" and "inhibiting" have their commonly understood meaning of lessening or decreasing.

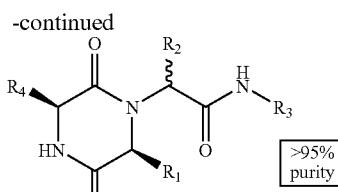
The present invention also provides a method of reducing virulence in a microbe comprising the step of contacting said microbe with a quorum sensing compound, as shown above.

44 45 Further, in another embodiment, the present invention provides a method of modulating biofilm formation in a microbe comprising the step of contacting said microbe with a quorum sensing compound as shown in this disclosure. The invention also provides methods of regulating microbial disease resistance or susceptibility to a microbial disease in a subject comprising the step of contacting said microbe with a quorum sensing compound.

46 50 55 In certain embodiments, the present invention encompasses administering the compounds useful in the present invention to a patient or subject. A "patient" or "subject", used equivalently herein, refers to a plant or an animal. In particular, an animal refers to a mammal, preferably a human. The subject either: (1) has a pathogen remediable or treatable by administration of a quorum sensing compound; or (2) is susceptible to a pathogen that is preventable by administering a quorum sensing compound.

The present invention also provides a method of synthesis of a quorum sensing compound, comprising the steps shown in any of SCHEMES I-III.

60 The compounds shown above, or compounds made using the synthesis steps shown in SCHEMES I-III have industrial uses as well. For example, in industries where biofouling occurs, such as, for example, paper making, power generating and water treating, equipment and/or machinery subject to biofilms can be treated with quorum sensing antagonists to prevent biofilm formation. Such treatment can include spraying or painting the susceptible machinery, component or

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>95% purity

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instrument with a quorum sensing compound or maintaining an effective concentration of a quorum sensing compound in a wash step or effluent step.

Bacteria affecting crops include beneficial species such as those of the *Rhizobium*, genus and other related agriculturally important bacteria such as *Sinorhizobium melilotii* and *Bradyrhizobium japonicum*. Detrimental quorum sensing bacteria, such as species of *Erwinia* are responsible for the soft rot of crops including carrots, potatoes, apples, pears and maize. Still other crop pathogens *Ralstonia solanacearum* (tomato and tobacco); *Xanthomonas campestris* pathovars (peppers and tomatoes); and *Pseudomonas syringae* pathovars (tomato, arabiopsis, legumes).

According to some exemplary embodiments of this invention, beneficial bacteria, such as *Rhizobium* can be encouraged to form symbiotic relationships with crops by treating soil, fertilizer or the like and applying an effective amount of a quorum sensing compound to the crops or tilling it into the soil. Similarly, destructive bacteria can be inhibited from attacking crop plants by applying a quorum sensing antagonist to the crops during growth, by treating the soil or applying as a spray. The crops can also be treated during harvest or distribution by applying a quorum sensing compound. Similar methods are used to encourage ripening of fruit by treatment with ethylene glycol.

Further, it should be appreciated that, in some exemplary embodiments, it may be desirable to treat a component, surgical instrument, machinery, food crop or the like with more than one quorum sensing compound. For example, when conditions may allow various pathogens to grow, it may be desirable to treat the area with a cocktail or mixture of quorum sensing compounds such that optimum compound is utilized for each suspect bacteria. In some embodiments, such quorum sensing compounds may all be inhibitory. In other embodiments some quorum sensing compounds could be inhibitory and some could be stimulatory. For example, in agriculture, a compound could be applied to the soil, much as fertilizer is that would encourage growth of nitrogen fixing bacteria while inhibiting the growth of various rot-causing bacteria. In some embodiments the quorum sensing compound may be applied in a prophylactic manner.

The present invention, thus, generally described, will be understood more readily by reference to the following examples, which are provided by way of illustration and are not intended to be limiting of the present invention.

EXPERIMENTAL BACKGROUND

Various exemplary embodiments of methods according to this invention are described in the following illustrative examples. In these examples, specific products identified by Arabic numerals (e.g., 1, 2, 3, etc.) refer to the specific structures so identified in the following description and synthesis route shown in SCHEME I (FIG. 1A). The present invention describes methods to synthesize AI analogs comprising both AHL agonists and antagonists and thereby create a combinatorial AHL library. In addition, the invention provides methods to screen novel quorum sensing compounds and assay their ability to interact and/or bind with the receptor/activator.

To meet these challenges, a solid-phase synthetic route to both natural and non-natural AHLs (Scheme 1) was developed. The solid-phase methods were chosen because they routinely give improved product purity relative to solution-phase methods, and enable combinatorial library construction (Ley, S. V.; Baxendale, I. R. *Nat. Rev. Drug. Discov.* 2002, 1, 573-586). To date, the use of combinatorial methods to systematically evaluate AHL analogs remains essentially unex-

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plored. To further expedite both solid-phase and library synthesis, microwave (microwave)-assisted reactions were incorporated throughout the route (Blackwell, H. E. *Org. Biomol. Chem.* 2003, 1, 1251-1255; Kappe, C. O. *Angew. Chem. Int. Ed.* 2004, 43, 6250-6284). The four-step synthetic approach developed (SCHEME I) entails first loading amino polystyrene resin (1) with N-Fmoc-L-methionine (2) using a microwave-assisted carbodiimide coupling (DIC) (microwave-assisted reactions were performed in a commercial microwave reactor available from, for example, Milestone, Inc. Shelton, Conn.). Next, thermal Fmoc group removal was followed by a second microwave-assisted DIC coupling with various carboxylic acids (4) or protected β -keto-acids (5) using established procedures (Rathke, M. W.; Nowak, M. A. *Synth. Commun.* 1985, 15, 1039-1049) to generate acylated resin 6. Finally, the classical reaction of cyanogen bromide (CNBr) with L-methionine was used in a microwave-assisted, tandem cyclization-cleavage step to release AHLs 7 and 8 from the solid-support (Ko, D. H.; Kim, D. J.; Lyu, C. S.; Min, I. K.; Moon, H.-s. *Tetrahedron Lett.* 1998, 39, 297-300 (b) Kappel, J. C.; Barany, G. J. *Comb. Chem.* 2005, 7, 78-84, and references therein).

Microwave-Assisted Solid-Phase Synthetic Route to
Natural and Non-Natural AHLs

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The solid-phase route to AHLs is significant because it is the first to provide access to the ca. 15 known natural AHLs from Gram-negative bacteria as well as access to structural analogs thereof. Further, the route delivers compounds in sufficient purity and quantity for biological research. As a test, the inventors synthesized the majority of the natural AHLs (7a-f, 8a-e) in good yields and excellent purities in under 60 minutes total reaction time (Table 2). AHLs 8b, 30 N-3-oxo-octanoyl L-homoserine lactone, (OOHL) and 8d, N-3-oxo-dodecanoyl L-homoserine lactone, (ODHL) were required as critical control molecules for biological work with *Agrobacterium tumefaciens* and *P. aeruginosa*, respectively (Examples 38-42). The high purities and shortened 35 reaction times accessible using this approach underscore the value of microwave-assisted solid-phase chemistry for AHL synthesis. Table 2 lists the naturally occurring AHL's synthesized by the route shown in SCHEME I (FIG. 1A), the organism that is its natural source, the percent purity and yield.

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TABLE 2
Naturally Occurring AHLs Synthesized via SCHEME I

compound	R ¹ or R ²	organism	purity [%] ^{a,b}	yield [%] ^c
7a	C ₈ H ₇	<i>P. aeruginosa</i>	98	65
7b	C ₉ H ₁₁	<i>R. leguminosarum</i>	98	64
7c	C ₉ H ₁₅	<i>Y. pseudotuberculosis</i>	98	76
7d	C ₉ H ₁₉	<i>B. pseudomallei</i>	97	80
7e	C ₁₁ H ₂₃	<i>S. meliloti</i>	98	70
7f	C ₁₃ H ₂₇	<i>R. capsulatus</i>	97	64
8a	C ₈ H ₇	<i>V. Fischeri</i>	>93	63
8b: OOHL ^d	C ₈ H ₁₁	<i>A. tumefaciens</i>	>93	65
8c	C ₅ H ₁₅	<i>V. anguillarum</i>	>93	76
8d: ODHL ^e	C ₉ H ₁₉	<i>P. aeruginosa</i>	>93	62
8e	C ₁₁ H ₂₃	<i>S. meliloti</i>	>93	62

^aPurities of 7a-f determined by integration of GC spectra.

^bPurities of 8a-e determined by ¹H NMR.

^cIsolated yields.

^dN-3-oxo-octanoyl L-homoserine lactone.

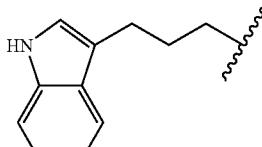
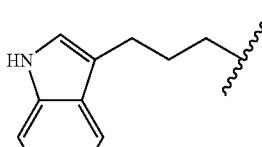
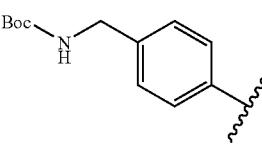
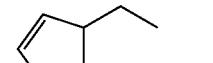
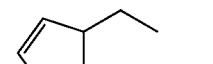
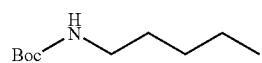
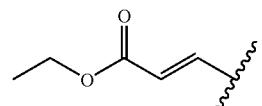
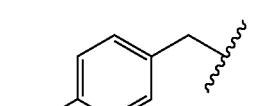
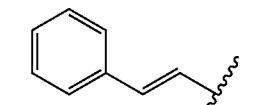
^eN-3-oxo-dodecanoyl L-homoserine lactone.

The solid-phase route of SCHEME I was used for the parallel synthesis of a library of non-natural AHLs used as test

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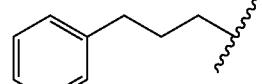
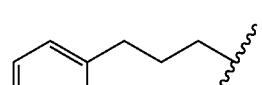
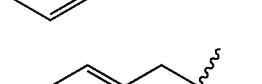
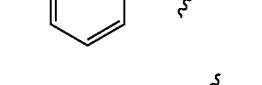
compounds (7g-q, 8f-h). The acyl substituents and stereochemistry of the AHL products were chosen to probe broadly the sterics and functionality present in the AHL binding site of LuxR-type proteins, as revealed in a recent X-ray structure of TraR from *A. tumefaciens*. Zhang, R. G.; Pappas, T.; Brace, J. L.; Miller, P. C.; Oulmassov, T.; Molyneaux, J. M.; Anderson, J. C.; Bashkin, J. K.; Winans, S. C.; Joachimiak, A. *Nature* 2002, 417, 971-974. The synthetic route proved robust and delivered the non-native AHLs in good yields (ca. 70%) and high purities (>93%) (Table 3).

TABLE 3

Non-Natural AHLs Synthesized via SCHEME 1.				
com- pound	R ¹ or R ²	stereo- chemistry ^a	Purity ^{b,c} [%]	yield ^d [%]
7g	C ₆ H ₁₃	L	98	81
7h		L	94	61
7i		D	93	52
7j	Boc- NH- 	L	93	74
7k		L	98	51
7l		D	97	54
7m	Boc- NH- 	L	>93	64
7n		L	93	47
7o		L	98	65
7p		L	93	76

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TABLE 3-continued

Non-Natural AHLs Synthesized via SCHEME 1.				
com- pound	R ¹ or R ²	stereo- chemistry ^a	Purity ^{b,c} [%]	yield ^d [%]
7q		L	95	56
7r		D	95	62
8f		L	>90	59
8g		D	>90	64
8h	C ₅ H ₁₁	D	>93	54

^aDetermined by the N-Fmoc methionine (2) starting material^bPurities of 7g-h, 7l-m, and 7o-r determined by integration of GC spectra (flame ionization and MS detection)^cPurities of 7i-k, 7nm and 8f-h determined by ¹H NMR analyses.^dIsolated yield.

Synthesis Steps: General Experimental Information

¹H NMR and ¹³C NMR spectra were recorded on a Bruker AC-300 spectrometer in deuterated solvents at 300 MHz and 75 Hz, respectively. Chemical shifts are reported in parts per million (ppm, δ) using tetramethyl silane (TMS) as an internal reference (0.0 ppm). Couplings are reported in hertz. Electrospray ionization (ESI) MS were obtained using a Shimadzu LCMS-2010 system (Columbia, Md.) equipped with two pumps (LC10ADvp), controller (SCL-10Avp), autoinjector (SIL-10ADvp), UV diode array detector (SPD-M10Avp), and single quadrupole analyzer. FT-IR and attenuated total reflectance (ATR)-IR spectra were recorded with a Bruker Tensor 27 spectrometer, outfitted with a single reflection MIRacle Horizontal ATR unit from Pike Technologies. A ZnSe crystal with spectral range 20,000 to 650 cm⁻¹ was used for ATR-IR measurements. UV spectra were recorded using an HP-8452 UV-Visible spectrometer running Chemstation software. GC-MS spectra were obtained using a Shimadzu GC-17A system (Columbia, Md.) equipped with a QP-5000 mass spectrometer. A Restek RTX-5 crossbond 95% polysiloxane GC column was used with following general GC gradient: injection temperature 300° C.; initial oven temperature 100° C.; hold 3 minutes; ramp at 20° C./minutes to 300° C.; hold 2-15 minutes for a total run time of 15-30 minutes. Optical rotations ([α]24D) were measured on a Perkin-Elmer 241 digital polarimeter at 25° C.

