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Kawaoka et al.

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(54) **HIGH TITER RECOMBINANT INFLUENZA VIRUSES WITH ENHANCED REPLICATION IN VERO CELLS**

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(51) **Int. Cl.**

C07K 14/005 (2006.01)

C12N 7/00 (2006.01)

A61K 39/00 (2006.01)

(52) **U.S. Cl.**

CPC **C07K 14/005** (2013.01); **C12N 7/00** (2013.01); **A61K 2039/525** (2013.01); **C12N 2760/16122** (2013.01); **C12N 2760/16152** (2013.01)

(58) **Field of Classification Search**

CPC **A61K 39/145**; **A61K 2039/5258**; **A61K 39/12**; **A61K 2039/5254**; **A61K 39/42**; **A61K 2039/525**; **C12N 2760/16134**; **C12N 2760/16123**; **C12N 2760/16222**; **C12N 2760/16251**; **C12N 2760/16162**; **C12N 2760/16243**; **C12N 2760/16111**; **C12N 2760/16161**

See application file for complete search history.

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(57)

ABSTRACT

The invention provides a composition useful to prepare high titer influenza viruses, e.g., in the absence of helper virus, which includes internal genes from an influenza virus vaccine strain or isolate, e.g., one that is safe in humans, for instance, one that does not result in significant disease, and genes from vaccine seed virus isolates which include a HA gene segment with a HA2 sequence encoding a HA2 that confers enhanced growth in cells in culture, such as Vero cells.

22 Claims, 20 Drawing Sheets

(56)

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PR8(Cambridge)

PB2

AGCGAAAGCAGGTCAATTATATTCAATATGGAAAGAATAAAAGAACTAAGAAATCTAATGTCGCACTCTCGCACCCGCGAGATA
CTCAGAAAAACCCGCTGGACCATATGGCCATAATCAAGAAGTACACATCAGGAAGACAGGAGAAGAACCCAGCACTTAGGATG
AAATGGATGATGGCAATGAAATATCCAATTACAGCAGACAAGAGGATAACGGAAATGATTCCTGAGAGAAATGAGCAAGGACAA
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CCAATGACAAATACAGTTCATTATCCAAAAATCTACAAAATCTATTTTGAAGAGTGAAGGCTAAAGCATGGAACCTTTGGC
CCTGTCCATTTTGAAGAACCAAGTCAAAATACGTCCGAGAGTTGACATAAACTCTGGTTCATGCAGATCTCAGTGCCAAGGAGGCA
CAGGATGTAATCATGGAAGTTGTTTTCCCTAACGAAGTGGGAGCCAGGATACTAACATCGGAATCGCAACTAACGATAACCAAA
GAGAAGAAAGAAGAACTCCAGGATTGCAAAATTTCTCCTTTGATGGTTGCATACATGTTGGAGAGAGAACTGGTCCGCAAAACG
AGATTCCTCCCACTGGCTGGTGGAAACAAGCAGTGTGTACATTGAAGTGTTCATTTGACTCAAGGAACATGCTGGGAACAGATG
TATACTCCAGGAGGGGAAGTGAAGAATGATGATGTTGATCAAGCTTGATTATTGCTGCTAGGAACATAGTGAGAAGAGCTGCA
GTATCAGCAGACCCACTAGCATCTTTATTGGAGATGTGCCACAGCACACAGATTGGTGGAAATAGGATGGTAGACATCCTTAAG
CAGAACCCCAACAGAAGAGCAAGCCGTGGATATATGCAAGGCTGCAATGGGACTGAGAATTAGTCACTCTTCAGTTTGGTGGGA
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GTGCATGAGGGATCTGAAGAGTTCACAAATGGTTGGGAGAAGAGCAACAGCCATACTCAGAAAAGCAACAGGAGATTGATTAG
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AAGGATGCGAAAGTGTCTTTTCAAAATTTGGGAGTTGAACCTATCGACAATGTGATGGGAATGATTGGGATATTGCCGACATG
ACTCCAAGCATCGAGATGTCAATGAGAGGAGTGAGAATCAGCAAAATGGGTGTAGATGAGTACTCCAGCACGGAGAGGGTAGTG
GTGAGCATTGACCGGTTCTTGAGAGTCAGGGACCAACGAGGAAATGTACTACTGTCTCCGAGGAGGTGAGTGAAACACAGGGA
ACAGAGAACTGACAATAACTTACTCATCTGCAATGATGTGGGAGATTAAATGGTCTGAATCAGTGTGGTCAATACCTATCAA
TGGATCATCAGAACTGGGAACTGTTAAATTCAGTGGTCCCAGAACCTACAATGCTATACAATAAAATGGAAATTTGAACCA
TTTCAGTCTTTAGTACCTAAGGCCATTAGAGGCCAATACAGTGGGTTTGAAGAACTCTGTTCCAACTAAATGAGGGATGTGCTT
GGGACATTTGATACCCACAGATAATAAACTTCTTCCCTTCGACGCCGCTCCACCAAAGCAAAGTGAATGCAGTTCCTCTCA
TTTACTGTGAATGTGAGGGGATCAGGAATGAGAATACTTGAAGGGCAATTTCTCTGTATTCACTACAACAAGGCCACGAAG
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AGGGGATTCTCATTCTGGGCAAGAAAGACAGGAGATATGGGCCAGCATTAAGCATCAATGAATGAGCAACCTTGCGAAAGGA
GAGAAGGCTAATGTGCTAATTTGGGCAAGGAGACGTGGTGTGGTAAATGAAACGAAACGGGACTCTAGCATACTTACTGACAGC
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SEQ ID NO:11

PB1

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GGAAGGCTCATAGACTTCTTAAGGATGTAATGGAGTCAATGAAAAAGAGAAATGGGGATCACAACCTATTTTCAAGAGAAAG
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GTCTGCTTAAAATGGGAATTTGATGGATGAGGATTACAGGGGGGTTTATGCAACCCACTGAACCCATTTGTCAAGCCATAAAGAA
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SEQ ID NO:10

Fig. 1A

PR8(Cambridge)

PA

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SEQ ID NO:12

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SEQ ID NO:13

M

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Fig. 1B

PR8(Cambridge)

GTCTCATAGGCAAATGGTGACAACAACCAACCCACTAATCAGACATGAGAACAGAATGGTTTTAGCCAGCACTACAGCTAAGGC
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SEQ ID NO:14

NS

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ACTTTCTCATTTAGCTTATTTAATAATAAAAAACACCCTTGTCTACT

SEQ ID NO:15

Fig. 1C

GROWTH PROPERTIES OF VERO-ADAPTED PR8 (PR8-VERO) VIRUS
IN VERO CELLS

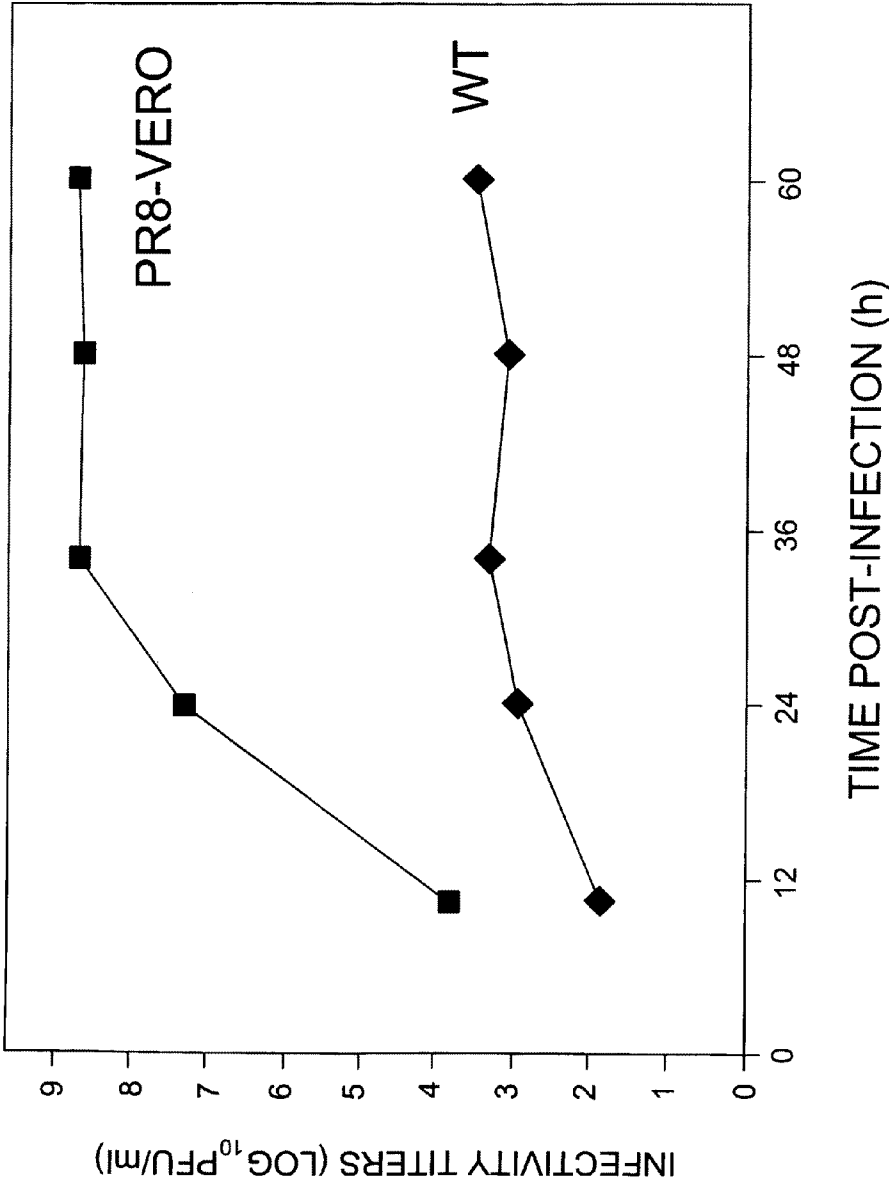


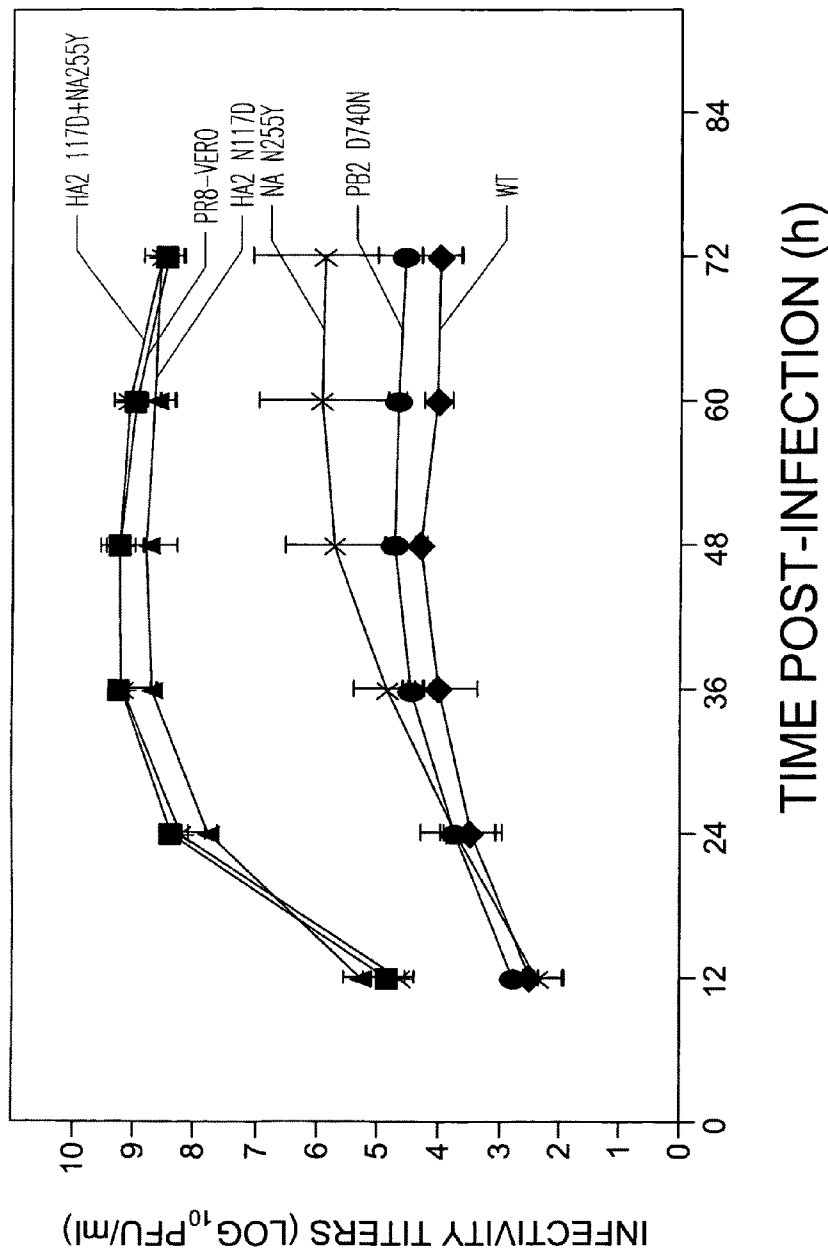
Fig. 2

COMPARISON OF AMINO ACID SEQUENCES
BETWEEN WT AND PR8-VERO

	POSITION	WT	PR8-VERO
HA2	117	N	D
NA	255	N	Y
PB2	740	D	N(2/4)

Fig. 3

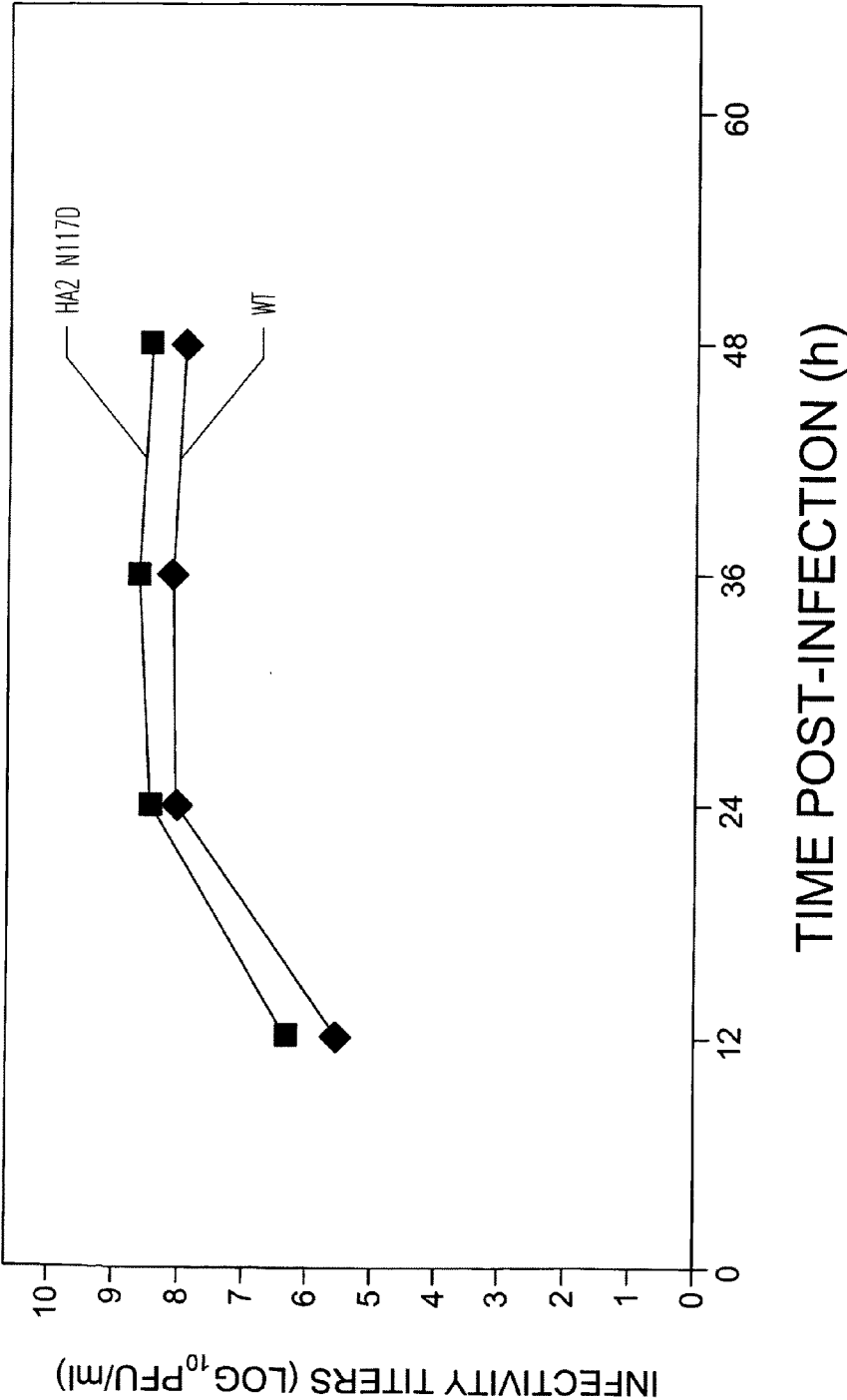
GROWTH PROPERTIES OF PR8 MUTANTS IN VERO CELLS



THE HA2 N117D MUTATION WAS MAINLY RESPONSIBLE FOR THE HIGH GROWTH PROPERTIES IN VERO CELLS.