All reagents were purchased from commercial sources (Alfa-Aesar, Ward Hill, Mass.; Aldrich, Milwaukee, Wis.; Acros Organics, Geel, Belgium; and Sigma, St. Louis Mo.) and used without further purification. Solvents were purchased from commercial sources (Aldrich and J. T. Baker, Phillipsburg, N.J.) and used as is, with the exception of dichloromethane (CH₂Cl₂), which was distilled over calcium hydride immediately prior to use. All solid-phase syntheses

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were performed using aminomethyl polystyrene resin (Nova-Biochem, Merck, KGaA, Darmstadt, Del.) 100-200 mesh; loading 1.1-1.2 mmol/g.

Microwave Instrumentation: Solid-phase reactions were carried out using either Milestone (Shelton, Conn.) or CEM (Mathews, N.C.) commercial microwave reactors. The Milestone MicroSYNTH Labstation is a multimodal microwave synthesis reactor equipped with a continuous power source (1000 W max). This instrument was interfaced with an Ehtos MicroSYNTH Lab Terminal PC running EasyWave reaction monitoring software. Using this reactor system, microwave irradiation was applied to reactions using either wattage (power) control or temperature control. The microwave reactor is equipped with a fiber-optic temperature sensor that allows direct monitoring of the internal temperature of reaction vessels, and an infrared sensor (installed in the side wall of the reactor cavity) that monitors the surface temperature of any reaction vessel inside the cavity. The system also has a rotating plate in the cavity and the capability for stirring (using magnetic stir-bars) during reactions.

The CEM Discover is a monomodal microwave synthesis reactor equipped with a 300 W (max) power source. The system has an attached Explorer automated synthesis workstation module, with four autosampler racks that each holds six samples. The instrument is interfaced with a Dell Inspiron PC running ChemDriver Discovery reaction monitoring software. Using this system, microwave irradiation can be applied to reactions using wattage, pressure, or temperature control. The CEM microwave reactor is equipped with an infrared temperature sensor positioned below the reaction vessel to control temperature. The system also has the capability for stirring (using magnetic stir-bars) during reactions.

All microwave-assisted reactions reported herein were performed using temperature control to monitor and control microwave irradiation.

Solid-Phase Library Synthesis Techniques

Solid-phase reactions were performed in either 100 mL round bottom flasks in the Milestone microwave reactor or 10 mL glass CEM microwave vessels (part #908035) in the CEM microwave reactor. Between synthesis steps, the solid-phase resin was washed with solvents stored in standard polypropylene Nalgene squirt bottles on a Vac-Man vacuum manifold (Promega, part #: A7231) using 8 mL polypropylene sample reservoirs (Alltech, part #: 210208) equipped with 20 μ m fits (Alltech, part #: 211408). Liquid reagents were dispensed during synthesis using Brinkman Eppendorf pipettmen (calibrated for variable solvent delivery) equipped with disposable polypropylene pipette tips.

Synthesis of 1,3-Dioxolane Protected β -Keto Acids

The 1,3-dioxolane protected β -keto acids building blocks (5) used in this study were prepared via a modified version of the methods reported by Barnick and Rathke (Barnick, J. W. F. K.; van der Baan, J. L.; Bickelhaupt, F. *Synthesis* 1979, 79, 787-788; Rathke, M. W.; Nowak, M. A. *Synth. Commun.* 1985, 15, 1039-1049). A representative synthesis is outlined below in Examples 1-3 and illustrated in SCHEMES IV-VI.

Example 1

Synthesis of 3-Oxoctanoic Acid (10b) (SCHEME IV)

A stirred solution of bis-trimethylsilyl malonate (21.6 g, 71.5 mmol) in 100 mL of anhydrous diethyl ether was cooled

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to -78° C. To this solution, n-butyl lithium (1.6 M in ether, 44.7 mL, 71.5 mmol) was added slowly, keeping the temperature below -60° C. Upon completion of addition, the reaction was allowed to warm to -10° C., at which time hexanoyl chloride (5 mL, 35.75 mmol) was added quickly and allowed to stir for 30 minutes. Next, 150 mL of a cold, aq. 5% sodium bicarbonate solution was added, and the resulting solution was stirred vigorously for 30 minutes. The aq. layer was separated out and acidified with cold 4N sulfuric acid until pH=2. The aq. layer was then extracted $2\times$ 50 mL with diethyl ether, dried over MgSO_4 , and concentrated down in vacuo to afford a white solid. This solid could be further purified by recrystallization from hexane, if necessary. 4.9 g, 87% yield. ^1H NMR (300 MHz, CDCl_3) δ =3.49 (s, 2H, CH_2), 2.59 (t, 2H, $J=7.3$ Hz, CH_2), 1.64 (p, 2H, $J=7.4$ Hz, CH_2), 1.34 (m, 4H, CH_2CH_2), 0.92 (t, 3H, $J=6.9$ Hz, CH_3); ^{13}C NMR (75 MHz, CDCl_3) δ =204.4, 182.8, 87.9, 48.1, 43.4, 31.3, 23.5, 14.1 ppm.

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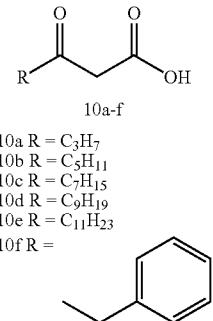
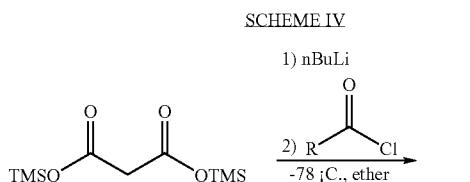
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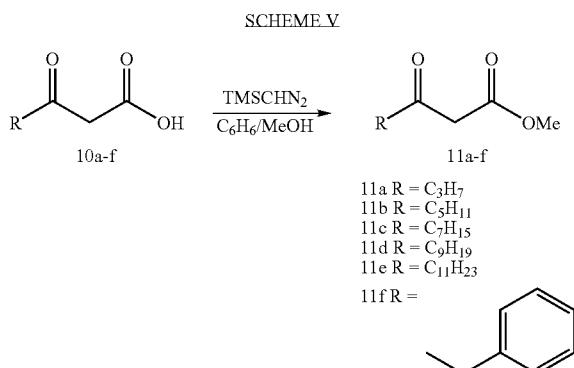


Example 2

Synthesis of Methyl-3-Oxoctanoate (11b) (SCHEME V)

To a stirred solution of 10b (5.5 g, 35 mmol) in 150 mL of a 4:1 mixture of benzene and methanol, TMSCHN_2 in diethyl ether (2M, 21 mL, 42 mmol) was added over a period of 10 minutes. The reaction was allowed to stir for 30 minutes, after which the reaction mixture was concentrated in vacuo to afford 11b as a yellow oil. This material was used in the subsequent step (SCHEME IV) with no further purification. 6.1 g, 95% yield. ^1H NMR (300 MHz, CDCl_3) δ =3.73 (s, 3H, CH_3) 3.44 (s, 2H, CH_2), 2.55 (t, 2H, $J=7.3$ Hz, CH_2), 1.64 (p, 2H, $J=7.4$ Hz, CH_2), 1.36 (m, 4H, CH_2CH_2), 0.91 (t, 3H, $J=6.9$ Hz, CH_3); ^{13}C NMR (75 MHz, CDCl_3) δ =202.9, 167.9, 128.5, 52.7, 49.2, 43.2, 31.3, 23.3, 14.0 ppm.

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Example 3

Synthesis of 3,3-Ethylenedioxooctanoic Acid (5b)
(SCHEME VI)

To a stirred solution of methyl-3-oxooctanoate (11b) (5.6 g, 32.5 mmol) in 125 mL of benzene, ethylene glycol (20.2 g, 325 mmol) and pTsOH (0.617 g, 3.25 mmol) was added. The flask was equipped with a condenser and Dean-Stark trap and heated to reflux for 24 h. The reaction mixture was concentrated in vacuo and diluted in 100 mL of diethyl ether. The organic layer was washed with 2×25 mL of 10% aq. NaOH, then 2×25 mL of saturated NaCl solution, dried over MgSO₄, and concentrated in vacuo to afford a clear oil. This oil was subjected to saponification by treatment with 1N NaOH (150 mL) and MeOH (75 mL) for 6 h. The basic solution was concentrated in vacuo, chilled in an ice bath, and acidified with cold concentrated HCl to a pH=2. The acidified solution was extracted with 2×75 mL of diethyl ether, dried over MgSO₄, and concentrated in vacuo to afford 3,3-ethylenedioxooctanoic acid 5b as a clear oil. 3.3 g, 50% yield. ¹H NMR (300 MHz, CDCl₃) δ=4.06-3.95 (m, 4H, OCH₂CH₂O), 2.70 (s, 2H, CH₂), 1.83 (t, 2H, J=7.3 Hz, CH₂), 1.42-1.18 (m, 6H, J=7.4 Hz, (CH₂)₃), 0.94 (t, 3H, J=6.9 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=175.3, 109.5, 65.3, 37.8, 32.0, 22.7, 14.1 ppm.

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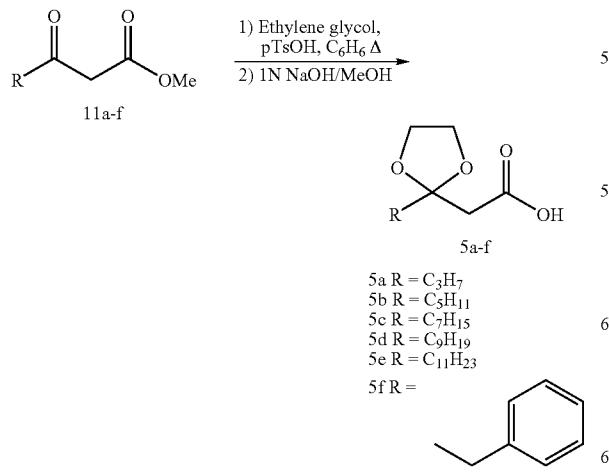
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SCHEME VI



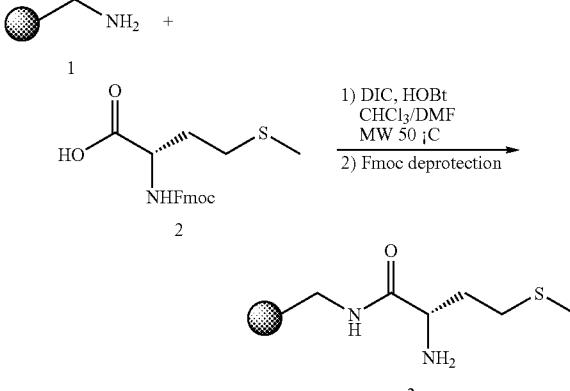
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Example 4

Synthesis of Natural and Unnatural Acylated Homoserine Lactone Analogs Representative n-Fmoc-Methionine Resin Loading Protocol
(SCHEME VII)

Aminomethyl polystyrene resin (1, 5.2 g, 6 mmol) was pre-swelled in 40 mL of CHCl₃ in a 100 mL round-bottom flask for 10 minutes at room temperature. In a separate flask, an activated solution of N-Fmoc-L-methionine (2, 6.7 g, 18 mmol), 1-hydroxybenzotriazole (HOBT (2.8 g, 21 mmol)), and N,N-diisopropyl-carbodiimide (DIC, 3.8 mL, 24 mmol) was prepared in 50 mL of DMF. This activated solution was stirred for 10 minutes at room temperature and then added to the swelled resin. The reaction flask was equipped with a stir-bar and sealed with rubber septum that has been pierced to allow for the insertion of the Milestone fiber optic temperature probe (in a protective sheath). The reaction flask was placed into the Milestone Microsynth Labstation and irradiated for 10 minutes at 50° C. (3 minutes ramp to 50° C. with a max of 600 W, hold 10 minutes at 50° C. with a max of 600 W). The resin then was filtered and washed with 250 mL each of DMF, water, EtOH, and CH₂Cl₂ and dried in vacuo. This coupling process was repeated 1× to yield N-Fmoc-L-methionine resin at 0.8-0.9 mmol/g loading as quantified by UV absorbance.

SCHEME VII



Example 5

Representative UV Fmoc Quantitation Protocol

Approximately 20 mg of N-Fmoc-L-methionine loaded resin was submersed in 3 mL of a 4% 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) solution in DMF and stirred for 30 minutes at room temperature. Thereafter, 100 μL of the solution was removed and diluted to 3.0 mL in DMF. A 1.0 mL aliquot of this solution was withdrawn and the UV absorbance was read at 296 nm ($\epsilon_{296}=9500 \text{ M}^{-1} \text{ cm}^{-1}$) in a quartz cuvette; loadings were calculated according to standard methods.