Fig. 4

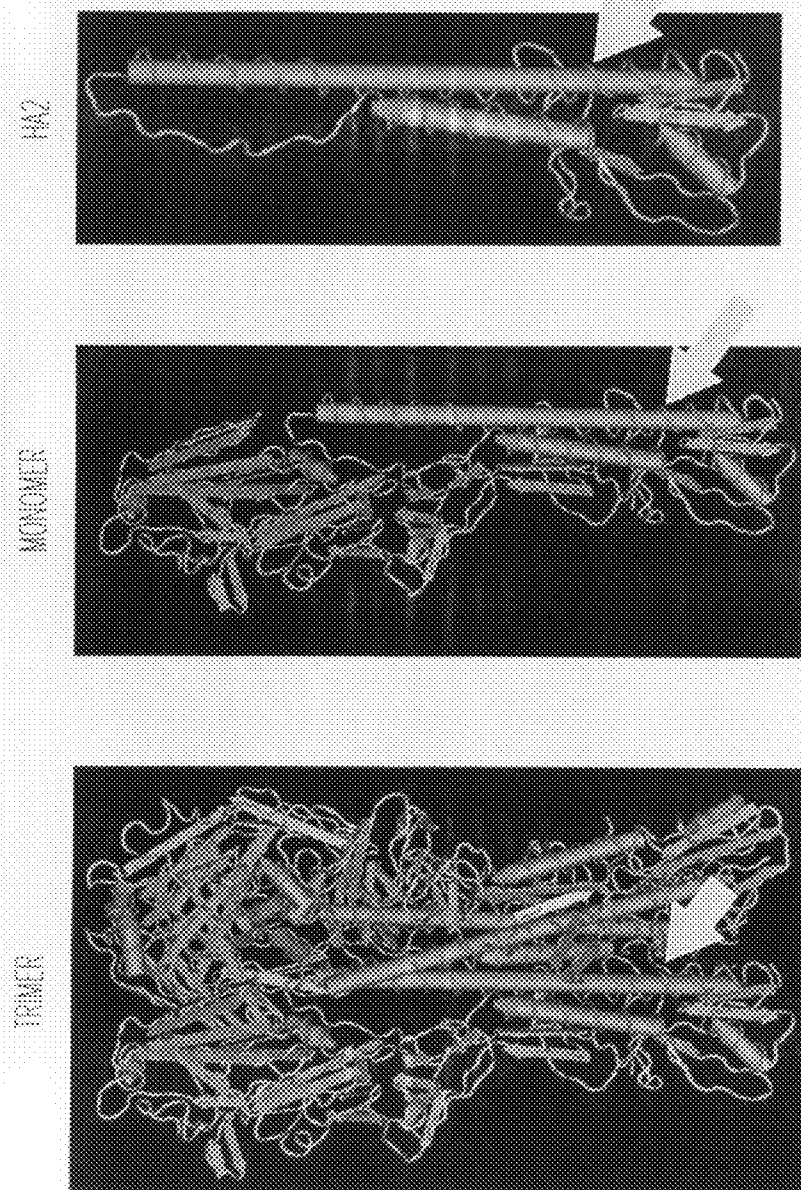
GROWTH PROPERTIES OF THE HA2 N117D MUTANT IN MDCK CELLS



REPLICATION EFFICIENCY WAS COMPARABLE BETWEEN THE WT AND THE MUTANT.

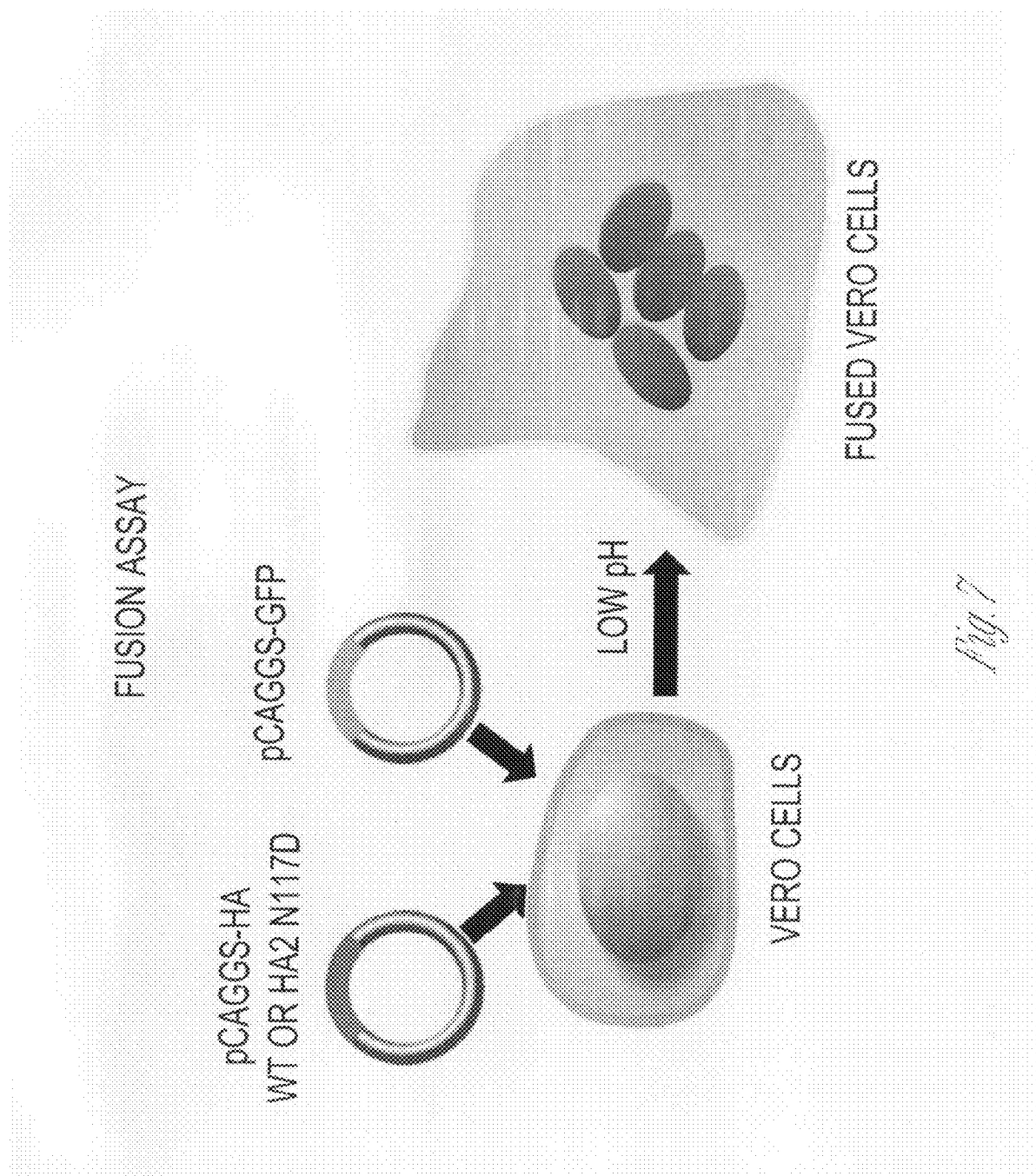
Fig. 5

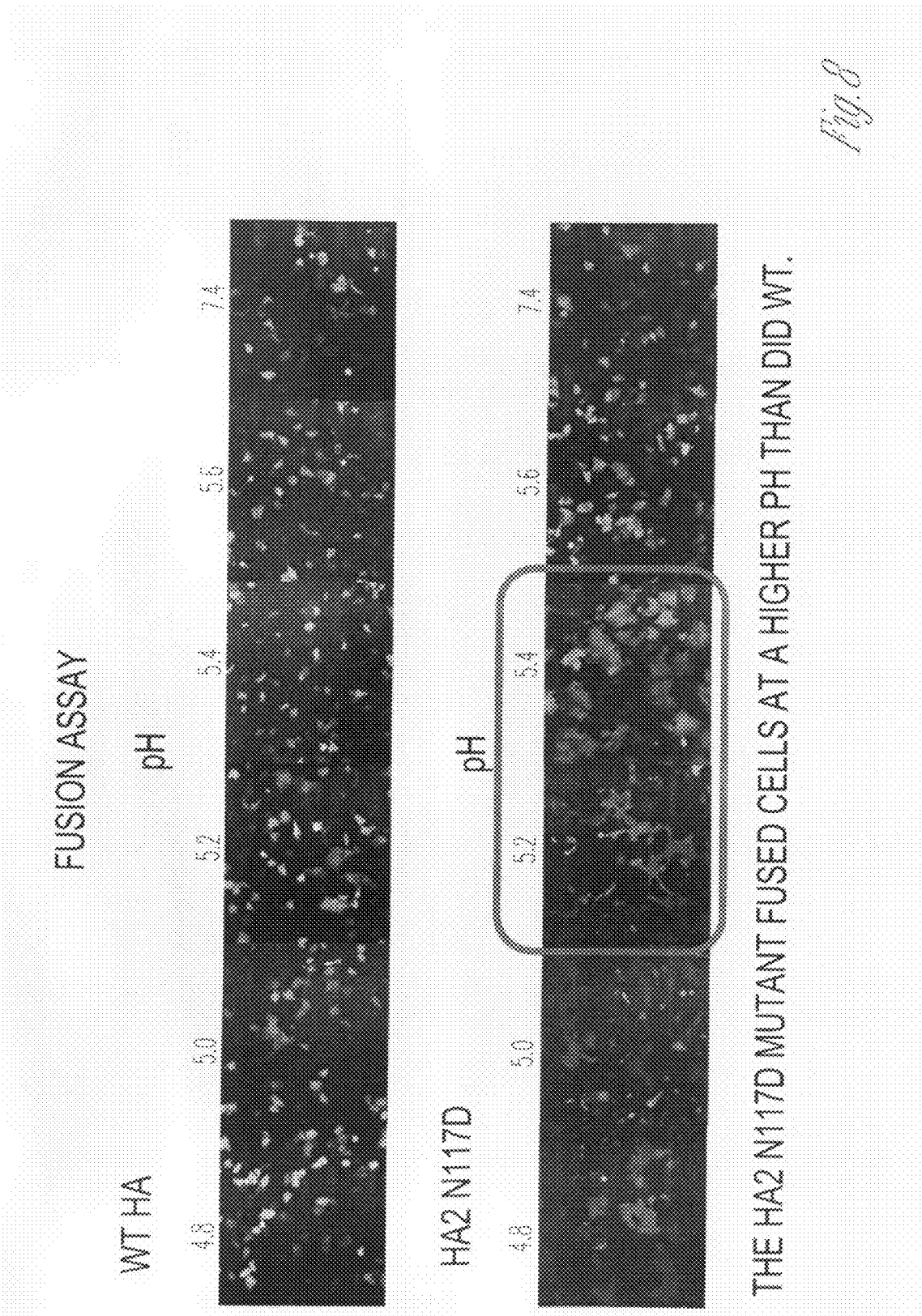
POSITION OF HA2 117 IN THE 3D STRUCTURE OF HA



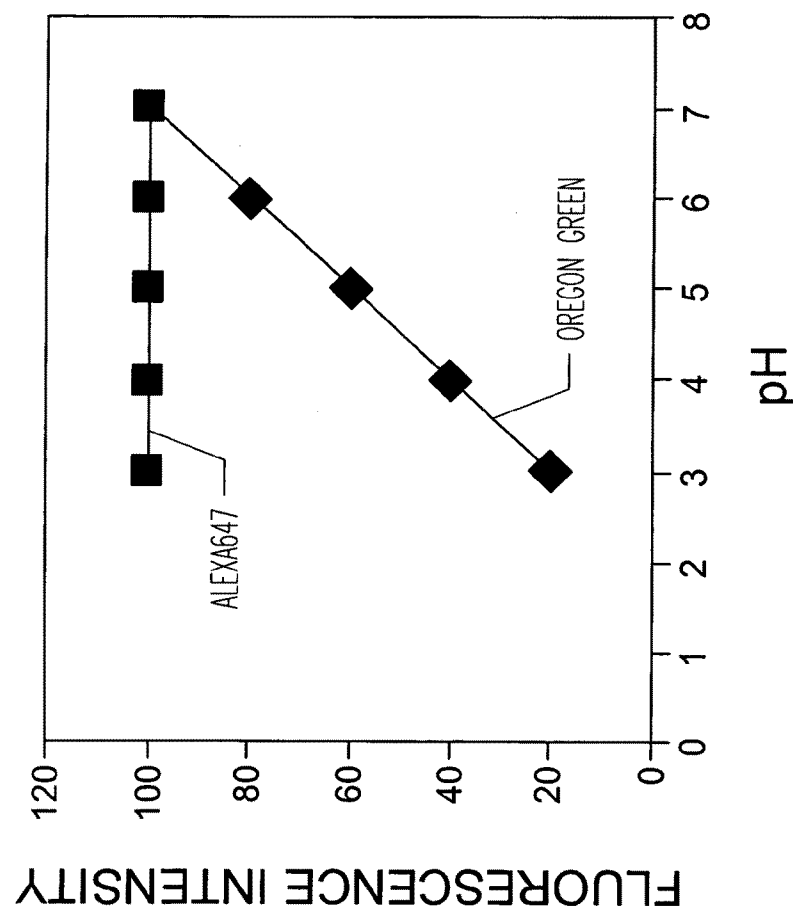
1934 HUMAN H1 HEMAGGLUTININ (MMDB ID: 26941, PDB ID: 1RU7)

Fig. 6





THE PRINCIPAL OF THE METHOD OF COMPARISON OF ENDOSOMAL
pH BETWEEN TWO DIFFERENT CELLS (MDCK VS. VERO CELLS)



FLUORESCENCE INTENSITY OF OREGON GREEN IS SENSITIVE TO LOW PH
ALTHOUGH INTENSITY OF ALEXA647 IS NOT SENSITIVE TO PH VALUE.

PH CAN BE COMPARED BY MEASURING THE INTENSITY AND CALCULATING THE RATIO BETWEEN ALEXA647 AND OREGON GREEN.

Fig. 9A

THE METHOD OF COMPARISON OF ENDOSOMAL pH
BETWEEN MDCK CELLS AND VERO CELLS

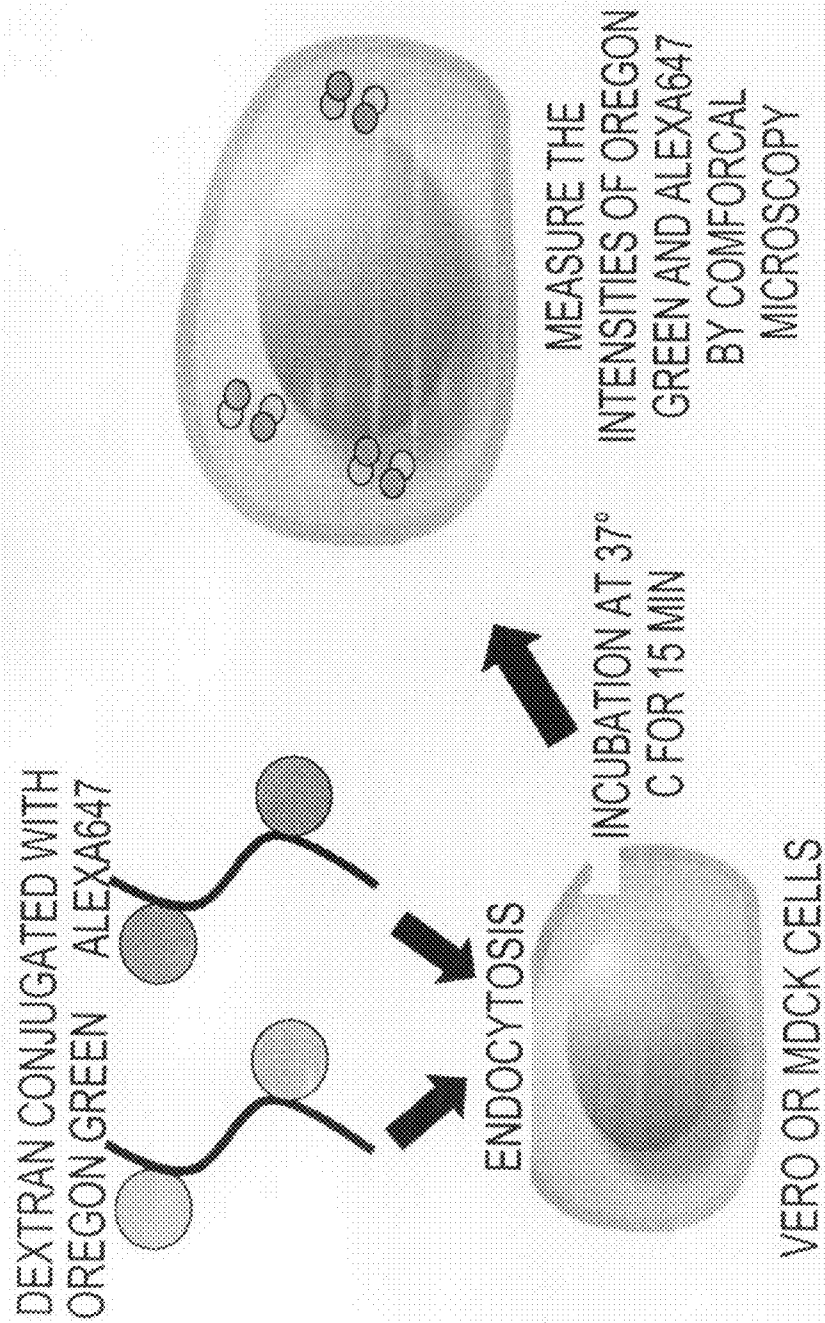


Fig. 9B

COMPARISON OF ENDOSOMAL pH
BETWEEN MDCK CELLS AND VERO CELLS

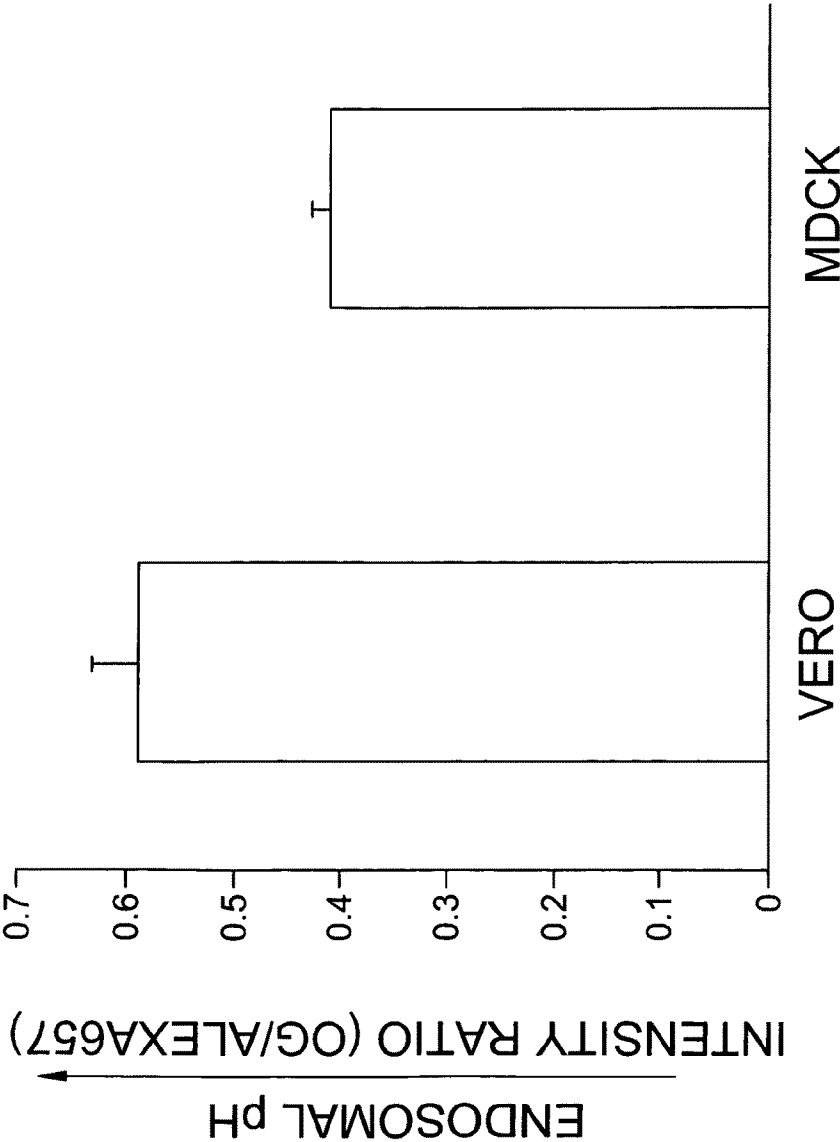


Fig. 10

THE HA2 N117D MUTATION ENHANCES THE REPLICATION EFFICIENCY OF
THE A/KAWASAKI/173/2001 (H1N1) 6:2 REASSORTANT WITH A PR8
DONOR IN VERO CELLS

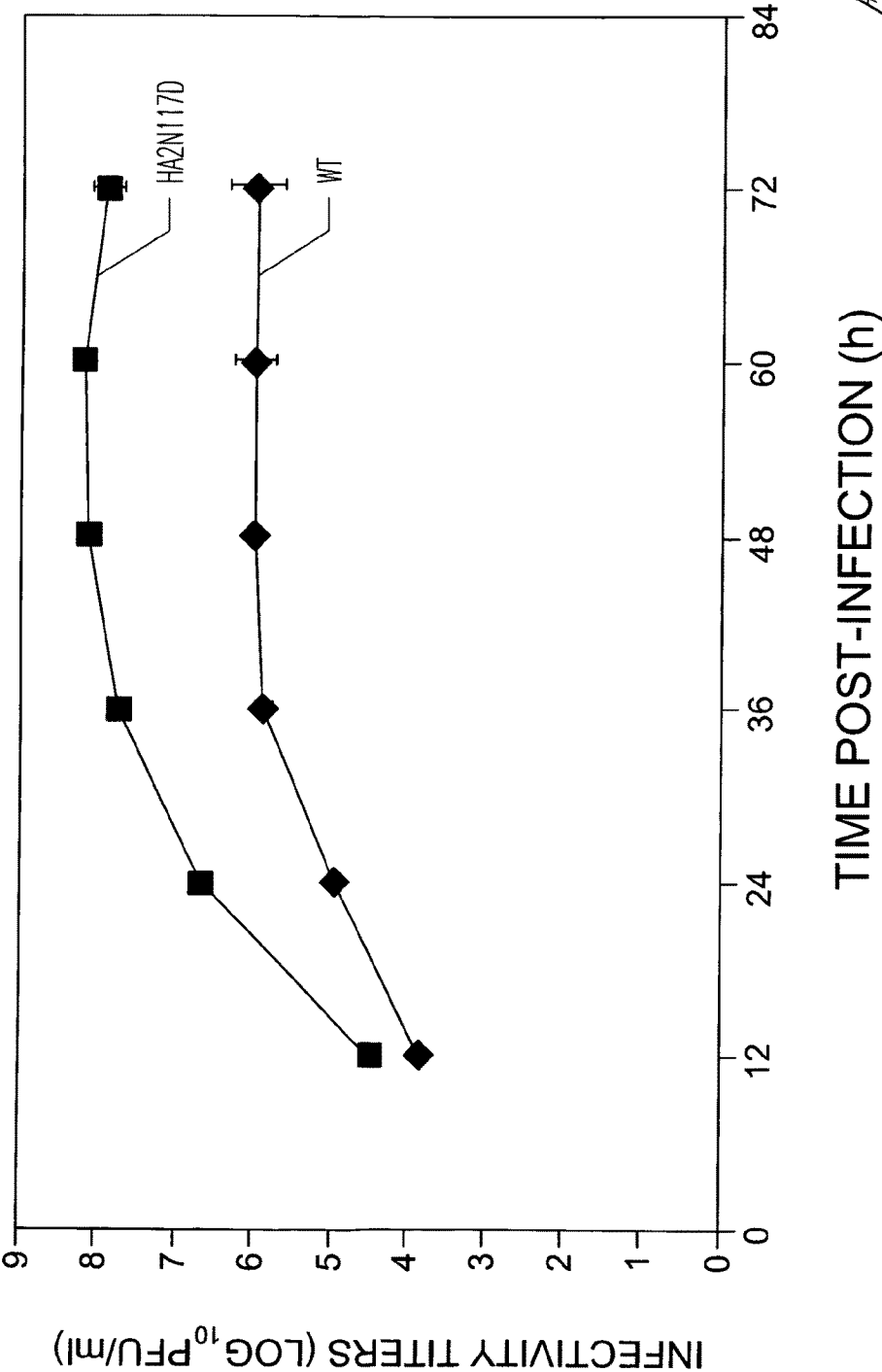


Fig. 11A

THE HA2 N117D MUTATION ENHANCES THE REPLICATION EFFICIENCY OF
THE A/KAWASAKI/UTK-4/2009 (H1N1) 6:2 REASSORTANT WITH A PR8
DONOR IN VERO CELLS.

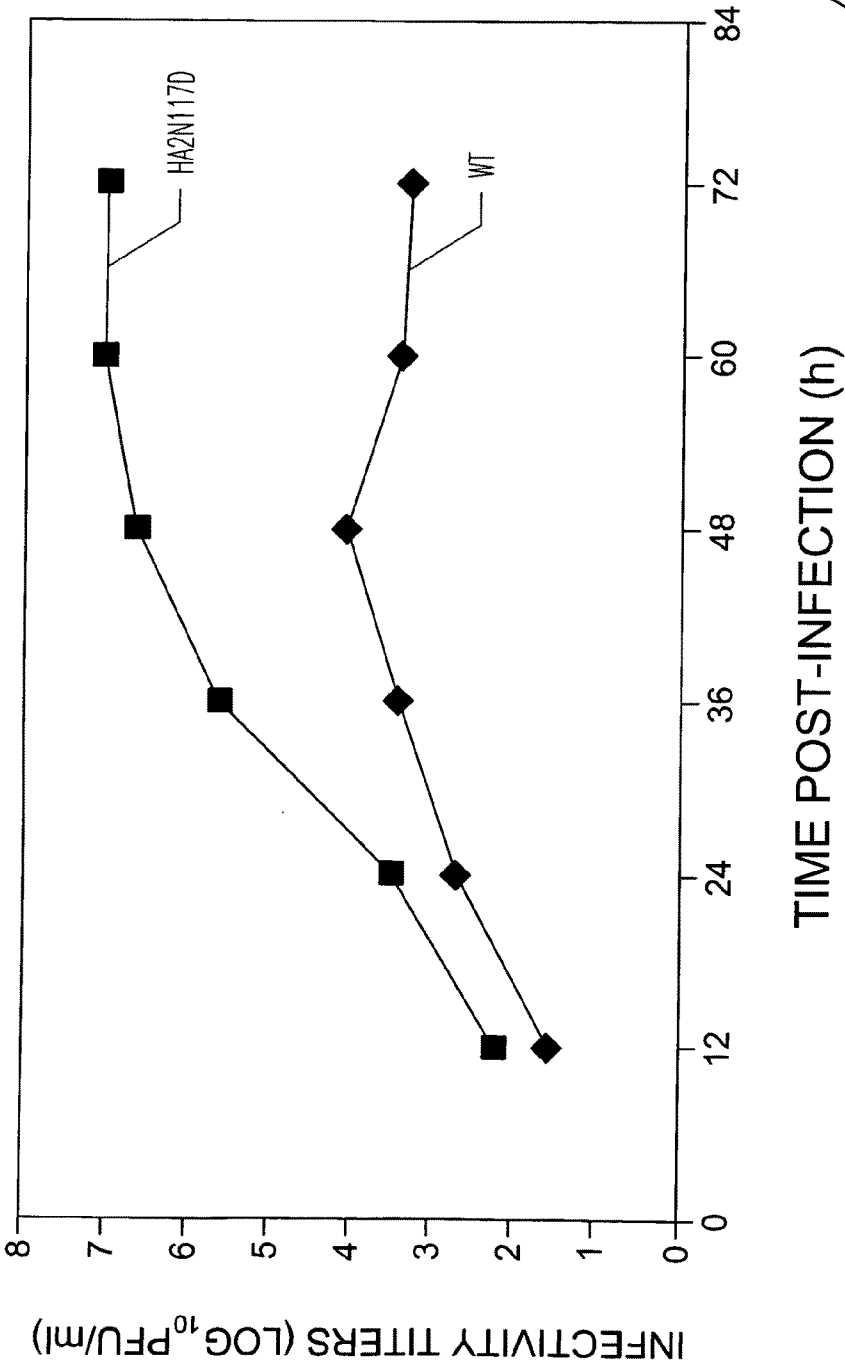


Fig. 11B

THE HA2 N117D MUTATION ENHANCES THE REPLICATION EFFICIENCY OF
THE AYOKOHAMA/2017/2003 (H3N2) 6:2 REASSORTANT WITH A PR8
DONOR IN VERO CELLS.

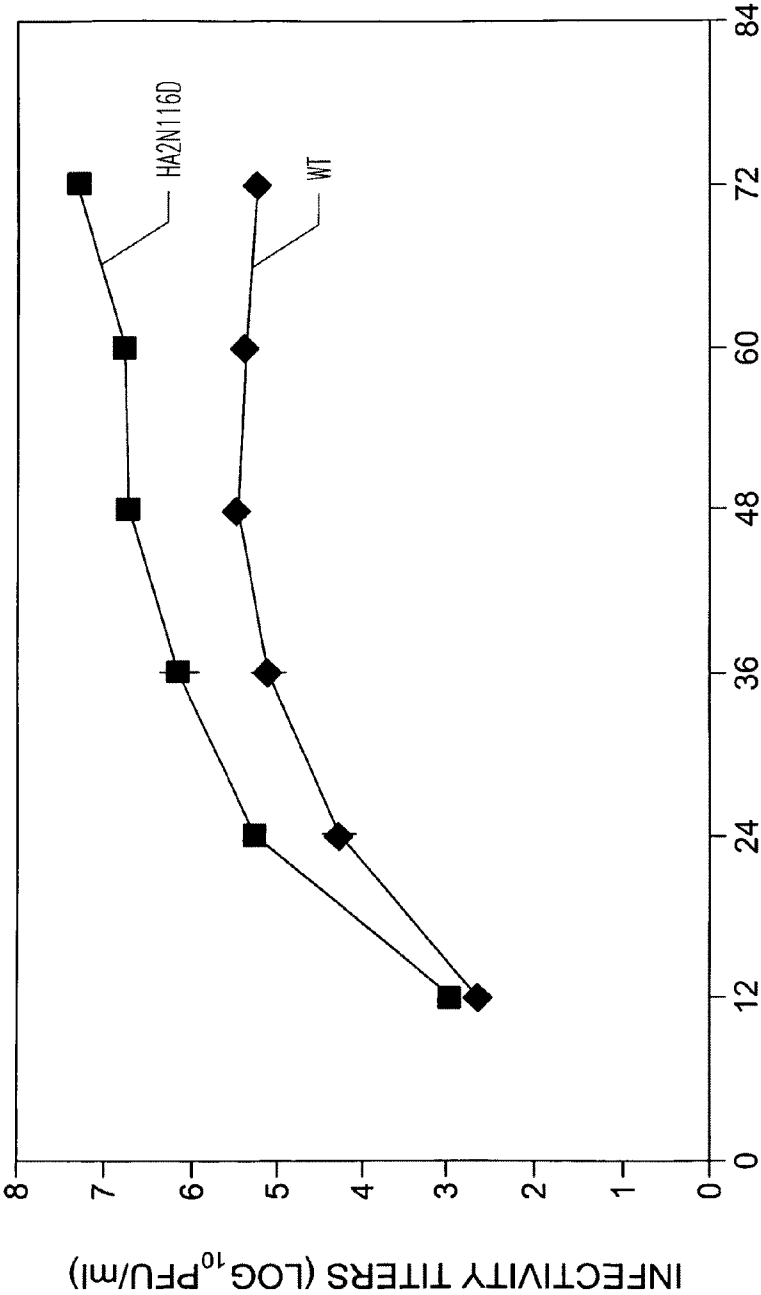


Fig. 11C

TIME POST-INFECTION (h)

HA1	11	107
H3HU	ATLCGLGHAVNGTLVKTITDDQIEVTNATELVQSSSTGKICN.NPHRILDGIDCTLIDALLGDPHCDVFQN.ETWDLFVERSKAFS.NCYPYDVPDYAS	
H5AV	DQI I Y NNSTEQ D MEKN T H QDILEKTHN L DL GVKP ILR SVAGW N M E L VPE SYI KDNPVNGL ENFN EE	
H5HU	DQI I Y NNSTEQ D MEKN T H QDILERTHN L DL GVKP ILR SVAGW N M E I VPE SYI KASPANDL GNFN EE	
H9SW	DKI I YQSTNSTET D L ETN P H K LHTEHN ML AT LGHP ILDT IEGLIY N S LLLGGRE SYI PS VNGM GN ENLEE	
H9HU	DKI I QSTNSTET D L ETN P H K LHTEHN ML ATSLGHP ILDT IEGLVY N S LLLGGRE SYI S VNGT GN ENLEE	
	108	203
H3HU	LRSLVASSGTLEFITEGF...TWTGVTON.CGSNACKRPGSGFFSRNLNWLTKSGSTYPVLNVTMPNNDNFDKLYIMGIHPSTNQEQTSLYVQASGRVT	
H5AV	KH LS TNHF K RI.IPRSS SNHDASS V S PYNGR S RNVV I KNN A TIKRSYN TNQE L IL NDAA K QNPPTY S	
H5HU	KH LSRINF K QI.IPKSS SNHDASS V S PYLGR S RNVV I KN A TIKRSYN TNQE L VL NDAA K QNPPTY S	
H9SW	FS ASSYQR QI. PDTI .N SYS. T K S....DS RSMR QKNN A QDAQYT RGKSI M N P DTV N TRTDTTTS	
H9HU	T FS ASSYQR QI. PDT .N YT. T R S....GS RSMR QKSGF QDAQYT RGKSI P YT N RNDTTTS	
	204	302
H3HU	VSTRRSQQTIPNIGSRPWVRLSSRISYIWTIVKPGDVLVINSNGNLIAPR.GYFKMRTGKSSIMRSDAPIDTCISECITPNGSIPNDKPFQNVNKITY	
H5AV	G STLN RS E AT K N Q G MEF L N AINFE F EYA KIVKK G A K GLEYGN NTK Q M A NSSM H HPL I	
H5HU	G STLN RL E AT K N Q G MEF L N AINFE F EYA KIVKK D T K ELEYGN NTK Q M A NSSM H HPL I	
H9SW	T EDINR FK V P L N HG DY S L QT R R WY HILSGESHGR LKT LNSGN VQ Q ER GLNTTL H S YA	
H9HU	T EDLNR FK V P L N QG DY S L QT R R WY HVLSGGSHGR LKT LKGGN VQ Q EK GLNSTL H S YA	
	303	328 *
H3HU	GACPKYVKONTLKLATGMERNVPEKQT....R	SEQ ID NO:16
H5AV	E SGR V L ORE	SEQ ID NO:17
H5HU	E S R V L T QRERRKK	SEQ ID NO:18
H9SW	N GVKS V L ARSS....	SEQ ID NO:19
H9HU	T RV S V L ARSS....	SEQ ID NO:20