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Example 6

Representative Microwave-Assisted Fmoc-Deprotection Protocol

Approximately 300 mg of N-Fmoc-L-methionine loaded resin was placed in a 10 mL CEM microwave vial with 4 mL

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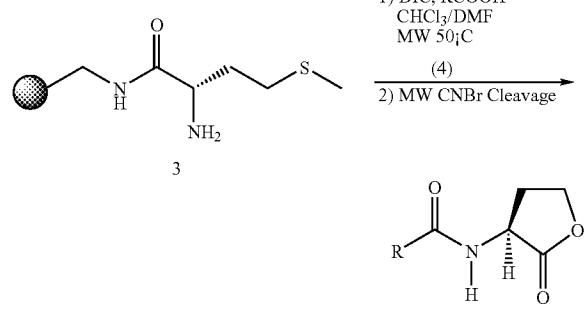
of DMF and irradiated in the CEM Discover at 150° C. for 6 minutes (300 W max. wattage). UV Fmoc-quantitation was performed as described above.

Example 7**Alternate Room Temperature Fmoc Deprotection Protocol**

N-Fmoc-L-methionine loaded resin was placed into the appropriate size container and stirred with a solution of 20% piperidine/DMF solution for 30 minutes. The solution was drained and the process was repeated. UV Fmoc-quantitation was performed as described above.

Example 8**Representative Synthesis of AHLs (7a-r) (SCHEME VIII)**

In a 10 mL CEM microwave vial equipped with a stir-bar, methionine resin 3 (300 mg, 0.295 mmol) was pre-swelled in 1.5 mL of CHCl₃ for 5 minutes at room temperature. An activated solution of carboxylic acid 4 (1.03 mmol) and DIC (1.5 mmol) was prepared in a separate vial in 2 mL of DMF and stirred for 5 minutes. The activated solution was added to the swelled resin, and the reaction mixture was subjected to microwave irradiation for 10 minutes at 50° C. (ramp time 30 sec, hold for 10 minutes at 50° C., max. wattage 300 W). The resin then was filtered and washed 2× with 50 mL each of DMF, H₂O, EtOH, and CH₂Cl₂ and dried in vacuo. To obtain the best yields, this process was repeated again. To affect compound cleavage, resin was treated with 7 mL of a 5:2 solution of 1.5 M CNBr in CHCl₃ and 1% aqueous TFA solution and subjected to microwave irradiation for 30 minutes at 60° C. (ramp time 60 sec, hold for 30 minutes at 60° C., max. wattage 300 W). The AHL product was eluted from the resin with 5 mL of CHCl₃, washed with water (3×10 mL), dried over MgSO₄, and concentrated in vacuo to yield the AHL as a white powder. Purities and yields for 7a-h are reported in Tables 2 and 3.

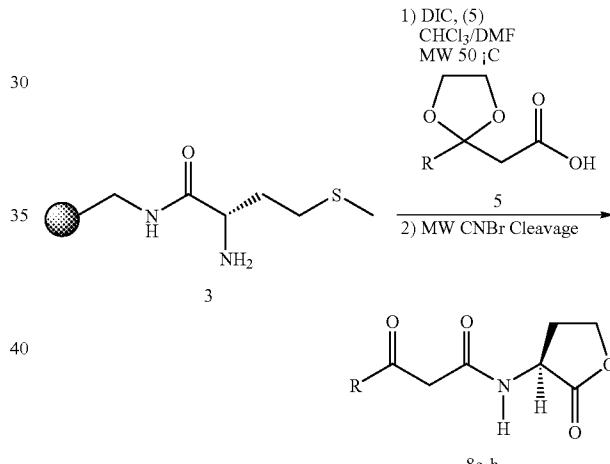
SCHEME VIII**Example 9****Representative Synthesis of 3-Oxo AHLs (8a-h) (SCHEME IX)**

In a 10 mL CEM microwave vial, methionine resin 3 (300 mg, 0.295 mmol) was pre-swelled in 1.5 mL of CHCl₃ for 5

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minutes at room temperature. An activated solution of 3,3-ethylenedioxocarboxylic acid 5 (1.03 mmol) and DIC (1.5 mmol) was prepared in a separate vial in 2 mL of DMF and stirred for 5 minutes. The activated solution was added to the swelled resin, and the reaction mixture was subjected to microwave irradiation for 10 minutes at 50° C. (ramp time 30 sec, hold for 10 minutes at 50° C., max. wattage 300 W). The resin then was filtered and washed 2× with 50 mL each of DMF, H₂O, EtOH, and CH₂Cl₂ and dried in vacuo. To obtain the best yields, this process was repeated again. To effect compound cleavage, resin was treated with 7 mL of a 5:2 solution of 1.5 M CNBr in CHCl₃ and 1% aqueous TFA solution and subjected to microwave irradiation for 30 minutes at 60° C. (ramp time 60 sec, hold for 30 minutes at 60° C., max. wattage 300 W). AHL was eluted from the resin with 5 mL of CHCl₃, washed with water (3×10 mL), dried over MgSO₄, and concentrated in vacuo. This material was stirred for 30 minutes in 5 mL of 50% TFA/CH₂Cl₂ to affect β-keto deprotection. The solution was washed with water (2×5 mL), dried over MgSO₄, and concentrated in vacuo to yield 3-oxo AHL product as a white powder. Purities and yields for 8a-h are reported in Tables 2 and 3.

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SCHEME IX**Example 10****Alternative Room Temperature Microwave Cleavage Protocol**

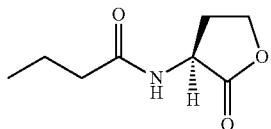
To effect compound cleavage, resin was stirred at room temperature in 7 mL of a 5:2 solution of 1.5 M CNBr in CHCl₃ and 50% aqueous TFA solution for 24 h. The AHL product was eluted from the resin with 5 mL of CHCl₃, washed with water (3×10 mL), dried over MgSO₄, and concentrated in vacuo to yield the 3-oxo AHL as a white powder.

FIGS. 2 through 27 show GC-MS and/or NMR spectra (as noted) used to confirm the identity and purity of the compounds of the synthetic AHL analogs listed in Tables 2 and 3. Each of the compounds 7a-7r and 8a-8h were synthesized using the overall synthesis route illustrated in SCHEME I to provide both the naturally occurring and non-natural AHLs.

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Example 11

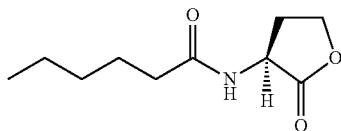
N-butanoyl-L-homoserine lactone (7a)



N-butanoyl-L-homoserine lactone (7a) was synthesized by the method described in Example 8. FIG. 2A shows the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 2B is the NMR spectrum where: ¹H NMR (300 MHz, CDCl₃) δ=5.97 (s, 1H, NH), 4.59 (ddd, 1H, J=5.9 Hz, CH-lac), 4.51 (td, 1H, J=1.1 Hz, CH-lac), 4.34 (ddd, 1H, J=5.9 Hz, CH-lac), 2.93 (ddd, 1H, J=1.1 Hz, CH-lac), 2.27 (t, 2H, J=7.5 Hz, CH₂), 2.17 (ddd, 1H, J=1.7 Hz, CH-lac), 1.29 (h, 2H, J=7.4 Hz, CH₂), 0.99 (t, 3H, J=7.1 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=175.7, 173.7, 65.9, 48.9, 35.9, 31.4, 30.1, 29.0, 28.8, 25.2, 22.4, 13.8; GC-MS: expected m/z=227, observed [M+]=227; [α_D]=+15.9 (c=3.4 mg/mL; CHCl₃); IR (cm⁻¹): 3684, 3619, 3427, 3020, 2930, 2858, 2401, 1780, 1673, 1511, 1423, 1381, 1216, 1019, 929, 772, 669.

Example 12

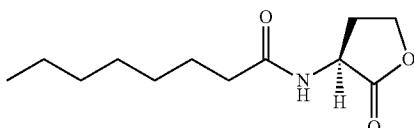
N-hexanoyl-L-homoserine lactone (7b)



N-hexanoyl-L-homoserine lactone (7b) was synthesized by the method described in Example 7. FIG. 3A shows the GC-MS spectrum assaying purity (inset illustrates compound structure). FIG. 3B is the NMR spectrum where: ¹H NMR (300 MHz, CDCl₃) δ=6.22 (s, 1H, NH), 4.63 (ddd, 1H, J=6.1 Hz, CH-lac), 4.51 (td, 1H, J=1.0 Hz, CH-lac), 4.34 (ddd, 1H, J=5.9 Hz, CH-lac), 2.89 (dddd, 1H, J=1.4 Hz, CH-lac), 2.28 (t, 2H, J=7.4 Hz, CH₂), 2.20 (ddd, 1H, J=2.2 Hz, CH-lac), 1.71 (t, 2H, J=7.4 Hz, CH₂), 1.34 (m, 4H, (CH₂)₂), 0.93 (t, 3H, J=7.1 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=175.9, 174.1, 66.4, 49.4, 36.4, 31.6, 30.8, 25.4, 22.6, 14.1; GC-MS: expected m/z=199, observed [M+]=199; [α_D]=+15.9 (c=3.4 mg/mL; CHCl₃); IR (cm⁻¹): 3689, 3620, 3426, 3020, 2963, 2401, 1781, 1675, 1510, 1381, 1216, 1020, 929, 757, 669.

Example 13

N-octanoyl-L-homoserine lactone (7c)



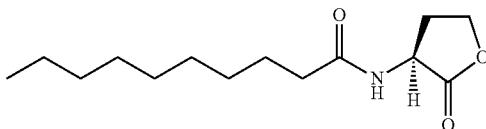
N-octanoyl-L-homoserine lactone (7c) was synthesized by the method described in Example 8. FIG. 4A shows the GC-

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MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 4B shows the NMR spectrum where: ¹H NMR (300 MHz, CDCl₃) δ=6.44 (s, 1H, NH), 4.65 (ddd, 1H, J=6.5 Hz, CH-lac), 4.50 (t, 1H, J=9.2 Hz, CH-lac), 4.34 (ddd, 1H, J=2.3 Hz, CH-lac), 2.85 (dddd, 1H, J=6.1 Hz, CH-lac), 2.28 (t, 2H, J=7.4 Hz, CH₂), 2.22 (ddd, 1H, J=2.3 Hz, CH-lac), 1.67 (t, 2H, J=7.1 Hz, CH₂), 1.30 (m, 8H, (CH₂)₄), 0.91 (t, 3H, J=6.3 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=175.5, 173.7, 65.9, 48.9, 35.9, 31.4, 30.1, 29.0, 28.8, 25.2, 22.4, 13.8; GC-MS: expected m/z=227, observed [M+]=227; [α_D]=+15.9 (c=3.4 mg/mL; CHCl₃); IR (cm⁻¹): 3684, 3619, 3427, 3020, 2930, 2858, 2401, 1780, 1673, 1511, 1423, 1381, 1216, 1019, 929, 772, 669.

Example 14

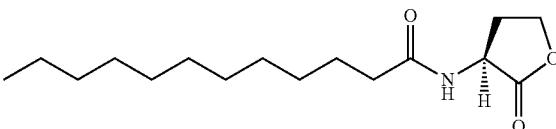
N-decanoyl-L-homoserine lactone (7d)



N-decanoyl-L-homoserine lactone (7d) was synthesized by the method described in Example 8. FIG. 5A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 5B is an NMR spectrum where: ¹H NMR (300 MHz, CDCl₃) δ=6.16 (s, 1H, NH), 4.62 (ddd, 1H, J=6.0 Hz, CH-lac), 4.51 (td, 1H, J=1.0 Hz, CH-lac), 4.34 (ddd, 1H, J=5.9 Hz, CH-lac), 2.90 (dddd, 1H, J=8.3 Hz, CH-lac), 2.28 (t, 2H, J=6.9 Hz, CH₂), 2.15 (ddd, 1H, J=3.3 Hz, CH-lac), 1.70 (t, 2H, J=7.5 Hz, CH₂), 1.30 (m, 12H, (CH₂)₆), 0.91 (t, 3H, J=6.6 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=175.7, 173.9, 66.2, 49.3, 36.3, 31.9, 30.7, 29.5, 29.4, 29.3, 25.5, 22.7, 14.2; GC-MS: expected m/z=255, observed [M+]=255; [α_D]=+15.5 (c=2.9 mg/mL; CHCl₃); IR (cm⁻¹): 3684, 3620, 3427, 3020, 2928, 2857, 2401, 1780, 1673, 1511, 1423, 1381, 1216, 1019, 929, 772, 669.

Example 15

N-dodecanoyl-L-homoserine lactone (7e)

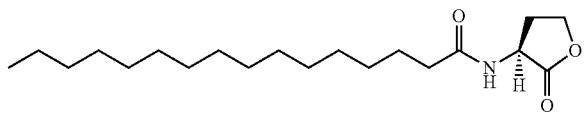


N-dodecanoyl-L-homoserine lactone (7e) was synthesized by the method described in Example 8. FIG. 6A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure) while 6B is the NMR spectrum where: ¹H NMR (300 MHz, CDCl₃) δ=6.22 (s, 1H, NH), 4.63 (ddd, 1H, J=6.1 Hz, CH-lac), 4.50 (t, 1H, J=9.1 Hz, CH-lac), 4.34 (ddd, 1H, J=5.9 Hz, CH-lac), 2.86 (dddd, 1H, J=5.9 Hz, CH-lac), 2.28 (t, 2H, J=6.9 Hz, CH₂), 2.19 (ddd, 1H, J=2.9 Hz, CH-lac), 1.67 (t, 2H, J=6.8 Hz, CH₂), 1.30 (m, 18H, (CH₂)₉), 0.91 (t, 3H, J=6.5 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=176.1, 174.3, 66.6, 49.7, 36.7, 32.4, 31.0, 30.1, 29.9, 29.7, 29.6, 25.9, 23.2, 14.6; GC-MS: expected m/z=283, observed [M+]=283; [α_D]=+15.1 (c=3.5 mg/mL; CHCl₃); IR (cm⁻¹): 3684, 3621, 3426, 3020, 2928, 2856, 2401, 1780, 1673, 1511, 1381, 1216, 1019, 929, 760, 669.