Fig. 12A

HA2
100
1 GLFGAIAAGFIENGWEGMIDGWYGFRHONSEGTOQAADLKSTQAAIDQINGKLN RVIEKTNEKFHQIEKFSEVEGRIQDLEKYVEDTKIDLWSYNAELLV
H3HU H S EQ S Y KE K GTTN V S D M TQ EA G>NNL R EN N KM GFL V T
H5AV G Q H S EQ S Y KE K G TN V S N M TQ EA GR>NNL R EN N KM GFL V T
H5HU G Q H S EQ S Y KE K G TN V S N M TQ EA GR>NNL R EN N KM GFL V T
H9SW G P L A Q S DQ V M RD K K TS V N D M KQ GI DH T LNMINNK D QIQ I T
H9HU G P L A Q S DQ V M RD K K TS V N D M KQ EI DH T LNMINNK D QIQ V A

101
199
101 ALENQHTIDLTDSMNKLFKTRRQLRENAEEMGNGCFKIYHKCDNACIESIRNGTYDHDVYRDEALNNRFFQIKGVELKSGYKDWILWI.SFAISCFLLC
H3HU LM ER L FH NVKN D V L D K L EF E M K YPQ SE RL EE S K E MGIYQ S Y TVA SLA A
H5AV LM ER L FH NVKN D V L D K L EF E M K YPQ SE RL EE S K E MGIYQ S Y TVA SLA A
H5HU L K L EH ANV N N VK A GS M D K EL DQ M T NRRK KE SKLE QK E K E EGTYK T Y TVA SLVIA
H9SW L K L EH ANV N N VK A GS M D K EL DQ M T NRRK E SRLE QK E -----
H9HU

200
221
H3HU VLLGFIMWACQGNIRCNICI SEQ ID NO:23
H5AV MIA LSL M SN SLQ R SEQ ID NO:24
H5HU MVA LSL M SN SLQ R SEQ ID NO:25
H9SW MGFAA LF MS----- SEQ ID NO:26
H9HU ----- SEQ ID NO:27

Fig. 12A CONT'D

1 MKAILLVLLY TFATANADTL CIGYHANNST DTVDTVLEKN VTVTHSVNLL EDKHNGKLCK
61 LRGVAPLHLG KCNIAGWILG NPECESLSTA SSWSYIVETP SSDNGTCYPG DFIDYEELRE
121 QLSSVSSFER FEIFPKTSSW PNHDSNKGVT AACPHAGAKS FYKNLIWLK KGNSYPKLSK
181 SYINDKGKEV LVLWGIHHP TSADQQSLYQ NADAYVFVGS SRYSKFKPE IAI RPKVRDQ
241 EGRMYYWTL VEPGDKITFE ATGNLVVPY AFAMERNAGS GIIISDTPVH DCNTTCQTPK
301 GAIN TSLPFQ NIHPITIGK PKYVKSTKLR LATGLRNIPS IQSRGLFGAI AGFIEGGWTG
361 MVDGWYGYHH QNEQSGGYAA DLKSTQNAID EITNKVNSVI EKMNTQFTAV GKEFNHLEKR
421 IENLNKKVDD GFLDIWTYNA ELLVLLENER TLDYHDSNVK NLYEKVRSQ L KNNAKEIGNG
481 CFEFYHKCDN TCMESVKNGT YDYPKYSEEA KLNREEIDGV KLESTRIYQI LAIYSTVASS
541 LVLVSLGAI SFWMCSNGSL QCRICI SEQ ID NO:21

Fig. 12B

A/Kawasaki/173/2001 (H1N1)
GLFGAIAGFIEGGWTGMVDGWYGYHHQNEQSGYAADQKSTQNAINGITNKVNSVIEKMNTQFTAVG
KEFNKLERRMENLNKKVDDGFIDIWTYNAELLVLENERTLDFHDSNVKDLYEKVKSQLKNNAKEIGNGCF
EFYHKCNNECMESVKNGTYDYPKYSEESKLNREKIDGVKLESMGVYQILAIYSTVASSLVLVSLGAISFWM
CSNGSLQCRICI

SEQ ID NO:28

A/Kawasaki/UTK-4/2009 (H1N1)
GLFGAIAGFIEGGWTGMVDGWYGYHHQNEQSGYAADQKSTQNAINGITNKVNSVIEKMNTQFTAVG
KEFNKLERRMENLNKKVDDGFIDIWTYNAELLVLENERTLDFHDSNVKDLYEKVKSQLKNNAKEIGNGCFE
FYHKCNDECMESVKNGTYDYPKYSEESKLNREKIDGVKLESMGVYQILAIYSTVASSLVLVSLGAISFWM
SNGSLQCRICI

SEQ ID NO:29

A/Yokohama/2017/2003 (H3N2)
GIFGAIAGFIENGWEGMVDGWYGFRHQNSEGTGQAADLKSTQAAINQINGKLNRLIGKTNEKFHQIEKEF
SEVEGRIQDLEKYVEDTKIDLWSYNAELLVALENQHTIDLTDSEMDKLFERTKKQLRENAEDMNGCGCFKIYH
KCDNACIESIRNGTYDHDVYRDEALNNRFQIKGVELKSGYKDWILWISFAISCFLLCVALLGFIWACQKGN
IRCNICI

SEQ ID NO:30

Fig. 13

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HIGH TITER RECOMBINANT INFLUENZA VIRUSES WITH ENHANCED REPLICATION IN VERO CELLS

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of the filing date of U.S. application Ser. No. 61/254,795, filed on Oct. 26, 2009, the disclosure of which is incorporated by reference herein.

STATEMENT OF GOVERNMENT RIGHTS

This invention was made with United States government support awarded by the National Institutes of Health (grant NIH AI069274). The United States government has certain rights in this invention.

BACKGROUND

Influenza is a major respiratory disease in some mammals including horses and is responsible for substantial morbidity and economic losses each year. In addition, influenza virus infections can cause severe systemic disease in some avian species, leading to death. The segmented nature of the influenza virus genome allows for reassortment of segments during virus replication in cells infected with two or more influenza viruses. The reassortment of segments, combined with genetic mutation and drift, can give rise to a myriad of divergent strains of influenza virus over time. The new strains exhibit antigenic variation in their hemagglutinin (HA) and/or neuraminidase (NA) proteins, and in particular the gene coding for the HA protein has a high rate of variability. The predominant current practice for the prevention of flu is vaccination. Most commonly, whole virus vaccines are used. As the influenza HA protein is the major target antigen for the protective immune responses of a host to the virus and is highly variable, the isolation of influenza virus and the identification and characterization of the HA antigen in viruses associated with recent outbreaks is important for vaccine production. Based on prevalence and prediction, a vaccine is designed to stimulate a protective immune response against the predominant and expected influenza virus strains (Park et al., 2004).

There are three general types of influenza viruses, Type A, Type B and Type C, which are defined by the absence of serological crossreactivity between their internal proteins. Influenza Type A viruses are further classified into subtypes based on antigenic and genetic differences of their glycoproteins, the HA and NA proteins. All the known HA and NA subtypes (H1 to H15 and N1 to N9) have been isolated from aquatic birds, which are thought to act as a natural reservoir for influenza. The H1N1 "swine flu" virus has recently been declared to be a pandemic. While this virus may be less virulent than some circulating influenza viruses in certain populations, it is ubiquitous and has become the subject of significant public health efforts. Unfortunately, this virus appears to be less amenable than other viruses to high titer productions which may lead to challenges in vaccine manufacture.

SUMMARY OF THE INVENTION

The invention provides isolated recombinant, e.g., reassortant, influenza viruses with selected amino acid residues at specified positions in HA2, NA and/or PB2. In one embodiment, the recombinant reassortant influenza virus has an

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amino acid residue at position 117 in HA2 (position is based on H1 HA2 numbering; for example, position 117 in H1 HA2 corresponds to position 116 in H3 HA2) that results in enhanced growth in Vero cells relative to a corresponding virus with, for instance, an asparagine at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, the recombinant influenza virus has an amino acid residue at position 117 in HA2 that results in fusion of the virus with membranes in endosomes, e.g., late endosomes, at a higher pH relative to a corresponding virus with, for instance, an asparagine at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, the invention provides an isolated recombinant reassortant influenza virus having six "internal" gene segments from a vaccine influenza virus, a NA gene segment selected from a first influenza virus isolate, and a HA gene segment selected to encode an aspartic acid or glutamic acid at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. For example, the NA and HA gene segments may be from a strain for a seasonal flu vaccine or from a pandemic strain, and in one embodiment, the HA2 sequence in the HA gene segment is mutated to encode an aspartic acid or glutamic acid at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2.

As described herein, an influenza virus isolate useful as a vaccine virus (A/Puerto Rico/8/34 (PR8) to carry heterologous gene segments for NA and/or HA was serially passaged in Vero cells to obtain virus with enhanced replication in those cells. In one embodiment, viruses obtained after serial passage which have enhanced replication, have titers that are at least 2, 3, 4 or 5 logs higher than viruses that were not serially passaged. In one embodiment, viruses obtained after serial passage had substitutions in three gene segments, NA, HA and PB2, relative to the parent virus. It was determined that the substitution in HA2 was primarily associated with the enhanced growth phenotype. PR8 virus with HA2 N117D had at least a three log enhancement in titer in Vero cells. The HA2 N117D mutant fused cells at a higher pH than did wild-type HA. Three different recombinant (6:2 mutant reassortant) influenza viruses were prepared that had the same PR8 "internal" genes (i.e., those other than the HA and NA genes), and the NA and HA from a single isolate, and where the residue at position 117 (or position 116 in the H3 reassortant) in HA2 was altered to aspartic acid. All of the 6:2 mutant reassortants showed enhanced growth in Vero cells relative to the corresponding parent 6:2 reassortant. Thus, for vaccine viruses that are to be grown or passaged in cells in culture, e.g., Vero cells, replacement of the residue at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2, e.g., by mutation, or selection of a HA gene segment with a residue that confers enhanced growth of the virus in cultured cells, can result in significantly higher viral titers. Thus, the invention provides a method to select for influenza viruses with enhanced replication in cell culture. The method includes providing cells suitable for influenza vaccine production; serially culturing one or more influenza virus isolates in the cells; and isolating serially cultured virus with enhanced growth relative to the one or more isolates prior to serial culture. In one embodiment, the cells are rodent or primate, e.g., human, cells. Also provided is a method to identify a HA2 that confers altered growth of a recombinant influenza virus. The method includes introducing one or more substitutions in influenza virus HA2 into a HA gene segment to yield a mutant HA gene segment; and identifying whether the mutant HA gene segment, when present in a replication competent recombinant influenza virus, results in enhanced replication of the recombinant influenza virus in a cell relative

to a corresponding replication competent influenza virus without the one or more substitutions in HA2. In one embodiment, at least one substitution is at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2, e.g., the at least one substitution is to aspartic acid or glutamic acid. In one embodiment, the cells are rodent or primate cells. In one embodiment, the one or more substitutions are to an amino acid residue with an acidic side chain.

In one embodiment, the influenza virus of the invention is a recombinant influenza virus having a mutant HA2 protein with at least one substitution that replaces an amino acid residue with an aliphatic side chain, amide-containing side chain, basic side chain, or sulfur containing side chain with a residue with an aromatic side chain or acidic side chain (a nonconservative substitution), e.g., at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, the influenza virus is a recombinant influenza virus having a HA2 protein with a residue with an aromatic side chain or acidic side chain at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, the recombinant influenza virus has a mutant HA2 protein with at least one substitution that replaces a neutral or positively charged residue with a polar or negatively charged residue, e.g., at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, the influenza virus is a recombinant influenza virus having a HA2 protein with a residue with a polar or negatively charged residue at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. The presence of the residue with the aromatic side chain or acidic side chain, or the polar or negatively charged residue, at position 117 in HA2 may alter the efficiency or rate of conformational change of HA or pH dependent membrane fusion. In one embodiment, the recombinant reassortant influenza virus comprises a HA gene segment selected to encode an aspartic acid or glutamic acid at position 117 in HA2, wherein recombinant virus has enhanced replication in Vero cells relative to a corresponding virus that does not have aspartic acid or glutamic acid at position 117 in HA2, e.g., where the corresponding virus has an alanine, asparagine, arginine or lysine at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, the recombinant virus has a NA gene segment with a tyrosine at position 255, wherein the numbering for NA residues is that for N1.

In one embodiment, the invention provides isolated influenza type A virus with a characteristic residue or substitution at position 117 of HA2, e.g., the residue at position 117 of HA2 is not asparagine, alanine, arginine or lysine, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, the isolated influenza type A virus of the invention with a characteristic residue or substitution at position 117 of HA2, has an HA2 amino acid sequence with at least 80%, e.g., 90%, 92%, 95%, 97% or 99%, including any integer between 80 and 99, contiguous amino acid sequence identity to a polypeptide encoded by one of SEQ ID NOs:16-20 or 22. In one embodiment, the isolated influenza type A virus of the invention with a characteristic residue or substitution at position 117 of HA2, has an HA1 from any one of subtypes 1-15 of HA. In one embodiment, an isolated influenza A virus of the invention has a nonconservative substitution at residue 117 of HA2, e.g., an asparagine to an aspartic acid substitution, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, the isolated influenza virus of the invention has an aspartic acid or glutamic acid at position 117 of HA2, wherein the numbering for HA2 residues is that for H1 HA2. Conservative amino acid substitu-

tions refer to the interchangeability of residues having similar side chains. For example, a group of amino acids having aliphatic side chains is glycine, alanine, valine, leucine, and isoleucine; a group of amino acids having aliphatic-hydroxyl side chains is serine and threonine; a group of amino acids having amide-containing side chains is asparagine and glutamine; a group of amino acids having aromatic side chains is phenylalanine, tyrosine and tryptophan; a group of amino acids having basic side chains is lysine, arginine and histidine; and a group of amino acids having sulfur-containing side chain is cysteine and methionine. In one embodiment, conservative amino acid substitution groups are: threonine-valine-leucine-isoleucine-alanine; phenylalanine-tyrosine; lysine-arginine; alanine-valine; glutamic-aspartic; and asparagine-glutamine.

In one embodiment, a mutation is introduced into a HA gene segment of an influenza virus isolate, e.g., via recombinant DNA techniques including site-specific mutagenesis or replacing a portion of the HA coding sequence that includes residue 117 of HA2 with a portion that includes the characteristic residue(s), wherein the numbering for HA2 residues is that for H1 HA2.

In another embodiment, a HA gene segment with a residue that confers enhanced replication in Vero cells is combined with a compatible NA segment, and internal gene segments of an influenza vaccine virus. In one embodiment, the substitution(s) in the HA2 protein, or the characteristic residue in the HA2 protein, that results in the enhanced replication, is/are at or within about 1 to 10 residues, or any integer in between, for instance, at or within 1 to 5, residues, of residue 117 of the HA2 protein of influenza A virus, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, a NA protein has at least one substitution, or has the characteristic residue discussed herein, such as one that results in enhanced replication, at or within about 1 to 10 residues, or any integer in between, e.g., at or within 1 to 5 residues of the codon for residue 255 of the NA protein of influenza A virus, wherein the numbering for NA residues is that for N1.

The invention provides a plurality of influenza virus vectors of the invention, e.g., those useful to prepare reassortant viruses including 6:1:1 reassortants, 6:2 reassortants and 7:1 reassortants. A 6:1:1 reassortant within the scope of the present invention is an influenza virus with 6 internal gene segments from a vaccine virus, a NA gene segment from a different (second) viral isolate, and a HA gene segment with a characteristic residue or substitution at position 117 of HA2 as described herein, where the HA gene segment is from a different viral source than the vaccine virus or the first viral isolate; a 6:2 reassortant within the scope of the present invention is an influenza virus with 6 internal gene segments from a vaccine virus, and a NA gene segment and a HA gene segment from a different (second) viral isolate, where the HA gene segment has the characteristic residue or a substitution at position 117 of HA2 as described herein; and a 7:1 reassortant within the scope of the present invention is an influenza virus with 6 internal gene segments and a NA gene segment from a vaccine virus, and a HA gene segment with a characteristic residue or substitution at position 117 of HA2 as described herein, where the HA gene segment is from a different viral source than the vaccine virus, or an influenza virus with 6 internal gene segments and a HA gene segment with the characteristic residue or substitution at position 117 of HA2 as described herein, and a NA gene segment is from a different viral source than the vaccine virus.

In one embodiment of the invention, the plurality includes vectors for vRNA production selected from a vector comprising a promoter operably linked to an influenza virus PA DNA

linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus PB1 DNA linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus PB2 DNA linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus HA DNA linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus NP DNA linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus NA DNA linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus M DNA linked to a transcription termination sequence, and a vector comprising a promoter operably linked to an influenza virus NS DNA linked to a transcription termination sequence. In one embodiment, the DNAs for vRNA production of PB1, PB2, PA, NP, M, and NS, have sequences from an influenza virus that replicates to high titers in cultured mammalian cells such as Vero cells or PER.C6® cells and also optionally embryonated eggs, and/or from a vaccine virus, e.g., one that does not cause significant disease in humans. The DNA for vRNA production of NA may be from any NA, e.g., any of N1-N9, and the DNA for vRNA production of HA may be from any HA, e.g., H1-H16. In one embodiment, the DNAs for vRNA production may be for an influenza B or C virus. For example, the DNAs for vRNA production include influenza B virus PA, PB1, PB2, NP, NS, and M or influenza B virus PA, PB1, PB2, NP, NS, M, and NA, wherein the vRNA for HA has a HA2 with a characteristic amino acid at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. The DNAs for vRNA production of NA and HA may be from different strains or isolates (6:1:1 reassortants) or from the same strain or isolate (6:2 reassortants), or the NA may be from the same strain or isolate as that for the internal genes (7:1 reassortant), where the HA2 sequence is selected to result in enhanced replication in Vero cells relative to a corresponding virus with, for example, an asparagine at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. The plurality also includes vectors for mRNA production selected from a vector encoding influenza virus PA, a vector encoding influenza virus PB1, a vector encoding influenza virus PB2, and a vector encoding influenza virus NP, and optionally one or more vectors encoding NP, NS, M, e.g., M1 and M2, HA or NA. The vectors encoding viral proteins may further include a transcription termination sequence.

Viruses that may provide the internal genes for reassortants within the scope of the invention include viruses that have high titers in Vero cells, e.g., titers of at least about 10^5 PFU/mL, e.g., at least 10^6 PFU/mL, 10^7 PFU/mL or 10^8 PFU/mL; high titers in embryonated eggs, e.g., titers of at least about 10^7 EID₅₀/mL, e.g., at least 10^8 EID₅₀/mL, 10^9 EID₅₀/mL or 10^{10} EID₅₀/mL; high titers in MDCK cells, e.g., titers of at least about 10^7 PFU/mL, e.g., at least 10^8 PFU/mL, or high titers in two of more of those host cells.

In one embodiment, the titers of the reassortant viruses of the invention in cells such as Vero cells may be over 1 log, 2 logs, 3 logs, or greater, than titers of the corresponding virus without a HA2 substitution or that lacks the selected residue at position 117 of HA2, wherein the numbering for HA2 residues is that for H1 HA2.

Other reassortants with internal genes from other PR8 isolates or vaccine viruses may be employed in recombinant reassortant viruses of the invention. In particular, 5:1:2 reassortants having PR8(UW) PB1, PB2, PA, NP, and M ("5") and PR8(Cam) NS ("1"); 6:1:1 reassortants having PR8(UW) NA, PB1, PB2, PA, NP, and M ("6") and PR8(Cam) NS ("1");

and 7:1 reassortants having PR8(UW) PB1, PB2, PA, NP, M, NA, and NS ("7") may be employed.

In one embodiment, the DNAs for the internal genes for PB1, PB2, PA, NP, M, and NS encode proteins with substantially the same activity as a corresponding polypeptide encoded by one of SEQ ID NOs:1-6 or 10-15. As used herein, "substantially the same activity" includes an activity that is about 0.1%, 1%, 10%, 30%, 50%, 90%, e.g., up to 100% or more, or detectable protein level that is about 80%, 90% or more, the activity or protein level, respectively, of the corresponding full-length polypeptide. In one embodiment, the nucleic acid a sequence encoding a polypeptide which is substantially the same as, e.g., having at least 80%, e.g., 90%, 92%, 95%, 97% or 99%, including any integer between 80 and 99, contiguous amino acid sequence identity to, a polypeptide encoded by one of SEQ ID NOs:1-6 or 10-15. In one embodiment, the isolated and/or purified nucleic acid molecule comprises a nucleotide sequence which is substantially the same as, e.g., having at least 50%, e.g., 60%, 70%, 80% or 90%, including any integer between 50 and 100, or more contiguous nucleic acid sequence identity to one of SEQ ID NOs:1-6 or 10-15 or and, in one embodiment, also encodes a polypeptide having at least 80%, e.g., 90%, 92%, 95%, 97% or 99%, including any integer between 80 and 99, contiguous amino acid sequence identity to a polypeptide encoded by one of SEQ ID NOs:1-6 or 10-15. In one embodiment, the influenza virus polypeptide has one or more, for instance, 2, 5, 10, 15, 20 or more, conservative amino acids substitutions, e.g., conservative substitutions of up to 10% or 20% of the residues, relative to a polypeptide encoded by one of SEQ ID NOs:1-6 or 10-15. Conservative amino acid substitutions refer to the interchangeability of residues having similar side chains. For example, a group of amino acids having aliphatic side chains is glycine, alanine, valine, leucine, and isoleucine; a group of amino acids having aliphatic-hydroxyl side chains is serine and threonine; a group of amino acids having amide-containing side chains is asparagine and glutamine; a group of amino acids having aromatic side chains is phenylalanine, tyrosine and tryptophan; a group of amino acids having basic side chains is lysine, arginine and histidine; and a group of amino acids having sulfur-containing side chain is cysteine and methionine. In one embodiment, conservative amino acid substitution groups are: valine-leucine-isoleucine; phenylalanine-tyrosine; lysine-arginine; alanine-valine; glutamic-aspartic; and asparagine-glutamine. In one embodiment, the influenza virus polypeptide has one or more, for instance, 2, 3 or 4, nonconservative amino acid substitutions, relative to a polypeptide encoded by one of SEQ ID NOs:1-6 or 10-15.

The invention thus includes the use of isolated and purified vectors or plasmids, which express or encode influenza virus proteins, or express or encode influenza vRNA, both native and recombinant vRNA. The vectors comprise influenza cDNA, e.g., influenza A (e.g., any influenza A gene including any of the 16 HA or 9 NA subtypes), B or C DNA (see Fields *Virology* (Fields et al. (eds.), Lippincott, Williams and Wilkins (2006), which is specifically incorporated by reference herein). Any suitable promoter or transcription termination sequence may be employed to express a protein or peptide, e.g., a viral protein or peptide, a protein or peptide of a nonviral pathogen, or a therapeutic protein or peptide.

A composition or plurality of vectors of the invention may also comprise a heterologous gene or open reading frame of interest, e.g., a foreign gene encoding an immunogenic peptide or protein useful as a vaccine or in gene replacement, for instance may encode an epitope useful in a cancer therapy or vaccine, or a peptide or polypeptide useful in gene therapy.

When preparing virus, the vector or plasmid comprising the gene or cDNA of interest may substitute for a vector or plasmid for an influenza viral gene or may be in addition to vectors or plasmids for all influenza viral genes. Thus, another embodiment of the invention comprises a composition or plurality of vectors as described above in which one of the vectors is replaced with, or further comprises, 5' influenza virus sequences optionally including 5' influenza virus coding sequences or a portion thereof, linked to a desired nucleic acid sequence, e.g., a desired cDNA, linked to 3' influenza virus sequences optionally including 3' influenza virus coding sequences or a portion thereof. In one embodiment, the desired nucleic acid sequence such as a cDNA is in an anti-sense (antigenomic) orientation. The introduction of such a vector in conjunction with the other vectors described above to a host cell permissive for influenza virus replication results in recombinant virus comprising vRNA corresponding to the heterologous sequences of the vector.

The promoter in a vector for vRNA production may be a RNA polymerase I promoter, a RNA polymerase II promoter, a RNA polymerase III promoter, a T7 promoter, or a T3 promoter, and optionally the vector comprises a transcription termination sequence such as a RNA polymerase I transcription termination sequence, a RNA polymerase II transcription termination sequence, a RNA polymerase III transcription termination sequence, or a ribozyme. Ribozymes within the scope of the invention include, but are not limited to, tetrahymena ribozymes, RNase P, hammerhead ribozymes, hairpin ribozymes, hepatitis ribozyme, as well as synthetic ribozymes. In one embodiment, the RNA polymerase I promoter is a human RNA polymerase I promoter.

The promoter or transcription termination sequence in a vRNA or virus protein expression vector may be the same or different relative to the promoter or any other vector. In one embodiment, the vector or plasmid which expresses influenza vRNA comprises a promoter suitable for expression in at least one particular host cell, e.g., avian or mammalian host cells such as canine, feline, equine, bovine, ovine, or primate cells including human cells, or for expression in more than one host.

In one embodiment, at least one vector for vRNA comprises a RNA polymerase II promoter linked to a ribozyme sequence linked to viral coding sequences linked to another ribozyme sequences, optionally linked to a RNA polymerase II transcription termination sequence. In one embodiment, at least 2, e.g., 3, 4, 5, 6, 7 or 8, vectors for vRNA production comprise a RNA polymerase II promoter, a first ribozyme sequence, which is 5' to a sequence corresponding to viral sequences including viral coding sequences, which is 5' to a second ribozyme sequence, which is 5' to a transcription termination sequence. Each RNA polymerase II promoter in each vRNA vector may be the same or different as the RNA polymerase II promoter in any other vRNA vector. Similarly, each ribozyme sequence in each vRNA vector may be the same or different as the ribozyme sequences in any other vRNA vector. In one embodiment, the ribozyme sequences in a single vector are not the same.

In one embodiment, the invention provides a plurality of influenza virus vectors for a reassortant, comprising a vector for vRNA production comprising a promoter operably linked to an influenza virus PA DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus PB1 DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus PB2 DNA linked to a transcription termination sequence, a vector for vRNA production comprising a

promoter operably linked to an influenza virus HA DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus NP DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus NA DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus M DNA linked to a transcription termination sequence, and a vector for vRNA production comprising a promoter operably linked to an influenza virus NS cDNA linked to a transcription termination sequence, wherein the DNAs for PB1, PB2, PA, NP, NS, and M from one or more influenza vaccine seed viruses, wherein the DNA for NA has sequences for a heterologous NA, and wherein the DNA for HA selected to encode an aspartic acid or glutamic acid at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2; and a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus PA, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus PB1, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus PB2, and a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus NP, and optionally a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus HA, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus NA, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus M1, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus M2, or a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus NS2. In one embodiment, at least one vector comprises sequences corresponding to those encoding PB1, PB2, PA, NP, M, or NS, or a portion thereof, having substantially the same activity as a corresponding polypeptide encoded by one of SEQ ID NOs: 1-6 or 10-15, e.g., a sequence encoding a polypeptide with at least 80%, e.g., 85%, 90%, 92%, 95%, 98%, 99% or 100%, including any integer between 80 and 100, amino acid identity to a polypeptide encoded by one of SEQ ID NOs: 1-6 or 10-15. Optionally, two vectors may be employed in place of the vector comprising a promoter operably linked to an influenza virus M cDNA linked to a transcription termination sequence, e.g., a vector comprising a promoter operably linked to an influenza virus M1 cDNA linked to a transcription termination sequence and a vector comprising a promoter operably linked to an influenza virus M2 cDNA linked to a transcription termination sequence.

A plurality of the vectors of the invention may be physically linked or each vector may be present on an individual plasmid or other, e.g., linear, nucleic acid delivery vehicle. In one embodiment, each vRNA production vector is on a separate plasmid. In one embodiment, each mRNA production vector is on a separate plasmid.

The invention also provides a method to prepare influenza virus. The method comprises contacting a cell with a plurality of the vectors of the invention, e.g., sequentially or simultaneously, in an amount effective to yield infectious influenza virus. The invention also includes isolating virus from a cell contacted with the plurality of vectors. Thus, the invention further provides isolated virus, as well as a host cell contacted with the plurality of vectors or virus of the invention. In another embodiment, the invention includes contacting the

cell with one or more vectors, either vRNA or protein production vectors, prior to other vectors, either vRNA or protein production vectors. In one embodiment, the promoter for vRNA vectors employed in the method is a RNA polymerase I promoter, a RNA polymerase II promoter, a RNA polymerase III promoter, a T3 promoter or a T7 promoter. In one embodiment, the RNA polymerase I promoter is a human RNA polymerase I promoter. In one embodiment, each vRNA vector employed in the method is on a separate plasmid. In one embodiment, the vRNA vectors employed in the method are on one plasmid or on two or three different plasmids. In one embodiment, each mRNA vector employed in the method is on a separate plasmid. In one embodiment, the mRNA vectors for PA, PB1, PB2 and NP employed in the method are on one plasmid or on two or three different plasmids.

In one embodiment, the invention provides a method to select for influenza viruses with enhanced replication in cell culture. The method includes providing cells suitable for influenza vaccine production; serially culturing one or more influenza virus isolates in the cells; and isolating serially cultured virus with enhanced growth relative to the one or more isolates prior to serial culture. In one embodiment, the cells are rodent or primate cells.

Also provided is a method to identify a HA2 that confers altered growth of a recombinant influenza virus. The method includes introducing one or more substitutions in influenza virus HA2 into a HA gene segment to yield a mutant HA gene segment; and identifying whether the mutant HA gene segment, when present in a replication competent recombinant influenza virus, results in enhanced replication of the recombinant influenza virus in a cell relative to a corresponding replication competent influenza virus without the one or more substitutions in HA2. In one embodiment, at least one substitution is at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2, e.g., at least one substitution is to aspartic acid or glutamic acid. In one embodiment, the cell is a rodent or primate cell. In one embodiment, the one or more substitutions are to an amino acid residue with an acidic side chain.

In one embodiment, the invention provides a method to prepare a recombinant influenza virus with a HA gene segment having a mutant HA2. The method includes altering influenza virus HA nucleic acid at position 117 in HA2 to aspartic acid or glutamic acid; and expressing the altered nucleic acid in a cell having vectors for influenza vRNA production and viral protein production in an amount effective to yield recombinant influenza virus with a HA gene segment having the aspartic acid or glutamic acid at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2. In one embodiment, the cell is a mammalian, e.g., a human cell, or avian cell.

The methods of producing virus described herein, which do not require helper virus infection, are useful in viral mutagenesis studies, and in the production of vaccines (e.g., for AIDS, influenza, hepatitis B, hepatitis C, rhinovirus, filoviruses, malaria, herpes, and foot and mouth disease) and gene therapy vectors (e.g., for cancer, AIDS, adenosine deaminase, muscular dystrophy, ornithine transcarbamylase deficiency and central nervous system tumors). Thus, a virus for use in medical therapy (e.g., for a vaccine or gene therapy) is provided.

The invention also provides isolated viral polypeptides, and methods of preparing and using recombinant virus of the invention. The methods include administering to a host organism, e.g., a mammal, an effective amount of the influenza virus of the invention, e.g., an inactivated virus prepa-

ration, optionally in combination with an adjuvant and/or a carrier, e.g., in an amount effective to prevent or ameliorate infection of an animal such as a mammal by that virus or an antigenically closely related virus. In one embodiment, the virus is administered intramuscularly while in another embodiment, the virus is administered intranasally. In some dosing protocols, all doses may be administered intramuscularly or intranasally, while in others a combination of intramuscular and intranasal administration is employed. The vaccine may further contain other isolates of influenza virus including recombinant influenza virus, other pathogen(s), additional biological agents or microbial components, e.g., to form a multivalent vaccine. In one embodiment, intranasal vaccination, for instance containing with inactivated influenza virus, and a mucosal adjuvant may induce virus-specific IgA and neutralizing antibody in the nasopharynx as well as serum IgG.

The influenza virus of the invention may employed with other anti-virals, e.g., amantadine, rimantadine, and/or neuraminidase inhibitors, e.g., may be administered separately in conjunction with those anti-virals, for instance, administered before, during and/or after.

The invention also provides a method in which the pH of media in which cells suitable for propagating influenza virus are cultured, is altered during virus propagation to allow for enhanced influenza virus replication in those cells. Thus, for cells with late endosomes having a pH that is higher than that in MDCK cells, altering media pH to maintain a higher pH during virus replication over time, may enhance virus production in the absence of a HA2 protein with a characteristic residue, such as aspartic acid, at position 117, wherein the numbering for HA2 residues is that for H1 HA2.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1. Nucleotide sequence for PR8(Cambridge) genes (SEQ ID NOs:10-15).

FIG. 2. Growth properties of Vero cell-adapted PR8 virus in Vero cells.

FIG. 3. Comparison of amino acid sequence differences between PR8 and Vero cell-adapted PR8.

FIG. 4. Growth properties of Vero cell-adapted PR8, non Vero cell-adapted "wild-type" PR8, and recombinant viruses with one or two substitutions relative to wild-type virus in Vero cells.

FIG. 5. Growth properties of HA2 N117D virus and wild-type PR8 in MDCK cells.

FIG. 6. Three dimensional structure of HA as a trimer (A), HA as a monomer (B) and HA2 (C).

FIG. 7. Schematic of fusion assay which expresses full length HA.

FIG. 8. Photomicrographs of Vero cells expressing wild-type PR8HA or HA2 N117D virus at various pH conditions.

FIGS. 9A-B. pH sensitivity of Alexa647 and Oregon Green dyes. A) The fluorescence intensity of Oregon Green dye is sensitive to variations in pH while the fluorescence intensity of Alexa647 does not vary over pH 3 to 7. B) Schematic of assay to detect endosomal pH.

FIG. 10. Comparison of endosomal pH in MDCK cells and Vero cells.

FIGS. 11A-C. HA2 N117D substitution mutants have enhanced infectivity titers in Vero cells. A) Vero cells were infected with A/Kawasaki/173/2001 (H1N1) and A/Kawasaki/173/2001 HA2 N117D and the titers over time determined. B) Vero cells were infected with A/Kawasaki/UTK-4/2009 (H1N1) and A/Kawasaki/UKT-4/2009 HA2 N117D and the titers over time determined. C) Vero cells were

infected with A/Yokohama/2017/2003 (H3N2) and A/Yokohama/2017/2003 HA2 N116D and the titers over time determined.

FIG. 12. A) Alignment of HA2 sequences from A/Aichi/2/68; A/Dk/Sing/97; A/HK/486/97; A/Sw/9/98; and A/HongKong/1073/99 (SEQ ID Nos.16-20 and 23-27). B) Amino acid sequence of HA sequence from A/California/08/2009 (SEQ ID NO:21). HA2 sequences correspond to residues 336-566 (SEQ ID NO:22)

FIG. 13. HA2 sequences for A/Kawasaki/173/2001, A/Kawasaki/UKT-4/2009, and A/Yokohama/2017/2003 (SEQ ID NOs:28-30). According to the NCBI database, influenza virus HA2 sequences for H1, H2, H3, H5, H7, and H9 HAs were generally conserved at position 116 or 117 (N116 or N117) (more than 99%).

DETAILED DESCRIPTION OF THE INVENTION

Definitions

As used herein, the term “isolated” refers to in vitro preparation and/or isolation of a nucleic acid molecule, e.g., vector or plasmid, peptide or polypeptide (protein), or virus of the invention, so that it is not associated with in vivo substances, or is substantially purified from in vitro substances. An isolated virus preparation is generally obtained by in vitro culture and propagation, and/or via passage in eggs, and is substantially free from other infectious agents.