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Example 16

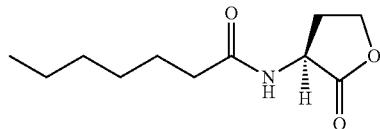
N-hexadecanoyl-L-homoserine lactone (7f)



N-hexadecanoyl-L-homoserine lactone (7f) was synthesized by the method described in Example 8. FIG. 7A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 7B is the NMR spectrum where: ^1H NMR (300 MHz, CDCl_3) δ =6.03 (s, 1H, NH), 4.59 (ddd, 1H, J =5.6 Hz, CH-lac), 4.50 (t, 1H, J =8.2 Hz, CH-lac), 4.33 (ddd, 1H, J =5.7 Hz, CH-lac), 2.92 (dddd, 1H, J =1.3 Hz, CH-lac), 2.28 (t, 2H, J =6.9 Hz, CH_2), 2.20 (ddd, 1H, J =8.8 Hz, CH-lac), 1.69 (t, 2H, J =7.1 Hz, CH_2), 1.31 (m, 24H, $(\text{CH}_2)_{12}$), 0.91 (t, 3H, J =6.4 Hz, CH_3); ^{13}C NMR (75 MHz, CDCl_3) δ =175.8, 174.0, 66.4, 49.6, 36.5, 32.2, 31.0, 30.0, 29.7, 29.6, 29.5, 25.7, 23.0, 14.4; GC-MS: expected m/z=286, observed [M+]=286; $[\alpha_D]=+14.2$ ($c=2.6$ mg/mL; CHCl_3); IR (cm^{-1}): 3684, 3620, 3480, 3426, 3020, 2977, 2401, 2362, 2254, 1780, 1674, 1603, 1514, 1217.

Example 17

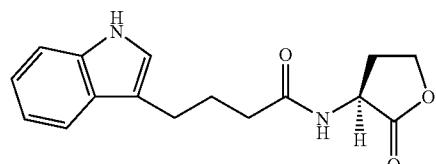
N-heptanoyl-L-homoserine lactone (7g)



N-heptanoyl-L-homoserine lactone (7g) was synthesized by the method described in Example 8. FIG. 8A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 8B is the NMR spectrum where: ^1H NMR (300 MHz, CDCl_3) δ =6.28 (s, 1H, NH), 4.64 (ddd, 1H, J =6.4 Hz, CH-lac), 4.50 (td, 1H, J =1.2 Hz, CH-lac), 4.34 (ddd, 1H, J =5.8 Hz, CH-lac), 2.87 (dddd, 1H, J =8.4 Hz, CH-lac), 2.28 (t, 2H, J =6.7 Hz, CH_2), 2.20 (ddd, 1H, J =2.5 Hz, CH-lac), 1.70 (t, 2H, J =7.8 Hz, CH_2), 1.38 (m, 6H, $(\text{CH}_2)_3$), 0.91 (t, 3H, J =6.7 Hz, CH_3); ^{13}C NMR (75 MHz, CDCl_3) δ =175.5, 173.6, 65.9, 49.0, 36.0, 31.3, 30.3, 28.7, 25.2, 22.3, 13.8; GC-MS: expected m/z=213, observed [M+]=213; $[\alpha_D]=+12.6$ ($c=1.9$ mg/mL; CHCl_3); IR (cm^{-1}): 3685, 3621, 3427, 3020, 2931, 2401, 1780, 1674, 1510, 1423, 1381, 1216.

Example 18

N-(indole-3-butanoyle)-L-homoserine lactone (7h)



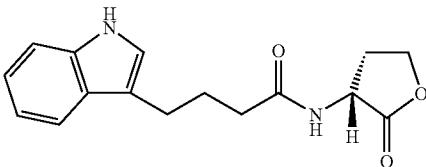
N-(indole-3-butanoyle)-L-homoserine lactone (7h) was synthesized by the methods described in Example 8. FIG. 9A

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is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 9B is the NMR spectrum where: ^1H NMR (300 MHz, CDCl_3) δ =8.08 (s, 1H, NH-indole), 7.59 (d, 1H, J =7.6 Hz, CH—Ar), 7.35 (d, 1H, J =7.1 Hz, CH—Ar), 7.20 (td, 1H, J =1.2 Hz, CH—Ar), 7.12 (td, 1H, J =1.2 Hz, CH—Ar), 6.97 (d, 1H, J =2.3 Hz, CH—Ar), 6.05 (d, 1H, J =5.8 Hz, NH), 4.54 (ddd, 1H, J =6.3 Hz, CH-lac), 4.43 (td, 1H, J =0.9 Hz, CH-lac), 4.26 (ddd, 1H, J =6.0 Hz, CH-lac), 2.82 (t, 2H, J =7.3 Hz, CH_2), 2.76 (m, 1H, CH-lac), 2.28 (t, 2H, J =7.7 Hz, CH_2), 2.10 (m, 3H, $\text{CH}_2+\text{CH-lac}$); ^{13}C NMR (75 MHz, CDCl_3) δ =122.2, 121.9, 119.5, 119.1, 115.6, 111.3, 66.2, 49.4, 35.7, 30.7, 25.8, 24.6; GC-MS: expected m/z=286, observed [M+]=286; $[\alpha_D]=+14.2$ ($c=2.6$ mg/mL; CHCl_3); IR (cm^{-1}): 3684, 3620, 3480, 3426, 3020, 2977, 2401, 2362, 2254, 1780, 1674, 1603, 1514, 1217.

Example 19

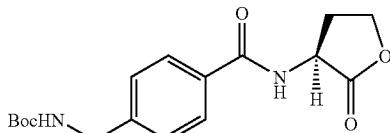
N-(indole-3-butanoyle)-D-homoserine lactone (7i)



N-(indole-3-butanoyle)-D-homoserine lactone (7i) was synthesized by the method described in Example 8. FIG. 10A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 10B is the NMR spectrum where: ^1H NMR (300 MHz, CDCl_3) δ =7.99 (s, 1H, NH), 7.61 (d, 1H, J =0.6 Hz, CH—Ar), 7.37 (d, 1H, J =0.4 Hz, CH—Ar), 7.26 (m, 2H, CH—Ar), 7.01 (d, 1H, J =2.1 Hz, CH—Ar), 5.90 (d, 1H, J =4.4 Hz, NH), 4.55 (ddd, 1H, J =5.9 Hz, CH-lac), 4.46 (td, 1H, J =8.3 Hz, CH-lac), 4.29 (ddd, 1H, J =5.9 Hz, CH-lac), 2.85 (m, 3H), 2.32 (m, 2H), 2.17 (m, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ =122.2, 121.8, 119.5, 111.3, 66.3, 49.4, 35.7, 30.7, 25.8, 24.6; GC-MS: expected m/z=286, observed [M+]=286; $[\alpha_D]=-11.6$ ($c=1.95$ mg/mL; CHCl_3); IR (cm^{-1}): 3359, 2921, 2692, 2358, 1771, 1648, 1558, 1220, 1019.

Example 20

N-Boc-(4-aminomethyl)-N-benzoyl-L-homoserine lactone (7j)



N-Boc-(4-aminomethyl)-N-benzoyl-L-homoserine lactone (7j) was synthesized by the method described in Example 8. FIG. 11 is the NMR spectrum where: ^1H NMR (300 MHz, CDCl_3) δ =7.78 (d, 2H, Ar—H), 7.35 (d, 2H, Ar—H), 6.02 (d, 1H, NH), 4.98 (s, 1H, NH), 4.79 (ddd, 1H, J =6.0 Hz, CH-lac), 4.57 (td, 1H, J =0.9 Hz, CH-lac), 4.40 (m, 3H, J =8.7 Hz, $\text{CH}_2+\text{CH-lac}$), 3.00 (dddd, 1H, J =1.2 Hz, CH-

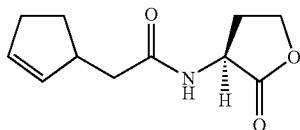
39

lac), 2.35 (ddd, 1H, J=8.8 Hz, CH-lac), 1.47 (s, 9H, BocCH₃); ¹³C NMR (75 MHz, CDCl₃) δ=131.7, 127.3, 66.1, 49.6, 30.4, 28.2; MS(ESI): expected m/z=334, observed [M+Na]=357; [α_D]=+21.6 (c=2.7 mg/mL; CHCl₃); IR (cm⁻¹): 3621, 3453, 3020, 2401, 1780, 1692, 1527, 1216.

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Example 21

N-(2-cyclopentene-1-acetanoyl)-L-homoserine lactone (7k)



N-(2-cyclopentene-1-acetanoyl)-L-homoserine lactone (7k) was synthesized by the method described in Example 8. FIG. 12A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 12B is the NMR spectrum where: ¹H NMR (300 MHz, CDCl₃) δ=6.01 (d, 1H, NH), 5.81 (m, 1H, CH-vinyl), 5.69 (m, 1H, CH-vinyl), 4.57 (ddd, 1H, J=3.0 Hz, CH-lac), 4.5 (td, 1H, J=9.5 Hz, CH-lac), 4.33 (ddd, 1H, J=6.0 Hz, CH-lac), 3.15 (m, 1H, J=2.6 Hz, CH), 2.91 (ddd, 1H, J=1.2 Hz, CH-lac) 2.39-2.09 (m, 6H), 1.52 (ddd, 1H, J=5.0 Hz, CH-lac); ¹³C NMR (75 MHz, CDCl₃) δ=175.6, 173.0, 133.7, 132.1, 66.3, 49.5, 42.6, 42.4, 32.1, 30.9, 29.8, 29.7; GC-MS: expected m/z=209, observed [M+]=209; [α_D]=+10.8 (c=2.6 mg/mL; CHCl₃); IR (cm⁻¹): 3311, 2923, 2851, 1775, 1643, 1546, 1173, 1016.

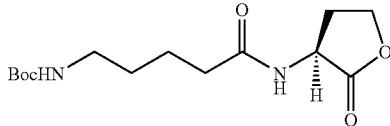
10

Example 21

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Example 23

N-Boc-aminocapranoyl-L-homoserine lactone (7m)

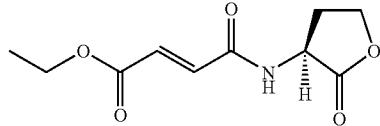


N-Boc-aminocapranoyl-L-homoserine lactone (7m) was synthesized by the method described in Example 8. FIG. 14 is the NMR spectrum where: ¹H NMR (300 MHz, CDCl₃) δ=6.50 (d, 1H, NH), 4.65 (m, 2H, CH-lac, NH), 4.49 (td, 1H, J=9.0 Hz, CH-lac), 4.32 (ddd, 1H, J=6.1 Hz, CH-lac), 3.11 (q, 2H, J=6.3 Hz, CH₂), 2.79 (ddd, 1H, J=4.5 Hz, CH-lac), 2.28 (t, 2H, J=7.5 Hz, CH₂), 2.17 (ddd, 1H, J=6.8 Hz, CH-lac) 1.69 (p, 2H, J=7.4 Hz, CH₂), 1.52 (m, 12H); ¹³C NMR (75 MHz, CDCl₃) δ=175.8, 173.7, 156.2, 49.2, 40.5, 36.0, 31.1, 30.3, 29.9, 28.6, 26.4, 25.1, 16.5; GC-MS: expected m/z=314, observed [M+Na]=337; [α_D]=+5.7 (c=2.1 mg/mL; CHCl₃); IR (cm⁻¹): 3684, 3455, 3020, 2401, 1781, 1693, 1511, 1216.

25

Example 24

N-monoethyl fumaroyl-L-homoserine lactone (7n)

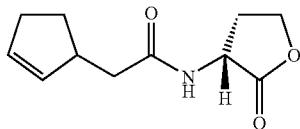


N-monoethyl fumaroyl-L-homoserine lactone (7n) was synthesized by the method described in Example 8. FIG. 15A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 15B is the NMR spectrum where: ¹H NMR (300 MHz, CDCl₃) δ=7.0 (d, 2H, Ar—H), 6.87 (d, 2H, Ar—H), 4.72 (ddd, 1H, J=6.3 Hz, CH-lac), 4.54 (td, 1H, J=1.3 Hz, CH-lac), 4.37 (ddd, 1H, J=6.0 Hz, CH-lac), 4.29 (q, 2H, J=7.1 Hz, CH₂), 2.91 (ddd, 1H, J=1.4 Hz, CH-lac), 2.31 (ddd, 1H, J=8.7 Hz, CH-lac) 1.47 (s, 2H, CH₂), 1.34 (t, 3H, J=7.1 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=175.2, 165.5, 164.4, 135.1, 131.8, 66.4, 61.6, 49.7, 30.4, 28.5, 14.3; GC-MS: expected m/z=227, observed [M+]=227; [α_D]=+4.7 (c=2.8 mg/mL; CHCl₃); IR (cm⁻¹): 3316, 2923, 1778, 1712, 1645, 1549, 1166.

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Example 22

N-(2-cyclopentene-1-acetanoyl)-D-homoserine lactone (7l)



N-(2-cyclopentene-1-acetanoyl)-D-homoserine lactone (7l) was synthesized by the method described in Example 8. FIG. 13A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 13B is the NMR spectrum where: ¹H NMR (300 MHz, CDCl₃) δ=6.07 (s, 1H, NH), 5.81-5.70 (m, 1H, CH-vinyl), 5.69-5.66 (m, 1H, CH-vinyl), 4.60 (ddd, 1H, J=2.9 Hz, CH-lac), 4.47 (td, 1H, J=8.3 Hz, CH-lac), 4.33 (ddd, 1H, J=6.0 Hz, CH-lac), 3.15-3.10 (m, 1H), 2.91 (ddd, 1H, J=1.3 Hz, CH-lac) 2.40-2.08 (m, 6H), 1.54-1.43 (m, 1H, CH-lac); ¹³C NMR (75 MHz, CDCl₃) δ=173.0, 033.8, 133.7, 132.1, 132.0, 66.3, 49.5, 42.6, 42.4, 32.0, 30.8, 29.7; GC-MS: expected m/z=209, observed [M+]=209; [α_D]=-12.9 (c=2.7 mg/mL; CHCl₃); IR (cm⁻¹): 3310, 2923, 2851, 2385, 1775, 1643, 1546, 1173, 1016.

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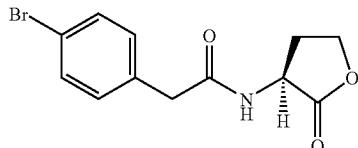
40

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Example 25

N-(4-bromophenylacetanoyl)-L-homoserine lactone (7o)



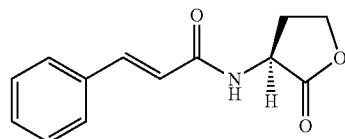
41

N-(4-bromophenylacetanoyl)-L-homoserine lactone (7o) was synthesized by the method described in Example 8. FIG. 16A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 16B is the NMR spectrum where: ^1H NMR (300 MHz, CDCl_3) δ =7.49 (d, 2H, $J=2.7$ Hz, CH—Ar), 7.17 (d, 2H, $J=2.7$ Hz, CH—Ar), 6.02 (s, 1H, NH), 4.58 (ddd, 1H, $J=6.2$ Hz, CH-lac), 4.45 (td, 1H, $J=1.2$ Hz, CH-lac), 4.33 (ddd, 1H, $J=5.9$ Hz, CH-lac), 3.59 (s, 2H, CH_2), 2.87 (ddd, 2H, $J=1.4$ Hz, CH-lac), 2.19 (dd, 1H, $J=8.9$ Hz, CH-lac); ^{13}C NMR (75 MHz, CDCl_3) δ =132.8, 131.9, 130.9, 65.8, 49.2, 30.1; GC-MS: expected m/z=297, observed [M+]=297; $[\alpha_D]=+3.8$ ($c=2.55$ mg/mL; CHCl_3); IR (cm^{-1}): 3419, 3020, 2401, 1782, 1672, 1216.

Example 26

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N-(trans-cinamoyl)-L-homoserine lactone (7p)

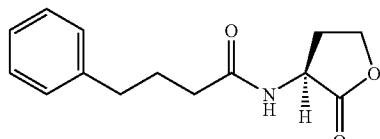


N-(trans-cinamoyl)-L-homoserine lactone (7p) was synthesized by the method described in EXAMPLE 8. FIG. 17A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 17B is the NMR spectrum: ^1H NMR (300 MHz, CDCl_3) δ =7.66 (d, 2H, Ar—H), 7.48 (m, 2H, Ar—H), 7.36 (m, 3H, Ar—H), 6.51 (d, 1H, $J=5.9$ Hz, NH), 6.49 (d, 1H, $J=15.7$ Hz, H-vinyl), 4.78 (ddd, 1H, $J=6.4$ Hz, CH-lac), 4.53 (td, 1H, $J=8.9$ Hz, CH-lac), 4.37 (ddd, 1H, $J=5.9$ Hz, CH-lac) 2.92 (ddd, 1H, $J=1.0$ Hz, CH-lac), 2.32 (td, 1H, $J=8.9$ Hz, CH-lac); ^{13}C NMR (75 MHz, CDCl_3) δ =176.0, 166.6, 142.6, 134.6, 130.2, 129.1, 128.2, 119.5, 66.5, 49.7, 30.7; GC-MS: expected m/z=231, observed [M+]=231; $[\alpha_D]=+30.2$ ($c=2.45$ mg/mL; CHCl_3); IR (cm^{-1}): 3312, 2923, 1774, 1644, 1547, 1173, 1016.

Example 27

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N-(4-phenylbutanoyl)-L-homoserine lactone (7q)



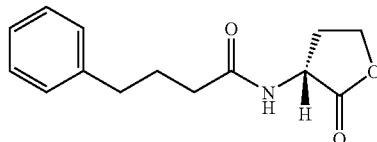
N-(4-phenylbutanoyl)-L-homoserine lactone (7q) was synthesized by the method described in Example 8. FIG. 18A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 18B is the NMR spectrum where: ^1H NMR (300 MHz, CDCl_3) δ =7.31 (m, 2H, Ar—H), 7.21 (m, 3H, Ar—H), 6.13 (d, 1H, $J=5.3$ Hz, NH), 4.59 (ddd, 1H, $J=6.2$ Hz, CH-lac), 4.47 (td, 1H, $J=1.0$ Hz, CH-lac), 4.30 (ddd, 1H, $J=5.7$ Hz, CH-lac), 2.85 (ddd, 1H, $J=1.1$ Hz, CH-lac), 2.68 (t, 2H, $J=7.3$ Hz, CH_2), 2.36 (t, 2H, $J=6.7$ Hz, CH_2), 2.18 (d, 1H, $J=3.2$ Hz, CH-lac), 2.04 (p, 2H, $J=7.4$ Hz, CH_2); ^{13}C NMR (75 MHz, CDCl_3) δ =175.7, 173.5, 167.7, 141.5, 128.7, 128.6, 126.2, 115.0, 66.3, 49.4, 35.5,

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35.3, 30.7, 27.0; GC-MS: expected m/z=247, observed [M+]=247; $[\alpha_D]=+27.9$ ($c=6.7$ mg/mL; CHCl_3); IR (cm^{-1}): 3314, 2935, 2358, 2331, 1771, 1652, 1447, 1173.

Example 28

N-(4-phenylbutanoyl)-D-homoserine lactone (7r)

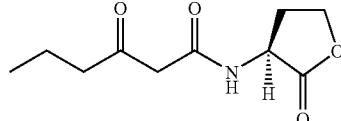


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N-(4-phenylbutanoyl)-D-homoserine lactone (7r) was synthesized by the method described in Example 8. FIG. 19A is the GC-MS spectrum assaying the purity of the synthesis (inset shows the chemical structure). FIG. 19B is the NMR spectrum where: ^1H NMR (300 MHz, CDCl_3) δ =7.31 (m, 2H, Ar—H), 7.21 (m, 3H, Ar—H), 6.10 (d, 1H, $J=4.7$ Hz, NH), 4.59 (ddd, 1H, $J=5.9$ Hz, CH-lac), 4.47 (td, 1H, $J=1.2$ Hz, CH-lac), 4.31 (ddd, 1H, $J=5.8$ Hz, CH-lac), 2.86 (ddd, 1H, $J=1.2$ Hz, CH-lac), 2.69 (t, 2H, $J=7.2$ Hz, CH_2), 2.36 (t, 2H, $J=7.8$ Hz, CH_2), 2.18 (ddd, 1H, $J=3.2$ Hz, CH-lac), 2.04 (p, 2H, $J=7.1$ Hz, CH_2); ^{13}C NMR (75 MHz, CDCl_3) δ =175.6, 173.5, 167.6, 163.9, 141.5, 128.7, 128.6, 126.2, 115.1, 115.0, 66.3, 59.4, 49.4, 35.5, 35.3, 30.7, 27.0; GC-MS: expected m/z=247, observed [M+]=247; $[\alpha_D]=-24.1$ ($c=4.0$ mg/mL; CHCl_3); IR (cm^{-1}): 3309, 2966, 2685, 23559, 2338, 1772, 1669, 1539, 1556, 1221, 1201.

Example 29

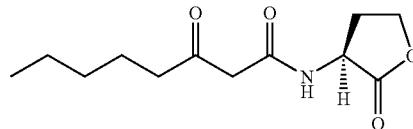
N-(3-oxo-hexanoyl)-L-homoserine lactone (8a)



N-(3-oxo-hexanoyl)-L-homoserine lactone (8a) was synthesized by the method described in Example 9. FIG. 20 is the NMR spectrum (inset shows the chemical structure) where: ^1H NMR (300 MHz, CDCl_3) δ =7.85 (s, 1H, NH), 4.67 (ddd, 1H, $J=6.7$ Hz, CH-lac), 4.57 (td, 1H, $J=1.3$ Hz, CH-lac), 4.34 (ddd, 1H, $J=6.1$ Hz, CH-lac), 3.52 (s, 2H, CH_2), 2.82 (ddd, 1H, $J=1.2$ Hz, CH-lac), 2.56 (t, 2H, $J=7.2$ Hz, CH_2), 2.33 (ddd, 1H, $J=2.4$ Hz, CH-lac), 1.64 (p, 2H, $J=7.2$ Hz, CH_2), 0.98 (t, 3H, $J=6.5$ Hz, CH_3); ^{13}C NMR (75 MHz, CDCl_3) δ =204.7, 173.1, 164.9, 64.2, 47.4, 46.3, 44.0, 28.1, 15.1, 11.8; MS(ESI): expected m/z=213, observed [M+Na]=236; $[\alpha_D]=+12.2$ ($c=2.7$ mg/mL; CHCl_3); IR (cm^{-1}): 3286, 3966, 1783, 1718, 1646, 1545, 1171.

Example 30

N-(3-oxo-octanoyl)-L-homoserine lactone (8b)

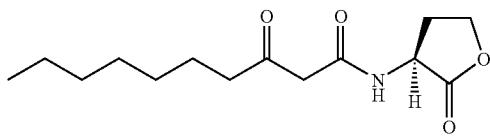


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N-(3-oxo-octanoyl)-L-homoserine lactone (8b) was synthesized by the method described in Example 9. FIG. 21 is the NMR spectrum (inset shows the chemical structure) where: ¹H NMR (300 MHz, CDCl₃) δ=7.95 (s, 1H, NH), 4.67 (ddd, 1H, J=6.7 Hz, CH-lac), 4.53 (td, 1H, J=1.3 Hz, CH-lac), 4.34 (ddd, 1H, J=6.1 Hz, CH-lac), 3.52 (s, 2H, CH₂), 2.81 (dd, 1H, J=1.2 Hz, CH-lac), 2.57 (t, 2H, J=7.2 Hz, CH₂) 2.34 (ddd, 1H, J=2.4 Hz, CH-lac), 1.64 (p, 2H, J=7.2 Hz, CH₂), 1.43 (m, 4H, J=6.4 Hz, (CH₂)₄), 0.91 (t, 3H, J=6.5 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=206.7, 175.0, 166.7, 66.3, 66.1, 49.3, 48.4, 44.0, 31.3, 30.0, 23.2, 22.6, 14.1; MS(ESI): expected m/z=241, observed [M+Na]=264; [α_D]=+16.9 (c=2.7 mg/mL; CHCl₃); IR (cm⁻¹): 3684, 3020, 2401, 1783, 1712, 1674, 1527, 1216.

Example 31

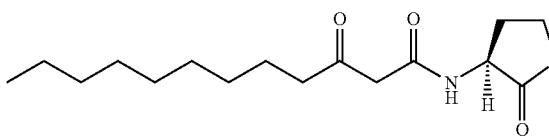
N-(3-oxo-decanoyl)-L-homoserine lactone (8c)



N-(3-oxo-decanoyl)-L-homoserine lactone (8c) was synthesized by the method described in Example 9. FIG. 22 shows the NMR spectrum (inset shows the chemical structure) where: ¹H NMR (300 MHz, CDCl₃) 7.68 (d, 1H, J=4.9 Hz, NH), 4.65 (ddd, 1H, J=9.2 Hz, CH-lac), 4.53 (t, 1H, J=8.9 Hz, CH-lac), 4.32 (ddd, 1H, J=6.2 Hz, CH-lac), 3.47 (s, 2H, CH₂), 2.79 (ddd, 1H, J=1.3 Hz, CH-lac), 2.55 (t, 2H, J=7.3 Hz, CH₂), 2.32 (ddd, 1H, J=2.4 Hz, CH-lac), 1.61 (m, 2H, CH₂), 1.28 (m, 8H, (CH₂)₄), 0.89 (t, 3H, J=5.4 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=206.7, 175.1, 166.6, 66.3, 66.1, 49.2, 48.4, 44.1, 31.8, 29.9, 29.2, 29.1, 23.6, 22.8, 14.2; MS(ESI): expected m/z=269, observed [M+Na]=292; [α_D]=+16.7 (c=3.4 mg/mL; CHCl₃); IR (cm⁻¹): 3295, 2923, 2851, 1775, 1716, 1645, 1547, 1176.