As used herein, “substantially purified” means the object species is the predominant species, e.g., on a molar basis it is more abundant than any other individual species in a composition, and preferably is at least about 80% of the species present, and optionally 90% or greater, e.g., 95%, 98%, 99% or more, of the species present in the composition.

As used herein, “substantially free” means below the level of detection for a particular infectious agent using standard detection methods for that agent.

A “recombinant” virus is one which has been manipulated in vitro, e.g., using recombinant DNA techniques, to introduce changes to the viral genome. Reassortant viruses can be prepared by recombinant or nonrecombinant techniques.

As used herein, the term “recombinant nucleic acid” or “recombinant DNA sequence or segment” refers to a nucleic acid, e.g., to DNA, that has been derived or isolated from a source, that may be subsequently chemically altered in vitro, so that its sequence is not naturally occurring, or corresponds to naturally occurring sequences that are not positioned as they would be positioned in the native genome. An example of DNA “derived” from a source, would be a DNA sequence that is identified as a useful fragment, and which is then chemically synthesized in essentially pure form. An example of such DNA “isolated” from a source would be a useful DNA sequence that is excised or removed from said source by chemical means, e.g., by the use of restriction endonucleases, so that it can be further manipulated, e.g., amplified, for use in the invention, by the methodology of genetic engineering.

As used herein, a “heterologous” influenza virus gene or gene segment is from an influenza virus source that is different than a majority of the other influenza viral genes or gene segments in a recombinant, e.g., reassortant, influenza virus.

The terms “isolated polypeptide”, “isolated peptide” or “isolated protein” include a polypeptide, peptide or protein encoded by cDNA or recombinant RNA including one of synthetic origin, or some combination thereof.

The term “recombinant protein” or “recombinant polypeptide” as used herein refers to a protein molecule expressed from a recombinant DNA molecule. In contrast, the term

“native protein” is used herein to indicate a protein isolated from a naturally occurring (i.e., a nonrecombinant) source. Molecular biological techniques may be used to produce a recombinant form of a protein with identical properties as compared to the native form of the protein.

Methods of alignment of sequences for comparison are well known in the art. Thus, the determination of percent identity between any two sequences can be accomplished using a mathematical algorithm.

Computer implementations of these mathematical algorithms can be utilized for comparison of sequences to determine sequence identity. Alignments using these programs can be performed using the default parameters. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). The algorithm may involve first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold. These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0) and N (penalty score for mismatching residues; always <0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when the cumulative alignment score falls off by the quantity X from its maximum achieved value, the cumulative score goes to zero or below due to the accumulation of one or more negative-scoring residue alignments, or the end of either sequence is reached.

In addition to calculating percent sequence identity, the BLAST algorithm may also perform a statistical analysis of the similarity between two sequences. One measure of similarity provided by the BLAST algorithm may be the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a test nucleic acid sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid sequence to the reference nucleic acid sequence is less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

The BLASTN program (for nucleotide sequences) may use as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program may use as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix. See <http://www.ncbi.nlm.nih.gov>. Alignment may also be performed manually by inspection.

For sequence comparison, typically one sequence acts as a reference sequence to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are input into a computer, subsequence coordinates are designated if necessary, and sequence algorithm program parameters are designated. The sequence comparison algorithm then calculates the percent sequence identity for the test sequence(s) relative to the reference sequence, based on the designated program parameters.

Influenza Virus Structure and Propagation

Influenza A viruses possess a genome of eight single-stranded negative-sense viral RNAs (vRNAs) that encode at least ten proteins. The influenza virus life cycle begins with binding of the hemagglutinin (HA) to sialic acid-containing receptors on the surface of the host cell, followed by receptor-mediated endocytosis. The low pH in late endosomes triggers a conformational shift in the HA, thereby exposing the N-terminus of the HA2 subunit (the so-called fusion peptide). The fusion peptide initiates the fusion of the viral and endosomal membrane, and the matrix protein (M1) and RNP complexes are released into the cytoplasm. RNPs consist of the nucleoprotein (NP), which encapsidates vRNA, and the viral polymerase complex, which is formed by the PA, PB1, and PB2 proteins. RNPs are transported into the nucleus, where transcription and replication take place. The RNA polymerase complex catalyzes three different reactions: synthesis of an mRNA with a 5' cap and 3' polyA structure, of a full-length complementary RNA (cRNA), and of genomic vRNA using the cRNA as a template. Newly synthesized vRNAs, NP, and polymerase proteins are then assembled into RNPs, exported from the nucleus, and transported to the plasma membrane, where budding of progeny virus particles occurs. The neuraminidase (NA) protein plays a crucial role late in infection by removing sialic acid from sialyloligosaccharides, thus releasing newly assembled virions from the cell surface and preventing the self aggregation of virus particles. Although virus assembly involves protein-protein and protein-vRNA interactions, the nature of these interactions is largely unknown.

Although influenza B and C viruses are structurally and functionally similar to influenza A virus, there are some differences. For example, influenza B virus does not have a M2 protein with ion channel activity but has BM2 and has a gene segment with both NA and NB sequences. Influenza C virus has only seven gene segments.

Cell Lines that can be Used in the Present Invention

Any cell, e.g., any avian or mammalian cell, such as a human, e.g., 293T or PER.C6® cells, or canine, bovine, equine, feline, swine, ovine, rodent, for instance mink, e.g., MvLu1 cells, or hamster, e.g., CHO cells, or non-human primate, e.g., Vero cells, including mutant cells, which supports efficient replication of influenza virus can be employed to isolate and/or propagate influenza viruses. Isolated viruses can be used to prepare a reassortant virus. In one embodiment, host cells for vaccine production are continuous mammalian or avian cell lines or cell strains. A complete characterization of the cells to be used, may be conducted so that appropriate tests for purity of the final product can be included. Data that can be used for the characterization of a cell includes (a) information on its origin, derivation, and passage history; (b) information on its growth and morphological characteristics; (c) results of tests of adventitious agents; (d) distinguishing features, such as biochemical, immunological, and cytogenetic patterns which allow the cells to be clearly recognized among other cell lines; and (e) results of tests for tumorigenicity. In one embodiment, the passage level, or population doubling, of the host cell used is as low as possible.

In one embodiment, the cells are WHO certified, or certifiable, continuous cell lines. The requirements for certifying such cell lines include characterization with respect to at least one of genealogy, growth characteristics, immunological markers, virus susceptibility tumorigenicity and storage conditions, as well as by testing in animals, eggs, and cell culture. Such characterization is used to confirm that the cells are free from detectable adventitious agents. In some countries, karyology may also be required. In addition, tumorigenicity may

be tested in cells that are at the same passage level as those used for vaccine production. The virus may be purified by a process that has been shown to give consistent results, before vaccine production (see, e.g., World Health Organization, 1982).

Virus produced by the host cell may be highly purified prior to vaccine or gene therapy formulation. Generally, the purification procedures result in extensive removal of cellular DNA and other cellular components, and adventitious agents. Procedures that extensively degrade or denature DNA may also be used.

Influenza Vaccines

A vaccine of the invention includes an isolated recombinant influenza virus of the invention, and optionally one or more other isolated viruses including other isolated influenza viruses, one or more immunogenic proteins or glycoproteins of one or more isolated influenza viruses or one or more other pathogens, e.g., an immunogenic protein from one or more bacteria, non-influenza viruses, yeast or fungi, or isolated nucleic acid encoding one or more viral proteins (e.g., DNA vaccines) including one or more immunogenic proteins of the isolated influenza virus of the invention. In one embodiment, the influenza viruses of the invention may be vaccine vectors for influenza virus or other pathogens.

A complete virion vaccine may be concentrated by ultrafiltration and then purified by zonal centrifugation or by chromatography. Viruses other than the virus of the invention, such as those included in a multivalent vaccine, may be inactivated before or after purification using formalin or beta-propiolactone, for instance.

A subunit vaccine comprises purified glycoproteins. Such a vaccine may be prepared as follows: using viral suspensions fragmented by treatment with detergent, the surface antigens are purified, by ultracentrifugation for example. The subunit vaccines thus contain mainly HA protein, and also NA. The detergent used may be cationic detergent for example, such as hexadecyl trimethyl ammonium bromide (Bachmeyer, 1975), an anionic detergent such as ammonium deoxycholate (layer & Webster, 1976); or a nonionic detergent such as that commercialized under the name TRITON X100. The hemagglutinin may also be isolated after treatment of the virions with a protease such as bromelain, and then purified. The subunit vaccine may be combined with an attenuated virus of the invention in a multivalent vaccine.

A split vaccine comprises virions which have been subjected to treatment with agents that dissolve lipids. A split vaccine can be prepared as follows: an aqueous suspension of the purified virus obtained as above, inactivated or not, is treated, under stirring, by lipid solvents such as ethyl ether or chloroform, associated with detergents. The dissolution of the viral envelope lipids results in fragmentation of the viral particles. The aqueous phase is recuperated containing the split vaccine, constituted mainly of hemagglutinin and neuraminidase with their original lipid environment removed, and the core or its degradation products. Then the residual infectious particles are inactivated if this has not already been done. The split vaccine may be combined with an attenuated virus of the invention in a multivalent vaccine.

Inactivated Vaccines.

Inactivated influenza virus vaccines are provided by inactivating replicated virus using known methods, such as, but not limited to, formalin or β -propiolactone treatment. Inactivated vaccine types that can be used in the invention can include whole-virus (WV) vaccines or subvirion (SV) (split) vaccines. The WV vaccine contains intact, inactivated virus, while the SV vaccine contains purified virus disrupted with

detergents that solubilize the lipid-containing viral envelope, followed by chemical inactivation of residual virus.

In addition, vaccines that can be used include those containing the isolated HA and NA surface proteins, which are referred to as surface antigen or subunit vaccines.

Live Attenuated Virus Vaccines.

Live, attenuated influenza virus vaccines, such as those including a recombinant virus of the invention can be used for preventing or treating influenza virus infection. Attenuation may be achieved in a single step by transfer of attenuated genes from an attenuated donor virus to a replicated isolate or reassorted virus according to known methods. Since resistance to influenza A virus is mediated primarily by the development of an immune response to the HA and/or NA glycoproteins, the genes coding for these surface antigens come from the reassorted viruses or clinical isolates. The attenuated genes are derived from an attenuated parent. In this approach, genes that confer attenuation generally do not code for the HA and NA glycoproteins.

Viruses (donor influenza viruses) are available that are capable of reproducibly attenuating influenza viruses, e.g., a cold adapted (ca) donor virus can be used for attenuated vaccine production. Live, attenuated reassortant virus vaccines can be generated by mating the ca donor virus with a virulent replicated virus. Reassortant progeny are then selected at 25° C. (restrictive for replication of virulent virus), in the presence of an appropriate antiserum, which inhibits replication of the viruses bearing the surface antigens of the attenuated ca donor virus. Useful reassortants are: (a) infectious, (b) attenuated for seronegative non-adult mammals and immunologically primed adult mammals, (c) immunogenic and (d) genetically stable. The immunogenicity of the ca reassortants parallels their level of replication. Thus, the acquisition of the six transferable genes of the ca donor virus by new wild-type viruses has reproducibly attenuated these viruses for use in vaccinating susceptible mammals both adults and non-adult.

Other attenuating mutations can be introduced into influenza virus genes by site-directed mutagenesis to rescue infectious viruses bearing these mutant genes. Attenuating mutations can be introduced into non-coding regions of the genome, as well as into coding regions. Such attenuating mutations can also be introduced into genes other than the HA or NA, e.g., the PB2 polymerase gene. Thus, new donor viruses can also be generated bearing attenuating mutations introduced by site-directed mutagenesis, and such new donor viruses can be used in the production of live attenuated reassortants vaccine candidates in a manner analogous to that described above for the ca donor virus. Similarly, other known and suitable attenuated donor strains can be reassorted with influenza virus to obtain attenuated vaccines suitable for use in the vaccination of mammals.

In one embodiment, such attenuated viruses maintain the genes from the virus that encode antigenic determinants substantially similar to those of the original clinical isolates. This is because the purpose of the attenuated vaccine is to provide substantially the same antigenicity as the original clinical isolate of the virus, while at the same time lacking pathogenicity to the degree that the vaccine causes minimal chance of inducing a serious disease condition in the vaccinated mammal.

The viruses in a multivalent vaccine can thus be attenuated or inactivated, formulated and administered, according to known methods, as a vaccine to induce an immune response in an animal, e.g., a mammal. Methods are well-known in the art for determining whether such attenuated or inactivated vaccines have maintained similar antigenicity to that of the

clinical isolate or high growth strain derived therefrom. Such known methods include the use of antisera or antibodies to eliminate viruses expressing antigenic determinants of the donor virus; chemical selection (e.g., amantadine or rimantidine); HA and NA activity and inhibition; and nucleic acid screening (such as probe hybridization or PCR) to confirm that donor genes encoding the antigenic determinants (e.g., HA or NA genes) are not present in the attenuated viruses.

Pharmaceutical Compositions

Pharmaceutical compositions of the present invention, suitable for inoculation, e.g., nasal, parenteral or oral administration, comprise one or more influenza virus isolates, e.g., one or more attenuated or inactivated influenza viruses, a subunit thereof, isolated protein(s) thereof, and/or isolated nucleic acid encoding one or more proteins thereof, optionally further comprising sterile aqueous or non-aqueous solutions, suspensions, and emulsions. The compositions can further comprise auxiliary agents or excipients, as known in the art. The composition of the invention is generally presented in the form of individual doses (unit doses).

Conventional vaccines generally contain about 0.1 to 200 µg, e.g., 30 to 100 µg, of HA from each of the strains entering into their composition. The vaccine forming the main constituent of the vaccine composition of the invention may comprise a single influenza virus, or a combination of influenza viruses, for example, at least two or three influenza viruses, including one or more reassortant(s).

Preparations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and/or emulsions, which may contain auxiliary agents or excipients known in the art. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Carriers or occlusive dressings can be used to increase skin permeability and enhance antigen absorption. Liquid dosage forms for oral administration may generally comprise a liposome solution containing the liquid dosage form. Suitable forms for suspending liposomes include emulsions, suspensions, solutions, syrups, and elixirs containing inert diluents commonly used in the art, such as purified water. Besides the inert diluents, such compositions can also include adjuvants, wetting agents, emulsifying and suspending agents, or sweetening, flavoring, or perfuming agents.

When a composition of the present invention is used for administration to an individual, it can further comprise salts, buffers, adjuvants, or other substances which are desirable for improving the efficacy of the composition. For vaccines, adjuvants, substances which can augment a specific immune response, can be used. Normally, the adjuvant and the composition are mixed prior to presentation to the immune system, or presented separately, but into the same site of the organism being immunized.

Heterogeneity in a vaccine may be provided by mixing replicated influenza viruses for at least two influenza virus strains, such as 2-20 strains or any range or value therein. Vaccines can be provided for variations in a single strain of an influenza virus, using techniques known in the art.

A pharmaceutical composition according to the present invention may further or additionally comprise at least one chemotherapeutic compound, for example, for gene therapy, immunosuppressants, anti-inflammatory agents or immune enhancers, and for vaccines, chemotherapeutics including, but not limited to, gamma globulin, amantadine, guanidine, hydroxybenzimidazole, interferon-α, interferon-β, interferon-γ, tumor necrosis factor-alpha, thiosemicarbazones, methisazone, rifampin, ribavirin, a pyrimidine analog, a

purine analog, foscarnet, phosphonoacetic acid, acyclovir, dideoxynucleosides, a protease inhibitor, or ganciclovir.

The composition can also contain variable but small quantities of endotoxin-free formaldehyde, and preservatives, which have been found safe and not contributing to undesirable effects in the organism to which the composition is administered.

Pharmaceutical Purposes

The administration of the composition (or the antisera that it elicits) may be for either a "prophylactic" or "therapeutic" purpose. When provided prophylactically, the compositions of the invention which are vaccines are provided before any symptom or clinical sign of a pathogen infection becomes manifest. The prophylactic administration of the composition serves to prevent or attenuate any subsequent infection. When provided prophylactically, the gene therapy compositions of the invention, are provided before any symptom or clinical sign of a disease becomes manifest. The prophylactic administration of the composition serves to prevent or attenuate one or more symptoms or clinical signs associated with the disease.

When provided therapeutically, a viral vaccine is provided upon the detection of a symptom or clinical sign of actual infection. The therapeutic administration of the compound(s) serves to attenuate any actual infection. When provided therapeutically, a gene therapy composition is provided upon the detection of a symptom or clinical sign of the disease. The therapeutic administration of the compound(s) serves to attenuate a symptom or clinical sign of that disease.

Thus, a vaccine composition of the present invention may be provided either before the onset of infection (so as to prevent or attenuate an anticipated infection) or after the initiation of an actual infection. Similarly, for gene therapy, the composition may be provided before any symptom or clinical sign of a disorder or disease is manifested or after one or more symptoms are detected.

A composition is said to be "pharmacologically acceptable" if its administration can be tolerated by a recipient mammal. Such an agent is said to be administered in a "therapeutically effective amount" if the amount administered is physiologically significant. A composition of the present invention is physiologically significant if its presence results in a detectable change in the physiology of a recipient patient, e.g., enhances at least one primary or secondary humoral or cellular immune response against at least one strain of an infectious influenza virus.

The "protection" provided need not be absolute, i.e., the influenza infection need not be totally prevented or eradicated, if there is a statistically significant improvement compared with a control population or set of mammals. Protection may be limited to mitigating the severity or rapidity of onset of symptoms or clinical signs of the influenza virus infection.

Pharmaceutical Administration

A composition of the present invention may confer resistance to one or more pathogens, e.g., one or more influenza virus strains, by either passive immunization or active immunization. In active immunization, an attenuated live vaccine composition is administered prophylactically to a host (e.g., a mammal), and the host's immune response to the administration protects against infection and/or disease. For passive immunization, the elicited antisera can be recovered and administered to a recipient suspected of having an infection caused by at least one influenza virus strain. A gene therapy composition of the present invention may yield prophylactic or therapeutic levels of the desired gene product by active immunization.

In one embodiment, the vaccine is provided to a mammalian female (at or prior to pregnancy or parturition), under conditions of time and amount sufficient to cause the production of an immune response which serves to protect both the female and the fetus or newborn (via passive incorporation of the antibodies across the placenta or in the mother's milk).

The present invention thus includes methods for preventing or attenuating a disorder or disease, e.g., an infection by at least one strain of pathogen. As used herein, a vaccine is said to prevent or attenuate a disease if its administration results either in the total or partial attenuation (i.e., suppression) of a clinical sign or condition of the disease, or in the total or partial immunity of the individual to the disease. As used herein, a gene therapy composition is said to prevent or attenuate a disease if its administration results either in the total or partial attenuation (i.e., suppression) of a clinical sign or condition of the disease, or in the total or partial immunity of the individual to the disease.

A composition having at least one influenza virus of the present invention, including one which is attenuated and one or more other isolated viruses, one or more isolated viral proteins thereof, one or more isolated nucleic acid molecules encoding one or more viral proteins thereof, or a combination thereof, may be administered by any means that achieve the intended purposes.

For example, administration of such a composition may be by various parenteral routes such as subcutaneous, intravenous, intradermal, intramuscular, intraperitoneal, intranasal, oral or transdermal routes. Parenteral administration can be accomplished by bolus injection or by gradual perfusion over time.

A typical regimen for preventing, suppressing, or treating an influenza virus related pathology, comprises administration of an effective amount of a vaccine composition as described herein, administered as a single treatment, or repeated as enhancing or booster dosages, over a period up to and including between one week and about 24 months, or any range or value therein.

According to the present invention, an "effective amount" of a composition is one that is sufficient to achieve a desired effect. It is understood that the effective dosage may be dependent upon the species, age, sex, health, and weight of the recipient, kind of concurrent treatment, if any, frequency of treatment, and the nature of the effect wanted. The ranges of effective doses provided below are not intended to limit the invention and represent dose ranges.

The dosage of a live, attenuated or killed virus vaccine for an animal such as a mammalian adult organism may be from about 10^2 - 10^{15} , e.g., 10^3 - 10^{12} , plaque forming units (PFU)/kg, or any range or value therein. The dose of inactivated vaccine may range from about 0.1 to 1000, e.g., 30 to 100 µg, of HA protein. However, the dosage should be a safe and effective amount as determined by conventional methods, using existing vaccines as a starting point.

The dosage of immunoreactive HA in each dose of replicated virus vaccine may be standardized to contain a suitable amount, e.g., 30 to 100 µg or any range or value therein, or the amount recommended by government agencies or recognized professional organizations. The quantity of NA can also be standardized, however, this glycoprotein may be labile during purification and storage.

The dosage of immunoreactive HA in each dose of replicated virus vaccine can be standardized to contain a suitable amount, e.g., 1-50 µg or any range or value therein, or the amount recommended by the U.S. Public Health Service (PHS), which is usually 15 µg, per component for older children. 3 years of age, and 7.5 µg per component for older

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children <3 years of age. The quantity of NA can also be standardized, however, this glycoprotein can be labile during the processor purification and storage (Kendal et al., 1980; Kerr et al., 1975). Each 0.5-ml dose of vaccine may contain approximately 1-50 billion virus particles, and preferably 10 billion particles.

The invention will be described by the following nonlimiting examples.

Example 1

Methods

Cells and Viruses

293T human embryonic kidney cells are maintained in Dulbecco's modified Eagle's minimal essential medium (DMEM) with 10% fetal calf serum and antibiotics. Madin-Darby canine kidney (MDCK) cells are grown in MEM with 5% newborn calf serum and antibiotics. African green monkey Vero WCB cells, which had been established after biosafety tests for use in human vaccine production (Sugawara et al., 2002), are maintained in serum-free VP-SFM medium (GIBCO-BRL) with antibiotics. Cells are maintained at 37° C. in 5% CO₂. A WHO-recommended vaccine seed virus is NIBRG-14.

Construction of Plasmids and Reverse Genetics

To generate reassortants of influenza A viruses, a plasmid-based reverse genetics (Neumann et al., 1999) is used. The full-length cDNAs were cloned into a plasmid under control of the human polymerase I promoter and the mouse RNA polymerase I terminator (Poll plasmids).

A previously produced series of Poll constructs, derived from A/WSN/33 (H5N1; WSN) or PR8 strains is used, for reverse genetics (Horimoto et al., 2006; Neumann et al., 1999). The World Health Organization (WHO) recommends A/Puerto Rico/8/34 (H1N1; PR8) as a donor virus, because of its safety in humans (Wood & Robertson, 2004; Webby & Webster, 2003).

Plasmids expressing WSN or PR8NP, PA, PB1, or PB2 under control of the chicken β -actin promoter are used for all reverse genetics experiments (Horimoto et al., 2006; Neumann et al., 1999). Briefly, Poll plasmids and protein expression plasmids are mixed with a transfection reagent, Trans-IT 293T (Panvera), incubated at room temperature for 15 minutes, and then added to 293T cells. Transfected cells are incubated in Opti-MEM I (GIBCO-BRL) for 48 hours. For reverse genetics in Vero WCB cells, an electroporator (Amaxa) is used to transfect the plasmid mixtures according to the manufacturer's instructions. Sixteen hours after transfection, freshly prepared Vero WCB cells were added onto the transfected cells and TPCK-trypsin (1 μ g/mL) is added to the culture 6 hours later. Transfected cells are incubated in serum-free VP-SFM for a total of 4 days. Supernatants containing infectious viruses are harvested, and may be biologically cloned by limiting dilution.

A recombinant virus having the HA and NA genes from A/Hong Kong/213/2003 (H5N1) and the remainder of the type A influenza virus genes from PR8(UW) was prepared. The titer of the recombinant virus was 10^{10.67} EID₅₀/mL, and the HA titer was 1:1600

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TABLE 1

Virus possessing PR8 genes together with the following	HA titer (HAU/mL) in each dilution						
	10-2	10-3	10-4	10-5	10-6	10-7	10-8
HA and NA genes							
WSN-HA NA	160	40	40	320	40	640	<1
HK-HAavir NA	400	800	400	400	400	800	<1

The sequences of PR8 (UW) genes are as follows:

PA (SEQ ID NO: 1)

AGCGAAAGCA GGTACTGATC CAAATGGAA GATTTTGTGC
GACAATGCCT CAATCCGATG ATTGTCGAGC TTGCGGAAAA
AACAATGAAA GAGTATGGGG AGGACCTGAA AATCGAACA
AACAAATTTG CAGCAATATG CACTCACTTG GAAGTATGCT
TCATGTATTC AGATTTTCAC TTCATCAATG AGCAAGGCGA
GTCAATAATC GTAGAACTTG GTGATCCAAA TGCATTTTG
AAGCACAGAT TTGAAATAAT CGAGGGAAGA GATCGCACAA
TGGCCTGGAC AGTAGTAAAC AGTATTTGCA ACACACAGG
GGCTGAGAAA CCAAAGTTTC TACCAGATTG GTATGATTAC
AAGGAGAATA GATTTCATCGA AATTGGAGTA ACAAGGAGAG
AAGTTCACAT ATACTATCTG GAAAAGGCCA ATAAATTA
ATCTGAGAAA ACACACATCC ACATTTTCTC GTTCACTGGG
GAAGAAATGG CCACAAAGGC AGACTACACT CTCGATGAAG
AAAGCAGGGC TAGGATCAAA ACCGAGACTAT TCACCATAG
ACAAGAAATG GCCAGCAGAG GCCTCTGGGA TTCTTTTCGT
CAGTCCGAGA GAGGAGAAGA GACAATTGAA GAAAGGTTTG
AAATCACAGG AACAATGCGC AAGCTTGCCG ACCAAAGTCT
CCGCGCAAC TTCTCCAGCC TTGAAATTT TAGAGCCTAT
GTGGATGGAT TCGAACCGAA CGGCTACATT GAGGGCAAGC
TGCTCTCAAT GTCCAAAGAA GTAATGCTA GAATTGAACC
TTTTTTGAAA ACAACACCAC GACCACTGAT ACTTCCGAAT
GGGCTCCCT GTTCTCAGCG GTCCAAATTC CTGCTGATGG
ATGCTTAAAT ATTAAGCATT GAGGACCCAA GTCATGAAGG
AGAGGGAATA CCGCTATATG ATGCAATCAA ATGCATGAGA
ACATTCCTTG GATGGAAGGA ACCCAATGTT GTTAAACCAC
ACGAAAAGGG AATAAATCCA AATTATCTTC TGTCATGGAA
GCAAGTACTG GCAGAACTGC AGGACATTGA GATGAGGAG
AAAATTCCAA AGACTAAAAA TATGAAGAAA ACAAGTCAGC
TAAAGTGGGC ACTTGGTGAG AACATGGCAC CAGAAAAGGT
AGACTTTGAC GACTGTAAAG ATGTAGGTGA TTTGAAGCAA
TATGATAGTG ATGAACCAGA ATTGAGGTCT CTGCAAGTT
GGATTCAGAA TGAGTTTAAAC AAGGCATGCG AACGTACAGA
TTCAAGCTGG ATAGAGCTCG ATGAGATTGG AGAAGATGTG
GCTCCAATTG AACACATTGC AAGCATGAGA AGGAATTATT
TCACATCAGA GGTGTCTCAC TGCAGAGCCA CAGAAATACAT
AATGAAGGGA GTGTACATCA ATACTGCCTT GCTTAATGCA
TCTGTGCAG CAATGGATGA TTCCAATTA ATTCCAATGA
TAAGCAAGTG TAGAACTAAG GAGGGAAGGC GAAAGACCAA
CTTGATGGT TTCAATATAA AAGGAAGATC CCCTTAAGG
AATGACACCG ACGTGGTAAA CTTTGTGAGC ATGGAGTTTT
CTCTCACTGA CCCAAGACTT GAACCAACATA AATGGGAGAA
GTACTGTGTT CTTGAGATAG GAGATATGCT TATAAGAAAT
GCCATAGGCC AGGTTTCAAG GCCCATGTTC TTGTATGTGA
GAACAAATGG AACCTCAAAA ATTAATAATGA AATGGGGAAAT
GGAGATGAGG CGTTGCCTCC TCCAGTCACT TCAACAAATT
GAGAGTATGA TTGAAGCTGA GTCTCTGTCT AAAGAGAAAG
ACATGACCAA AGAGTTCTTT GAGAACAAAT CAGAAACATG
GCCCATGGGA GAGTCCCCCA AAGGAGTGGA GGAAGTTTCC
ATTGGGAAGG TCTGCAGGAC TTTATTAGCA AAGTCGGTAT
TCAACAGCTT GTATGCATCT CCACAACATG AAGGATTTTC
AGCTGAATCA AGAAAACCTG TTCTTATCGT TCAGGCTCTT
AGGGACAACC TGGAACTTGG GACCTTTGAT CTGGGGGGG
TATATGAAGC AATTGAGGAG TGCCTGATTA ATGATCCCTG
GGTTTTGCTT AATGCTTCTT GGTTCACACT CTTCCTTACA
CATGCATTGA GTTAGTTGTG GCAGTGCTAC TATTTGCTAT
CCATACTGTC CAAAAAAGTA CCTTGTTTCT ACT

PB1

(SEQ ID NO: 2)

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CCTTACTTTT CTTAAAAGTG CCAGCACAAA ATGCTATAAG
CACAACCTTC CCTTATACTG GAGACCTTCC TTACAGCCAT
GGGACAGGAA CAGGATACAC CATGGATACT GTCAACAGGA
CACATCAGTA CTCAGAAAAG GGAAGATGGA CAACAAACAC

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 ATTGTGTATT GGAGGCGATG GCTTTCCTTG AGGAATCCCA
 TCCTGGTATT TTTGAAAAC CGTGTATTGA AACGATGGAG
 GTTGTTCAGC AAACACGAGT AGACAAGCTG ACACAAGGCC
 GACAGACCTA TGA CTGGACT CTAAATAGAA ACCAACCTGC
 TGCAACAGCA TTGGCCAAACA CAATAGAAGT GTTCAGATCA
 AATGGCCTCA CGGCCAATGA GTCTGGAAGG CTCATAGACT
 TCCTTAAGGA TGTAAATGGAG TCAATGAACA AAGAAGAAAT
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 GTTGGAGGCA ATGAGAGAGG AGCAAAAGTTG GCAAAATGTTG
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 CAGAATCCTC GGATGTTTTT GGCCATGATC ACATATATGA
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 AGGCGTCTCC ATCCTGAATC TTGGACAAA AGATACACAC
 AAGACTACTT ACTGGTGGGA TGGTCTTCAA TCCTCTGACG
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 TCAAGCCGGA GTCCGACAGG TTTATCGAAC CTGTAAGCTA
 CTTGGAATCA ATATGAGCAA GAAAAAGTCT TACATAAACCA
 GAACAGGTAC ATTTGAATTC ACAAGTTTTT TCTATCGTTA
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 GGGGTGTCTG GGATCAACGA GTCAGCGGAC ATGAGTATTG
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PB2

(SEQ ID NO: 3)

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 GATACTCACA AAAACACCG TGGACCATAT GGCCATAATC
 AAGAAGTACA CATCAGGAAG ACAGGAGAAG AACCCAGCAC
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NP

(SEQ ID NO: 4)

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 TTGAGAGGGT CGGTTGCTCA CAACTCTGTC CTGCCTGCCT
 GTGTGATGAG ACCTGCCGTA GCCAGTGGGT ACGACTTTGA
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 CTGCTTCAAA ACAGCCAAGT GTACAGCTTA ATCAGACCAA
 ATGGAATCC AGCACACAAG AGTCAACTGG TGTGGATGGC
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 TTCATCAAAG GGACGAAGGT GCTCCCAAGA GGGAGCTTTT
 GACTATGGAA AGTTCAAAT GCTTCCAATG AAAATATGGA
 GACTATGGAA TCAAGTACAC TTGAAGTGA AAGCAGGTAC
 TGGGCCATAA GGACAGAGAAG TGGAGGAAAC ACCAATCAAC
 AGAGGGCATC TGCGGGCCAA ATCAGCATAAC AACCTACGTT
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 ACCAGAAGAT GTGTCTTTCC AGGGGCGGG AGTCTTCGAG
 CTCTCGGACG AAAAGGCGAG GAGCCCGATC GTGCCCTTCT
 TTGACATGAG TAATGAAGGA TCTTATTCTC TCGGAGACAA
 TGCAGAGGAG TACGACAAAT AAGAAAAAT ACCCTTGTGT
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65

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M (SEQ ID NO: 5)

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 CTAAGACAA GACCAATCCT GTCACCTCTG ACTAAGGGGA
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 TCTCTTTGAA AATTTGCAGG CCTATCAGAA ACGAATGGGG
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High-titer A/PR/8/34 (H1N1, PR8(UW)) virus grows 10 times better than other A/PR/8/34 PR8 strains in eggs (10^{10} EID₅₀/mL; HA titer: 1:8,000). Thus, replacement of the HA and NA genes of PR8(UW) with those of a currently circulating strain of influenza virus results in a vaccine strain that can be safely produced, and validates the use of PR8(UW) as a master vaccine strain.

Genes that contribute to different growth properties between PR8(UW) and PR8 (Cambridge), which provides the non-HA and -NA genes of the NIBRG-14 vaccine strain (FIG. 1), were determined. Higher titers in eggs were obtained when the majority of internal genes were from PR8 (UW). Highest titers were with the M gene segment of PR8 (UW) and the NS gene of PR8 (Cambridge). The NS gene in PR8(UW) has a K (lysine) at residue 55 while the NS gene in PR8(Cam) has a E (glutamic acid). The polymerase subunit (PA, PB1, and PB2) and NP genes of PR8(UW) enhanced the growth of an H5N1 vaccine seed virus in chicken embryonated eggs, and the NS gene of PR8(Cambridge) enhanced the growth of an H5N1 vaccine seed virus in chicken embryo-

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nated eggs. A tyrosine (Y) at position 360 in PB2 of PR8(UW) likely contributes to the high growth rate of that virus in MDCK cells.

Example 2

To establish robust systems for influenza vaccine production, egg-free, cell culture-based systems are needed. Vero cells are approved for human use and so are candidate hosts for influenza virus vaccine production. To elucidate the molecular basis for efficient growth of influenza vaccine seed virus in Vero cells, A/Puerto Rico/8/34 (PR8) virus was passaged through Vero cells 12 times and the infectivity titer of the resulting virus was determined. Vero cell-adapted PR8 had over a 4 log increase in infectivity titers relative to non Vero cell-adapted PR8 (FIG. 2).

To determine the molecular basis for that growth difference, the genomes of both isolates were sequenced. Three amino acid differences were found: one in HA2, one in NA and one in PB2 (FIG. 3). To identify the contribution of each individual substitution, and of a combination of two of the substitutions, recombinant viruses with the individual substitution(s) were prepared and the growth of those recombinant viruses was compared to Vero cell-adapted PR8 and non Vero cell-adapted PR8 (FIG. 4). The results indicated that the substitution in HA2 was primarily responsible for the enhanced growth in Vero cells. The substitution in HA2 (N117D) did not enhance growth in MDCK cells (FIG. 5).