Example 32

N-(3-oxo-dodecanoyl)-L-homoserine lactone (8d)



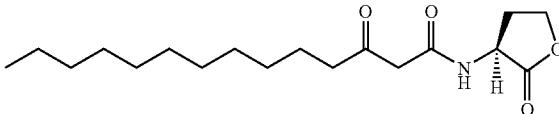
N-(3-oxo-dodecanoyl)-L-homoserine lactone (8d) was synthesized by the method described in Example 9. FIG. 23 is the NMR spectrum (inset shows the chemical structure) where: ¹H NMR (300 MHz, CDCl₃) 7.68 (d, 1H, J=6.0 Hz, NH), 4.65 (ddd, 1H, J=5.9 Hz, CH-lac), 4.50 (td, 1H, J=1.1 Hz, CH₂), 4.32 (ddd, 1H, J=6.1 Hz, CH-lac), 3.47 (s, 2H, CH₂), 2.79 (ddd, 1H, J=7.0 Hz, CH-lac), 2.55 (t, 2H, J=7.3 Hz, CH₂), 2.29 (ddd, 1H, J=2.2 Hz, CH-lac), 1.60 (m, 2H, CH₂), 1.26 (m, 12H, (CH₂)₆), 0.89 (t, 3H, J=6.1 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=206.7, 175.1, 174.1, 166.7, 66.3, 66.1, 49.4, 49.2, 48.9, 48.4, 44.1, 36.4, 32.0, 30.7, 29.9, 29.6, 29.5, 29.4, 29.2, 25.7; MS(ESI): expected

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m/z=297, observed [M+Na]=320; [α_D]=+11.8 (c=3.6 mg/mL; CHCl₃); IR (cm⁻¹): 3295, 2922, 2851, 1775, 1716, 1644, 1547, 1171, 1016.

Example 33

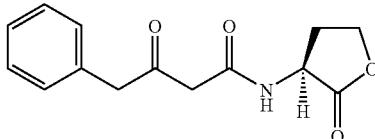
N-(3-oxo-tetradecanoyl)-L-homoserine lactone (8e)



N-(3-oxo-tetradecanoyl)-L-homoserine lactone (8e) was synthesized by the method described in Example 9. FIG. 24 is the NMR spectrum (inset shows the chemical structure) where: ¹H NMR (300 MHz, CDCl₃) 7.68 (d, 1H, J=6.0 Hz, NH), 4.66 (ddd, 1H, J=5.9 Hz, CH-lac), 4.56 (td, 1H, J=1.1 Hz, CH₂), 4.34 (ddd, 1H, J=6.1 Hz, CH-lac), 3.50 (s, 2H, CH₂), 2.86 (ddd, 1H, J=7.0 Hz, CH-lac), 2.30 (t, 2H, J=7.3 Hz, CH₂), 2.28 (ddd, 1H, J=2.2 Hz, CH-lac), 1.60 (m, 2H, CH₂), 1.26 (m, 12H, (CH₂)₆), 0.89 (t, 3H, J=6.1 Hz, CH₃); ¹³C NMR (75 MHz, CDCl₃) δ=175.0, 166.7, 66.3, 66.0, 49.5, 36.1, 32.1, 30.7, 29.8, 29.6, 29.5, 29.5, 29.4, 29.2, 25.7; MS(ESI): expected m/z=325, observed [M+Na]=348; [α_D]=+14.0 (c=2.5 mg/mL; CHCl₃); IR (cm⁻¹): 3921, 3315, 2852, 1777, 1716, 1645, 1548, 1174, 1014.

Example 34

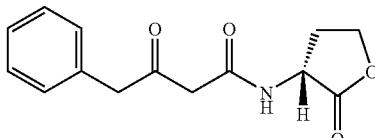
N-(3-oxo-4-phenylbutanoyl)-L-homoserine lactone (8f)



N-(3-oxo-4-phenylbutanoyl)-L-homoserine lactone (8f) was synthesized by the method described in EXAMPLE 9. FIG. 25 is the NMR spectrum (inset shows the chemical structure) where: ¹H NMR (300 MHz, CDCl₃) δ=7.66 (s, 1H, NH), 7.64 (m, 3H, CH—Ar), 7.20 (d, 2H, CH—Ar), 4.60 (ddd, 1H, J=2.0 Hz, CH-lac), 4.47 (td, 1H, J=1.4 Hz, CH-lac), 4.28 (ddd, 1H, J=3.2 Hz, CH-lac), 3.80 (s, 2H, CH₂), 3.53 (s, 2H, CH₂), 2.70 (ddd, 1H, J=1.1 Hz, CH-lac), 2.23 (ddd, 1H, J=8.9 Hz, CH-lac); ¹³C NMR (75 MHz, CDCl₃) δ=204.0, 175.1, 167.0, 132.9, 129.8, 129.6, 129.2, 127.8, 66.2, 50.8, 49.3, 47.6, 15.29.7; MS(ESI): expected m/z=261, observed [M+Na]=284; [α_D]=+14.6 (c=3.4 mg/mL; CHCl₃); IR (cm⁻¹): 3335, 3054, 2916, 1176, 1772, 1663, 1538, 1179, 1022.

Example 35

N-(3-oxo-4-phenylbutanoyl)-D-homoserine lactone (8g)

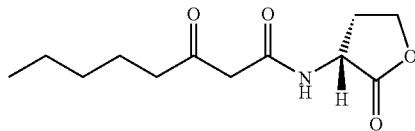


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N-(3-oxo-4-phenylbutanoyl)-D-homoserine lactone (8g) was synthesized by the method described in EXAMPLE 9. FIG. 26 is NMR spectrum (inset shows the chemical structure) where: ^1H NMR (300 MHz, CDCl_3) δ =7.70 (s, 1H, NH), 7.69 (m, 3H, CH—Ar), 7.21 (d, 2H, CH—Ar), 4.59 (ddd, 1H, $J=2.0$ Hz, CH-lac), 4.49 (td, 1H, $J=1.2$ Hz, CH-lac), 4.30 (ddd, 1H, $J=2.1$ Hz, CH-lac), 3.80 (s, 2H, CH_2), 3.53 (s, 2H, CH_2), 2.74 (ddd, 1H, $J=1.0$ Hz, CH-lac), 2.23 (ddd, 1H, $J=9.0$ Hz, CH-lac); ^{13}C NMR (75 MHz, CDCl_3) δ =204.1, 174.9, 167.1, 132.7, 129.7, 129.6, 129.2, 128.8, 127.8, 66.2, 50.9, 49.4, 47.3, 29.8; MS(ESI): expected m/z=261, observed [M+Na]=284; $[\alpha_D]=-6.2$ ($c=1.5$ mg/mL; CHCl_3); IR (cm^{-1}): 3335, 3054, 2916, 1176, 1772, 1663, 1538, 1179, 1022.

Example 36

N-(3-oxo-octanoyl)-D-homoserine lactone (8h)



N-(3-oxo-octanoyl)-D-homoserine lactone (8h) was synthesized by the method described in Example 9. FIG. 27 shows the NMR spectrum (inset shows the chemical structure) where: ^1H NMR (300 MHz, CDCl_3) δ =7.95 (s, 1H, NH), 4.67 (ddd, 1H, $J=6.7$ Hz, CH-lac), 4.53 (td, 1H, $J=1.3$ Hz, CH-lac), 4.34 (ddd, 1H, $J=6.1$ Hz, CH-lac), 3.52 (s, 2H, CH_2), 2.81 (dd, 1H, $J=1.2$ Hz, CH-lac), 2.57 (t, 2H, $J=7.2$ Hz, CH_2), 2.34 (ddd, 1H, $J=2.4$ Hz, CH-lac), 1.64 (p, 2H, $J=7.2$ Hz, CH_2), 1.43 (m, 4H, $J=6.4$ Hz, $(\text{CH}_2)_4$), 0.91 (t, 3H, $J=6.5$ Hz, CH₃); ^{13}C NMR (75 MHz, CDCl_3) δ =206.7, 175.0, 166.7, 66.3, 66.1, 49.3, 48.4, 44.0, 31.3, 30.0, 23.2, 22.6, 14.1; MS(ESI): expected m/z=241, observed [M+Na]=264; $[\alpha_D]=-19.6$ ($c=2.7$ mg/mL; CHCl_3); IR (cm^{-1}): 3684, 3020, 2401, 1783, 1712, 1674, 1527, 1216.

Example 37

Evaluation of AHL Mosher Amides

The enantiopurity of the AHLs synthesized via SCHEME I was assessed through the characterization of AHL Mosher amide derivatives. This solid-phase route was used to synthesize R-Mosher amide diastereomers starting from both the L- and D-methionine loaded resins (3). Analysis of the ^1H NMR (300 MHz, CDCl_3) spectrum of the diastereomers indicated that the AHL de was >95% (as calculated from 100(a-a') or 100(b-b') where a, a', b, b' are the fractional yields based on the integration of respective peaks in the NMR spectrum). FIGS. 28A and 28B show the ^1H NMR spectrum of the D and L homoserine enantiomers, respectively.

Biological Screenings

Compound Handling & Reagents: Stock solutions of synthetic compounds (10 mM and 100 mM) were prepared in either ethyl acetate or CHCl_3 and stored at -20° C. in sealed vials. The solutions were allowed to come to room temperature prior to use in assays. Solvent resistant polypropylene (Corning Costar cat. no. 3790) or polystyrene (Corning Costar cat. no. 3997) 96-well plates were used when appropriate.

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All biological reagents were purchased from Fisher and used according to enclosed instructions. AB Minimal media was prepared containing 1.0 g/L NH_4Cl , 0.3 g/L $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 0.15 g/L KCl, 0.01 g/L CaCl_2 , 2.5 mg/L $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$, 5.0 g/L glucose, 1.0 g/L NaH_2PO_4 , and 3.0 g/L K_2HPO_4 , pH=6.8. M9 media used in biofilm assays was prepared as described (De Kievit, T. R.; Gillis, R.; Marx, S.; Brown, C.; Iglesias, B. H. *Appl. Environ. Microbiol.* 2001, 67, 1865-1873). Buffers and solutions for Miller absorbance assays were prepared as described (Miller, J. H. *Experiments in Molecular Genetics*; Cold Spring, 1972).

Control Compounds: Compounds 7g (Zhu, J.; Beaber, J. W.; More, M. I.; Fuqua, C.; Eberhard, A.; Winans, S. C. *J. Bacteriol.* 1998, 180, 5398-5405) and 8h (Reverchon, S.; Chantegrel, B.; Deshayes, C.; Doutteau, A.; Cotte-Pattat, N. *Bioorg. Med. Chem. Lett.* 2002, 12, 1153-1157) were prepared according to the solid-phase methods described above. Compound 9 was synthesized using a modified solution-phase procedure (Smith, K. M.; Bu, Y.; Suga, H. *Chem. Biol.* 2003, 10, 563-571) as follows: β -keto acid 5d (483 mg, 1.87 mmol) and 2-aminophenol (204 mg, 1.87 mmol) were dissolved in 10 mL DMF, after which 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDC, 456 mg, 2.38 mmol), 4-dimethylaminopyridine (DMAP, 228 mg, 1.87 mmol), and N,N-diisopropylethylamine (DIPEA, 500 μL , 2.87 mmol) were added. The reaction mixture was stirred at room temperature until TLC indicated completion (12 h). The reaction mixture was diluted with 30 mL of diethyl ether and washed 2x with 30 mL 10% citric acid, 2x30 mL saturated NaHCO_3 , and 1x15 mL saturated NaCl. The ether layer was separated, dried with magnesium sulfate, and the solvent removed in vacuo. The resulting solid was immediately subjected to 95% $\text{TFA}/\text{H}_2\text{O}$ (30 minutes) to affect ketal deprotection. Extraction of the reaction mixture (3x15 mL) with CHCl_3 , drying over MgSO_4 , and removal of CHCl_3 in vacuo gave a yellow-brown solid. Purification by flash silica gel chromatography (1:1 EtOAc/hexane) yielded 9 as a white solid. 228 mg, 40% overall yield. Compounds 7g, 8h, and 9 were stored at -20° C. until required for use.

Instrumentation: Absorbance and fluorescence assay results were obtained using a PerkinElmer Wallac 2100 EnVision™ (PerkinElmer, Waltham, Mass.) multilabel plate reader using Wallac Manager v1.03 software. A filter of 600 nm was used for reading bacterial cell density. Filters of 420 nm and 550 nm were used for Miller-type absorbance assays (Griffith, K. L.; Wolf, R. E., Jr. *Biochem. Biophys. Res. Commun.* 2002, 290, 397-402). Filters of 485 nm for excitation and 535 nm for emission were used for evaluating the production of green fluorescent protein (GFP) in fluorescence assays. Biofilms were visualized with a Bio-Rad MRC-1024 laser scanning confocal microscope using Lasersharp v3.2 software. Biofilm visualization data was manipulated using Confocal Assistant v4.02 software (Available to the public from nephrology.iupui.edu/imaging/software.htm).

Example 38

Screening of Synthetic AHL Library

The AHL library constructed by the schema described above, was screened in two bacterial reporter strains for antagonism of quorum sensing: 1) *P. aeruginosa* PAO-JP2 (plasmid pLVAgfp), (De Kievit, T. R.; Gillis, R.; Marx, S.; Brown, C.; Iglesias, B. H. *Appl. Environ. Microbiol.* 2001, 67, 1865-1873) and 2) *A. tumefaciens* WCF47 (pCF372). Examination of these two strains was valuable for two reasons: 1) the direct clinical relevance of *P. aeruginosa*, and 2), the extensive body

of biochemical and structural data for TraR in *A. tumefaciens*. Both of these reporter strains lack their native AHL synthases, yet retain active LuxR-type receptors (LasR and TraR proteins, respectively); exogenous ligand is required for receptor activation, which can be measured by fluorescence (green fluorescent protein (GFP) for LasR) or absorbance (via β -galactosidase activity for TraR) measurements.

The antagonism screens revealed a suite of new quorum sensing inhibitors. In these experiments, illustrated in FIG. 29, the strains were treated with non-native AHL in the presence of native AHL ligand (8b or 8d), and a reduction in absorbance or fluorescence signal indicated that the non-native AHL was able to antagonize LuxR-type protein activity. Three compounds (7h, 7k and 7o) showed significant activity against TraR in *A. tumefaciens* and were one to two orders of magnitude more active than the previously reported LuxR-type protein antagonists examined as controls (7g, 8f, and 9 at 10 μ M in FIGS. 29a & 29c). Impressively, bromophenyl AHL 7o displayed 50% inhibition at an equimolar concentration of 8b (100 nM). Interestingly, the same three ligands were also identified as antagonists against LasR in *P. aeruginosa* (FIG. 29d). Here indol AHL 7h and bromophenyl AHL 7o were twice as active as the three controls (at 400 μ M), with indol AHL 7h displaying 50% inhibition at a 12.5:1 ratio with native ligand 8d. Notably, all three ligands contain bulky, hydrophobic acyl groups. This structural similarity, coupled with their cross activity, suggests that the ligands could be interacting with the TraR and LasR receptors in analogous manners; efforts to characterize these interactions are currently underway.

Further, the cross reactivity of ligands 7h, 7k and 7o illustrated that methods illustrated to construct the AHL combinatorial library are sufficient to create synthetic ligands for use in screening all bacteria having AHL-like ligand-receptors. Further, characterization of the synthetic ligands was made using reporter gene assays to determine agonism/antagonism effects of the non-natural ligands compared to naturally occurring ligands.

Investigation of Agonism/Antagonism Effects

Agonism/antagonism studies were performed for the synthetic analogs identified in Table 2 in both *A. tumefaciens* and *P. aeruginosa* models. The results of these experiments, illustrated in FIGS. 30 and 31, show not only the power of the reporter assay screen, but also the cross reactivity of the respective ligands in different hosts across the genera of quorum-sensing bacteria. Further, antagonism assays shown in FIGS. 32 and 33 illustrated the effectiveness of the AHL analogs across a concentration ranges and indicate that varying the concentration of synthetic ligands can result in an intermediate response, such that at high concentrations there is virtually no signal.

Example 39

Agrobacterium tumefaciens Reporter Gene Assay

An appropriate amount of concentrated AHL stock solution, to give a final concentration of 100 nM, was added to an empty culture tube, and the solvent was allowed to evaporate. An overnight culture of *A. tumefaciens* WCF47(pCF372) was diluted to an OD₆₀₀ (optical density) of 0.1 in fresh AB minimal medium containing 400 μ g/mL octopine and 50 μ g/mL streptomycin. A 1 mL portion of the diluted culture was added to the tubes containing AHLs. Tubes were grown at 28° C. for 12-16 h in a rotary shaking incubator (200 rpm).

The cultures were then assayed for β -galactosidase activity following the Miller assay method. Briefly, 100 μ L aliquots of bacteria from each of the tubes were added to wells of a polystyrene 96-well plate and the OD₆₀₀ of each well was recorded. Next, 50 μ L aliquots from each well were transferred to a solvent resistant 96-well plate containing 200 μ L Z buffer, 8 μ L CHCl₃ and 4 μ L, 0.1% aq. sodium dodecylsulfate (SDS). This suspension was mixed via repetitive pipetting, after which the CHCl₃ was allowed to settle. A 100 μ L aliquot from each well was transferred to a fresh polystyrene 96-well plate, and 20 μ L of substrate, o-nitrophenyl- β -D-galactopyranoside (ONPG, 4 μ g/mL in phosphate buffer), was added at time zero. After the development of appropriate yellow color (ca. 15-35 minutes), the reaction was terminated by the addition of 50 μ L of 1 M Na₂CO₃. Absorbance at 420 nm and 550 nm was measured for each well using a plate reader, and Miller units were calculated according to standard methods.

Similar methods were used for antagonism assays, except the concentration of AHL analog used was 10⁴ nM, and OOHL (8b) stock solution was added to each tube such that its concentration was 100 nM (FIG. 30). Compounds that showed good inhibitory activity at 10⁴ nM were then tested through a range of concentrations against OHHL (8b) at 100 nM, from 10⁴ nM to 1 nM (shown in FIG. 32). All assays were performed in triplicate.

FIG. 30, shows the results for the *A. tumefaciens* reporter gene screening data for AHL derivatives 7h-7r and 8f-8h. Data for control compounds 7g, 8f, and 9 are shown. Reporter strain: WCF47(pCF372). Miller units report relative β -galactosidase activity with ONPG as colorimetric substrate. Compound concentration in agonism assays: 100 nM. Compound concentration in antagonism assays: 10⁴ nM against 100 nM native ligand 8b (OOHL). Error bars are \pm one S.E.M., calculated from three replicate screens.

FIG. 31 shows antagonism screening data for AHL derivatives 7k and 7o in an *A. tumefaciens* reporter strain over a concentration range. Reporter strain: WCF47(pCF372). Miller units report relative β -galactosidase activity with ONPG as colorimetric substrate. Compounds screened at various concentrations against 100 nM native ligand 8b (OOHL). FIGS. 31A-31F are graphical displays of the results of dose response antagonism screening data for AHL derivatives 7h (FIG. 31A), 7k (FIG. 31B) and 7o (FIG. 31C) and control compounds 7g (FIG. 31D), 8f (FIG. 31E) and 9 (FIG. 31F) in an *A. tumefaciens* reporter strain. Reporter strain: WCF47(pCF372). Miller units report relative beta-galactosidase activity with ONPG as colometric substrate. Compounds screened at various concentrations against 100 nM native ligand 8b (OOHL). Error bars are \pm one S.E.M., calculated from at least three replicate screens.

Example 40

Pseudomonas aeruginosa Reporter Gene Assays

An appropriate amount of concentrated stock solution, to give a final concentration of 1 μ M, was added to a polypropylene 96-well plate, and the solvent was allowed to evaporate. An overnight culture of *P. aeruginosa* PAO-JP2(plasmid-LVAgfp) was diluted to an OD₆₀₀ of 0.1 in fresh LB medium containing 200 μ g/mL carbenicillin. 200 μ L of the diluted culture was added to each well of the plate and incubated at 37° C. for 6 h in a rotary shaking incubator (200 rpm). Cultures were transferred to a polystyrene 96-well plate, GFP expression was measured using a plate reader, and this value was normalized to cell density. Similar methods were used for antagonism assays, illustrated in FIG. 32, except the concentration of AHL analog used was 400 μ M and ODHL (8d) stock solution was added to each well such that its concen-

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tration was 1 μM . Compounds that showed good inhibitory activity at 400 μM were then tested through a range of concentrations against ODHL (8d) at 1 μM , from 400 μM to 4 nM (shown in FIG. 33). All assays were performed in triplicate.

FIG. 32, *P. aeruginosa* reporter gene screening data for AHL derivatives 7h-7r and 8f-8h. Data for control compounds 7g, 8f, and 9 are shown. Reporter strain: PAO-JP2 (plasI-LVAgfp). Fluorescence reported as percentage relative to control (8d, ODHL). Compound concentration in agonism assays: 1 μM . Compound concentration in antagonism assays: 400 μM against 1 μM native ligand 8d (ODHL). Error bars are \pm one S.E.M., calculated from three replicate screens.

FIGS. 33A-33F are graphical displays of the results of dose response antagonism screening data for AHL derivatives 7h (FIG. 33A), 7k (FIG. 33B) and 7o (FIG. 33C) and control compounds 7g (FIG. 33D), 8f (FIG. 33E) and 9 (FIG. 33F) in an *P. aeruginosa* reporter strain. Reporter strain: PAO-JP2 (plasI-LVAgfp). Fluorescence reported as percentage relative to native ligand 8d (ODHL). Compounds screened at various concentrations against 1 μM native ligand 8d (ODHL). Error bars are \pm one S.E.M., calculated from at least three replicate screens.

The data presented in FIGS. 31A-31F and FIGS. 33A-33F were fit to sigmoid dose-response curves to calculate IC₅₀ values, which are presented in Table 4, below.

TABLE 4

Selected antagonism screening data and IC ₅₀ inhibition values for AHL derivatives 7h, 7k and 7o and control compounds 7g, 8f, and 9 in <i>A. tumefaciens</i> and <i>P. aeruginosa</i> reporter strains. ^a						
compound	<i>A. tumefaciens</i>			<i>P. aeruginosa</i>		
	β -galactosidase activity [Miller units]	IC50 (μM) ^{c,d}	400 μM ^e	Relative fluorescence [%] ^b	12.5 μM ^e	IC50 (μM) ^{d,e}
8b: OOHL	— ^f	385 \pm 35 ^g	—	—	—	—
8d: ODHL	—	—	—	100 \pm 8 ^h	—	—
7h	150 \pm 14	245 \pm 14	1.1	14 \pm 4	52 \pm 8	14.8
7k	19 \pm 8	306 \pm 8	1.0	24 \pm 7	72 \pm 8	33.1
7o	0 \pm 4	193 \pm 9	0.25	14 \pm 5	58 \pm 21	16.1
7g	34 \pm 6	226 \pm 30	0.13	29 \pm 9	78 \pm 9	51.5
8f	51 \pm 22	310 \pm 6	0.59	27 \pm 9	82 \pm 18	53.6
9	186 \pm 15	396 \pm 27	4.7	23 \pm 7	63 \pm 8	37.3

^a *A. tumefaciens* strain: WCF47(pCF372); *P. aeruginosa* strain: PAO-JP2(plasI-LVAgfp).

^b Relative to agonism by 8d at 1 μM ; Set to 100%.

^c Screened against 100 nM 8b.

^d IC50 calculated from dose response data shown in FIG. S-5 and S-6 using nonlinear regression sigmoidal dose-response curve fit at 95% confidence with GraphPad Prism software (v. 4.00).

^e Screened against 1 μM 8d.

^f Not applicable.

^g Control: Agonism activity at 100 nM 8b.

^h Control: Relative agonism activity at 1 μM 8d (note: not at 12.5 μM).

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Standard static biofilm assays were performed using a *P. aeruginosa* (PAO1(pLVAgfp)) strain that constitutively produces GFP to facilitate visualization. Biofilms were grown in the absence of ligand (FIG. 34A) and in the presence of 50 μM synthetic ligands 7h (FIG. 34B) or 7o (FIG. 34C), for 48 h and visualized using scanning laser confocal microscopy. The treated biofilms appeared significantly less fluorescent relative to the untreated control, which has been used previously to establish that biofilms have reduced cell densities and are weakly organized. These data indicate that both non-natural quorum sensing compounds 7h and 7o strongly inhibit *P. aeruginosa* biofilm formation.

Example 42

Disruption of Biofilm Formation

Standard static biofilm assays were performed using a *P. aeruginosa* PAO1(pTdK-GFP). In brief, an overnight culture was diluted to an OD₆₀₀ of 0.1 in fresh M9 medium containing 200 $\mu\text{g}/\text{mL}$ carbenicillin. This diluted culture was added to a tube containing 50 μM antagonist. Biofilms were grown in the absence of ligand, FIG. 35A; in the presence synthetic ligand 7h, FIG. 35B; and in the presence of synthetic ligand

Example 41

Disruption of Biofilm Formation

As biofilm formation is largely under the control of LasR in *P. aeruginosa*, the inventors hypothesized that antagonists 7h and 7o could disrupt *P. aeruginosa* biofilm formation. Assays were performed according to standard procedures reported by Iglewski and co-workers.

7o, FIG. 35C (both at 50 μM) for 48 h and visualized using scanning laser confocal microscopy. A sterilized glass coverslip was added to each tube, and the culture was incubated at 37° C. for 48 h without shaking. Coverslips were removed, washed with phosphate buffered saline, and examined using scanning laser confocal microscopy methods. Top-down Z series were collected over a distance of ca. 100 μm . Images in FIGS. 35-37 are representative of several experiments performed on separate days.

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Example 43

Assay of Dose-Dependent Disruption of *P. aeruginosa* Biofilm: Compound 7o

The dose-dependent relationship of synthetic quorum sensing compound 7o was explored using the methods

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described above but with the treatment groups comprising: untreated (FIG. 36A); 50 μM 7o (FIG. 36B); 25 μM 7o (FIG. 36C) and 12.5 μM 7o (FIG. 36D). Composite stacked scanning confocal laser micrographs of *P. aeruginosa* (PAO1 (pLVAgfp)) clearly indicate a dose-dependent disruption of the biofilm.

Example 44

Assay of Dose-Dependent Disruption of *P. aeruginosa* Biofilm: Compound 7h

The dose-dependent relationship of synthetic quorum sensing compound 7h was explored using the methods described above but with the treatment groups comprising: 15 untreated (FIG. 37A); 50 μM 7h (FIG. 37B); 25 μM 7h (FIG. 37C); and 12.5 μM 7h (FIG. 37D). Composite stacked scanning confocal laser micrographs of *P. aeruginosa* (PAO1 (pLVAgfp)) clearly indicate a dose-dependent disruption of the biofilm.

The importance of these findings is highlighted by the fact that few inhibitors of bacterial biofilm formation are known. Moreover the effects illustrated in FIGS. 36A-D and FIGS. 37A-D illustrate that there is a dose-dependent effect for these compounds. This indicates intermediate responses are obtainable and that the mechanism of action may be similar for each of the compounds analyzed throughout a broad range of genera of quorum-sensing bacteria.

Quorum sensing compounds such as those herein described, are anticipated to have direct impact in health care and other industries where bacterial contamination is prevalent. The ability of a soluble, diffusible quorum sensing compound to inhibit biofilm formation when introduced or contacted to an environment has direct clinical and public health implications such as those related to bacterial contamination of surgical instruments and food and water resources. Moreover, the discovery of potent inhibitors from the small combinatorial library described herein highlights the potential utility of focused combinatorial methods for the discovery of additional small molecule modulators of quorum sensing.

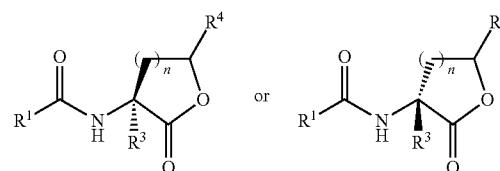
In summary, the invention provides a robust synthetic route to quorum sensing compounds, including AHL autoinducers, that provides access to both natural and unnatural AHLs in high purity. The synthetic route herein disclosed has been used to identify a set of non-native AHLs that are potent 45 inhibitors of bacterial quorum sensing. Further, the compounds developed by the disclosed methods allows the identification of both agonists and antagonists and provides methods for their rapid characterization and screening. Such compounds may be used to both inhibit cell to cell communication via quorum sensing as well as create miscommunication between such cells via the quorum sensing system. Manipulate the quorum sensing of pathogenic and destructive bacteria allows its use in research, health care, agriculture, food processing and other industries to either inhibit signaling and therefore inhibit the transcription of lethal genes or to 50 prematurely elicit the transcription of such genes so as to short-circuit the associated pathogenesis. In short, the compounds and methods described herein represent powerful new tools to control the growth, virulence and pathogenicity of 55 bacteria.

What is claimed is:

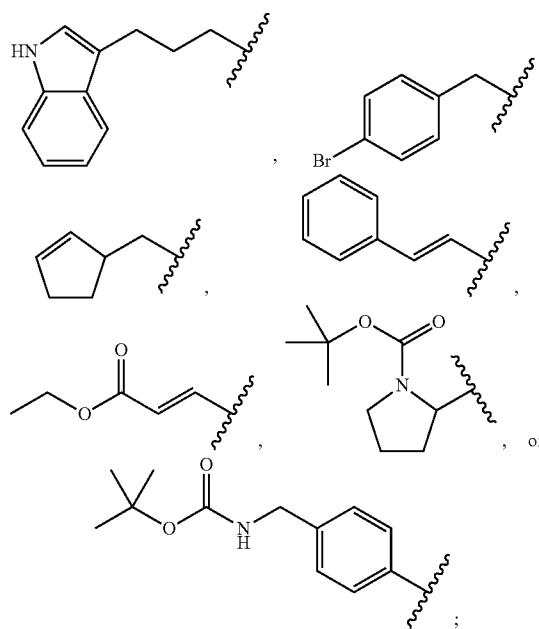
1. A method for modulating or regulating infectivity and pathogenicity of a quorum sensing bacterium which comprises contacting the quorum sensing bacterium with one or more compounds selected from the group consisting of:

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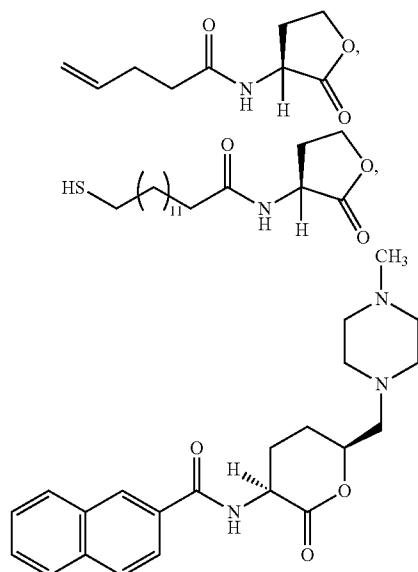
(a) a compound having formula:



where n is 1, 2 or 3,
R⁴ is —H, R³ is —H, —CH₃ or —CH₂—CH₃, and
R¹ is selected from:

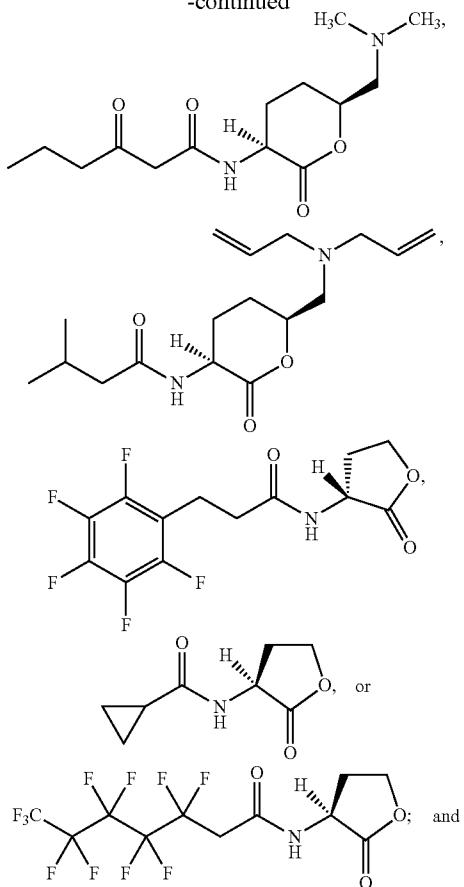


(b) a compound selected from compounds having formulas:

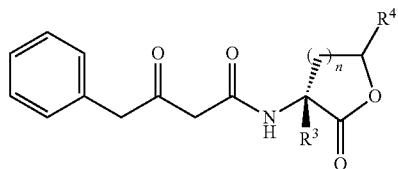


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-continued



(c) a compound of formula:



where n is 1, 2 or 3, R³ is selected from —H, —CH₂CH₃, or —CH₃, and R⁴ is selected from —H, or —CH₃.

2. The method of claim 1, wherein the one or more compounds act as agonists of native quorum sensing compounds.

3. The method of claim 1, wherein the one or more compounds act as antagonists of native quorum sensing compounds.

4. The method of claim 1, wherein the quorum sensing bacterium is a Gram negative bacterium.

5. The method of claim 1, wherein modulating or regulating infectivity and pathogenicity of a quorum sensing bacterium includes reducing the bacterial contamination of surfaces prone to bacterial biofilm contamination comprising contacting the surface with one or more of said compounds.

6. The method of claim 5, wherein contacting includes: spraying, painting, brushing, applying or treating the surface with a solution containing one or more of said compounds.

7. The method of claim 6, wherein the solution is an aqueous solution which contains a mixture of two or more of said compounds.

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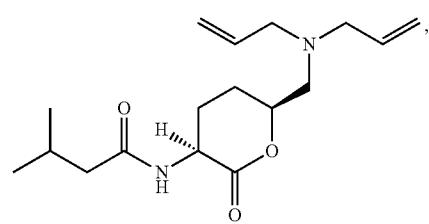
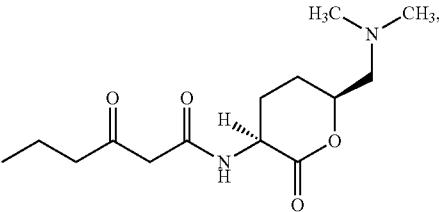
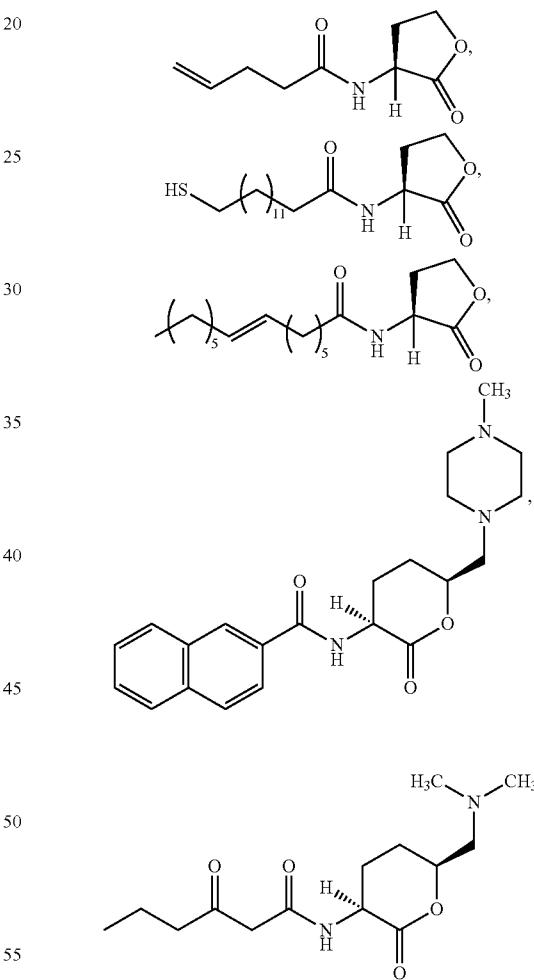
8. The method of claim 6 wherein modulating or regulating infectivity and pathogenicity of a quorum sensing bacterium includes inhibiting or diminishing biofilm production and the method comprises the step of contacting the bacterial biofilm with a composition comprising one or more of said compounds.

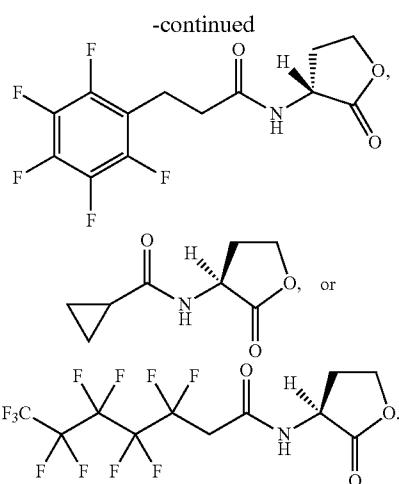
9. The method of claim 8, wherein the step of contacting includes the step of spraying, painting, brushing, applying or treating the bacterial biofilm with the composition.

10. The method of claim 1, wherein the contacting is performed in vitro.

11. The method of claim 1, wherein the contacting is performed in vivo.

12. The method of claim 1 wherein the one or more compounds are selected from:



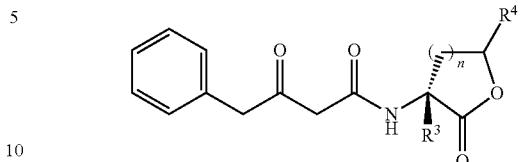
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13. The method of claim 12, wherein modulating or regulating infectivity and pathogenicity of quorum sensing bacteria comprises reducing the virulence of quorum sensing bacteria.

14. The method of claim 13, wherein reducing the virulence of a quorum sensing bacteria includes inhibiting or diminishing biofilm production.

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15. The method of claim 1 wherein the one or more compounds have formula:



where n is 1, 2 or 3, R³ is selected from —H, —CH₂CH₃, or —CH₃, and R⁴ is selected from —H, or —CH₃.

15 16. The method of claim 15, wherein the one or more compounds act as antagonists of native quorum sensing compounds.

17. The method of claim 15, wherein the one or more compounds act as agonists of native quorum sensing compounds.

20 18. The method of claim 15, wherein the quorum sensing bacterium is a Gram negative bacterium.

19. The method of claim 15, wherein R³ and R⁴ are —H.

20 20. The method of claim 15, wherein n is 1.

21. The method of claim 15, wherein n is 1 and R³ and R⁴ are —H.

* * * * *