Because HA2 has a fusion domain that is exposed after infection, a fusion assay was employed to compare the properties of wild-type PR8HA2 and HA2 N117D (FIGS. 7-8). The HA2 N117D mutant fused Vero cells at a higher pH than wild-type PR8. The endosomal pH in Vero cells and MDCK cells was determined using pH sensitive and insensitive dyes (FIGS. 9-10). The endosomes of Vero cells likely have a higher pH than those from MDCK cells. Thus, the HA2 N117D mutation may elevate the optimal pH for membrane fusion mediated by HA2, thereby enhancing virus replication efficiency in Vero cells.

To determine if the HA2 N117D mutation alone could enhance virus replication efficiency in different viruses in Vero cells, that substitution was introduced into two different H1N1 viruses (a AAT to GAT mutation) and one H3N2 virus (a AAC to GAC mutation) in a PR8 background (six gene segments were from Vero cell-adapted PR8; PA, PB1, PB2,

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M, NS and NP) (FIG. 11). The HA2 N117D mutation enhanced the replication efficiency of all three tested viruses in Vero cells. Such a strategy may be employed to prepare vaccine viruses with enhanced replication in Vero cells.

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- All publications, patents and patent applications are incorporated herein by reference. While in the foregoing specification this invention has been described in relation to certain preferred embodiments thereof, and many details have been set forth for purposes of illustration, it will be apparent to those skilled in the art that the invention is susceptible to additional embodiments and that certain of the details described herein may be varied considerably without departing from the basic principles of the invention.

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<400> SEQUENCE: 4

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agcaaaagca gggtagataa tcaactcactg agtgacatca aaatcatggc gtctcaaggc      60
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agagcatccg tcggaaaaat gattggtgga attggacgat tctacatcca aatgtgcacc      180
gaactcaaac tcagtgatta tgaggggacgg ttgatccaaa acagcttaac aatagagaga      240
atggtgctct ctgcttttga cgaaaggaga aataaatacc ttgaagaaca tcccagtgcg      300
gggaaagatc ctaagaaaac tggaggacct atatacagga gagtaaacgg aaagtggatg      360
agagaactca tcctttatga caaagaagaa ataaggcgaa tctggcgcca agctaataat      420
ggtgacgatg caacggctgg tctgactcac atgatgatct ggcatccaa tttgaatgat      480
gcaacttata agaggacaag agctcttggt cgcaccggaa tggatcccag gatgtgctct      540
ctgatgcaag gttcaactct ccctaggagg tctggagccg caggtgctgc agtcaaagga      600
gttggaacaa tggatgatga attggtcaga atgatcaaac gtgggatcaa tgatcggaac      660
ttctggaggg gtgagaatgg acgaaaaaca agaattgctt atgaaagaat gtgcaacatt      720
ctcaaaggga aatttcaaac tgtgcacaa aaagcaatga tggatcaagt gagagagagc      780
cggaaccag ggaatgctga gttcgaagat ctcacttttc tagcacggtc tgcactcata      840
ttgagagggt cggttgctca caagtcctgc ctgcctgcct gtgtgtatgg acctgccgta      900
gccagtgggt acgactttga aaggaggagg tactctctag tcggaataga ccctttcaga      960
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gcttccaatg aaaatatgga gactatggaa tcaagtacac ttgaactgag aagcaggtac     1200
tgggccataa ggaccagaag tggaggaaac accaatcaac agagggcatc tgcgggccaa     1260
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atggcagcat tcaatgggaa tacagagggg agaacatctg acatgaggac cgaaatcata     1380
aggatgatgg aaagtgaag accagaagat gtgtctttcc aggggcgggg agtcttcgag     1440
ctctcggacg aaaaggcagc gagcccgatc gtgccttcct ttgacatgag taatgaagga     1500
tcttatttct tcggagacaa tgcagaggag tacgacaatt aaagaaaaat accottgttt     1560
ctact                                             1565

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<210> SEQ ID NO 5
 <211> LENGTH: 1027
 <212> TYPE: DNA
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 5

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agcaaaagca ggtagatatt gaaagatgag tcttctaacc gaggtcgaaa cgtacgtact      60
ctctatcata ccgtcaggcc cctcctaaag cgagatcgca cagagacttg aagatgtctt     120
tgcagggaag aacaccgatc ttgaggttct catggaatgg ctaaagacaa gaccaatcct     180

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gtcacctctg actaagggga ttttaggatt tgtgttcacg ctcaccgtgc ccagtgagcg	240
aggactgcag cgtagacgct ttgtccaaaa tgccttaaat gggaacgggg atccaaataa	300
catggacaaa gcagttaaac tgtataggaa gctcaagagg gagataacat tccatggggc	360
caaagaaatc tcaactcagtt attctgctgg tgcacttgcc agttgtatgg gcctcatata	420
caacaggatg ggggctgtga ccactgaagt ggcatttggc ctggtatgtg caacctgtga	480
acagattgct gactcccagc atcgggtctca taggcaaatg gtgacaacaa ccaatccact	540
aatcagacat gagaacagaa tggtttttagc cagcactaca gctaaggcta tggagcaa	600
ggctggatcg agtgagcaag cagcagaggc catggagggt gctagtccag ctagacaa	660
ggtgcaagcg atgagaacca ttgggactca tcctagctcc agtgctggtc tgaaaaatga	720
tcttcttgaa aatttgcagg cctatcagaa acgaatgggg gtgcagatgc aacggttcaa	780
gtgatcctct cactattgcc gcaaatatca ttgggatctt gcacttgaca ttgtggattc	840
ttgatcgtct ttttttcaaa tgcatttacc gtcgctttaa atacggactg aaaggagggc	900
cttctacgga aggagtgcc aagtctatga ggaagaata tcgaaaggaa cagcagagt	960
ctgtggatgc tgacgatggt cattttgtca gcatagagct ggagtaaaaa actaccttgt	1020
ttctact	1027

<210> SEQ ID NO 6
 <211> LENGTH: 890
 <212> TYPE: DNA
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 6

agcaaaagca gggtgacaaa aacataatgg atccaaacac tgtgtcaagc tttcaggtag	60
attgctttct ttggcatgtc cgcaaacgag ttgcagacca agaactaggc gatgccccat	120
tccttgatcg gcttcgccga gatcagaaat ccctaagagg aaggggcagt actctcggtc	180
tggacatcaa gacagccaca cgtgctggaa agcagatagt ggagcggatt ctgaaagaag	240
aatccgatga ggcacttaaa atgaccatgg cctctgtacc tgcgtcgcgt tacctaactg	300
acatgactct tgaggaaaat tcaagggact ggtccatgct catacccaag cagaaagtgg	360
caggccctct ttgtatcaga atggaccagg cgatcatgga taagaacatc atactgaaag	420
cgaacttcag tgtgatTTTT gaccggctgg agactctaatt attgctaagg gctttcaccg	480
aagagggagc aattgttggc gaaatttcac cattgccttc tcttcaggga catactgctg	540
aggatgtcaa aaatgcagtt ggagtctca tcggaggact tgaatggaat gataacacag	600
ttcagagtctc tgaaactcta cagagattcg cttggagaag cagtaatgag aatgggagac	660
ctccactcac tccaaaacag aaacgagaaa tggcgggaac aattaggta gaagtgtgaa	720
gaaataagat ggttgattga agaagtgaga cacaaactga agataacaga gaatagtttt	780
gagcaataaa catttatgca agccttacat ctattgcttg aagtggagca agagataaga	840
actttctcgt ttcagcttat ttagtactaa aaaacaccct tgtttctact	890

<210> SEQ ID NO 7
 <211> LENGTH: 1775
 <212> TYPE: DNA
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 7

agcaaaagca ggggaaaata aaaacaacca aaatgaaggc aaacctactg gtctgttat	60
gtgcacttgc agctgcagat gcagacacaa tatgtatagg ctaccatgcg aacaattcaa	120

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ccgacactgt tgacacagta ctcgagaaga atgtgacagt gacacactct gttaacctgc	180
tcgaagacag ccacaacgga aaactatgta gattaaaagg aatagcccca ctacaattgg	240
ggaaatgtaa catcgccgga tggctcttgg gaaaccaga atgcgaccca ctgcttccag	300
tgagatcatg gtcctacatt gtagaaacac caaactctga gaatggaata tgttatccag	360
gagatttcat cgactatgag gagctgaggg agcaattgag ctgagtgtca tcattcgaaa	420
gattcgaaat atttcccaa gaaagctcat ggcccaacca caacacaaac ggagtaacgg	480
cagcatgtct ccatgagggg aaaagcagtt ttacagaaa tttgctatgg ctgacggaga	540
aggagggctc atacccaaag ctgaaaaatt cttatgtgaa caaaaaagg aaagaagtcc	600
ttgtactgtg gggatttcat caccgccta acagtaagga acaacagaat ctctatcaga	660
atgaaaatgc ttatgtctct gtagtgactt caaattataa caggagattt accccgaaa	720
tagcagaaag acccaaagta agagatcaag ctgggaggat gaactattac tggaccttgc	780
taaaaccgg agacacaata atatttgagg caaatggaaa tctaatagca ccaatgtatg	840
cttcgcact gagtagaggc tttgggtccg gcatcatcac ctcaaacgca tcaatgcatg	900
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atatacacc agtcacaata ggagagtgcc caaaatacgt caggagtgcc aaattgagga	1020
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ccggttttat tgaaggggga tggactggaa tgatagatgg atggtatggt tatcatcatc	1140
agaatgaaca gggatcaggc tatgcagcgg atcaaaaaag cacacaaat gccattaacg	1200
ggattacaaa caaggtgaac actgttatcg agaaaatgaa cattcaattc acagctgtgg	1260
gtaaagaatt caacaaatta gaaaaagga tggaaaattt aaataaaaaa gttgatgatg	1320
gatttctgga catttggaca tataatgcag aattgttagt tctactggaa aatgaaagga	1380
ctctggattt ccatgactca aatgtgaaga atctgtatga gaaagtaaaa agccaattaa	1440
agaataatgc caaagaaatc ggaatggat gttttgagtt ctaccacaag tgtgacaatg	1500
aatgcatgga aagtgtgaaga aatgggactt atgattatcc caaatattca gaagagtcaa	1560
agttgaacag ggaaggtga gatggagtga aattggaatc aatggggatc tatcagattc	1620
tggcgatcta ctcaactgtc gccagttcac tggtgctttt ggtctccctg ggggcaatca	1680
gtttctggat gtgttctaata ggatctttgc agtgcagaat atgcatctga gattagaatt	1740
tcagagatat gaggaaaaac acccttgttt ctact	1775

<210> SEQ ID NO 8

<211> LENGTH: 1413

<212> TYPE: DNA

<213> ORGANISM: Influenza virus

<400> SEQUENCE: 8

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gtctggtagt cggactaatt agcctaatat tgcaaatagg gaatataatc tcaatatgga	120
ttagccattc aattcaaaact ggaagtcaaa accatactgg aatatgcaac caaaacatca	180
ttacctataa aaatagcacc tgggtaaagg acacaacttc agtgatatta accggcaatt	240
catctctttg tcccatccgt ggggtgggcta tatacagcaa agacaatagc ataagaattg	300
gttccaaagg agacgttttt gtcataagag agccctttat ttcattgtct cacttggaa	360
gcaggacctt ttttctgacc caaggtgcct tactgaatga caagcattca agtgggactg	420

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ttaaggacag aagcccttat agggccttaa tgagctgccc tgctcggtgaa gctccgctccc	480
cgtacaattc aagatttgaa tcggttgctt ggtcagcaag tgcattgcat gatggcatgg	540
gctggctaac aatcggaatt tcaggtccag ataatggagc agtggctgta ttaaaataca	600
acggcataat aactgaaacc ataaaaagt ggaggaagaa aatattgagg acacaagagt	660
ctgaatgtgc ctgtgtaaat ggttcattgt ttactataat gactgatggc ccgagtgatg	720
ggctggcctc gtacaaaatt ttcaagatcg aaaaggggaa gggtactaaa tcaatagagt	780
tgaatgcacc taattctcac tatgaggaat gttcctgtta ccctgatacc ggcaaagtga	840
tgtgtgtgtg cagagacaat tggcatgggt cgaaccggcc atgggtgtct ttcgatcaaa	900
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aagatggaac aggcagctgt ggtccagtgt atgttgatgg agcaaacgga gtaaagggat	1020
tttcatatag gtatggtaat ggtgtttgga taggaaggac caaaagtcac agttccagac	1080
atgggtttga gatgatgttg gatcctaatt gatggacaga gactgatagt aagttctctg	1140
tgaggcaaga tggtgtggca atgactgatt ggtcagggtg tagcggaggt ttcgttcaac	1200
atcctgagct gacagggcta gactgtatga ggccgtgctt ctgggttgaa ttaatcagg	1260
gacgacctaa agaaaaaaca atctggacta gtgcgagcag catttctttt tgtggcgtga	1320
atagtatac ttagatttgg tcttggccag acggtgctga gttgccattc agcattgaca	1380
agtagtctgt tcaaaaaact cctgtttct act	1413

<210> SEQ ID NO 9

<400> SEQUENCE: 9

000

<210> SEQ ID NO 10

<211> LENGTH: 2341

<212> TYPE: DNA

<213> ORGANISM: Influenza virus

<400> SEQUENCE: 10

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ccagcacaaa atgctataag cacaactttc ccttataccg gagaccctcc ttacagccat	120
gggacaggaa caggatacac catggatact gtcaacagga cacatcagta ctcagaaaag	180
ggaagatgga caacaacac cgaaactgga gcaccgcaac tcaaccgat tgatgggcca	240
ctgccagaag acaatgaacc aagtgggtat gcccacacag attgtgtatt ggaagcaatg	300
gctttccttg aggaatccca tcttgggtatt ttgaaaaact cgtgtattga aacgatggag	360
gttgttcagc aaacacgagt agacaagctg acacaaggcc gacagacctg tgactggact	420
ttaaatagaa accagcctgc tgcaacagca ttggccaaca caatagaagt gttcagatca	480
aatggcctca cggccaatga gtcaggaagg ctcatagact tccttaagga tgtaattggag	540
tcaatgaaaa aagaagaaat ggggatcaca actcattttc agagaaagag acgggtgaga	600
gacaatatga ctaagaaaat gataacacag agaacaatag gtaaaaggaa acagagattg	660
aacaaaaggg gttatctaata tagagcattg accctgaaca caatgaccaa agatgctgag	720
agaggggaagc taaaacggag agcaattgca accccaggga tgcaaataag ggggtttgta	780
tactttgttg agacactggc aaggagtata tgtgagaaac ttgaacaatc aggggttgcca	840
gttgagggca atgagaagaa agcaaagttg gcaaatgttg taaggaagat gatgaccaat	900

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tctcaggaca	ccgaactttc	tttcaccatc	actggagata	acaccaaag	gaacgaaaat	960
cagaatccto	ggatgttttt	ggccatgatc	acatatatga	ccagaaatca	gcccgaatgg	1020
ttcagaaatg	ttctaagtat	tgctccaata	atgtttctca	acaaaatggc	gagactggga	1080
aaagggatata	tgtttgagag	caagagtatg	aaacttagaa	ctcaaatacc	tgacgaaatg	1140
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tgcaatttat	ttgaaaaatt	cttccccagc	agttcataca	gaagaccagt	cgggatatcc	2160
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ggaaggataa	agaaagaaga	gttcactgag	atcatgaaga	tctgttccac	cattgaagag	2280
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t						2341

<210> SEQ ID NO 11

<211> LENGTH: 2341

<212> TYPE: DNA

<213> ORGANISM: Influenza virus

<400> SEQUENCE: 11

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aagaagtaca	catcaggaag	acaggagaag	aaccacgac	ttaggatgaa	atggatgatg	180
gcaatgaaat	atccaattac	agcagacaag	aggataacgg	aatgatctcc	tgagagaaat	240
gagcaaggac	aaactttatg	gagtaaaatg	aatgatgccg	gatcagaccg	agtgatggta	300
tcacctctgg	ctgtgacatg	gtggaatagg	aatggaccaa	tgacaaatc	agttcattat	360
ccaaaaatct	acaaaactta	ttttgaaaga	gtcgaaaggc	taaagcatgg	aacctttggc	420
cctgtccatt	ttagaaacca	agtcaaaata	cgtcggagag	ttgacataaa	tcctgggtcat	480
gcagatctca	gtgccaaagg	ggcacaggat	gtaatcatgg	aagttgtttt	ccctaacgaa	540
gtgggagcca	ggatactaac	atcggaatcg	caactaacga	taaccaaaga	gaagaaagaa	600
gaactccagg	attgcaaaat	ttctcctttg	atggttgcat	acatgttgga	gagagaactg	660

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gtccgcaaaa cgagattcct ccagtggt ggtggaacaa gcagtgtgta cattgaagtg 720
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aatgatgatg ttgatcaaa cttgattatt gctgctagga acatagttag aagagctgca 840
gtatcagcag acccactagc atctttattg gagatgtgcc acagcacaca gattggtgga 900
attaggatgg tagacatcct taagcagaac ccaacagaag agcaagccgt ggatatatgc 960
aaggctgcaa tgggactgag aattagctca tcttcagtt ttggtggatt cacatttaag 1020
agaacaagcg gatcatcagt caagagagag gaagaggtgc ttacgggcaa tcttcaaca 1080
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cagtcgattg ccgaagcaat aattgtggcc atggtatctt cacaagagga ttgtatgata 1260
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ctactgtctc ccgaggaggt cagtgaacaa cagggaacag agaaactgac aataacttac 1620
tcactgtcaa tgatgtggga gattaatggt cctgaatcag tgttggtaa tacctatcaa 1680
tggatcatca gaaactggga aactgttaaa attcagtggg ccagaaccc tacaatgcta 1740
tacaataaaa tggaaattga accatttcag tcttttagtac ctaaggccat tagaggccaa 1800
tacagtgggt ttgtaagaac tctgttccaa caaatgaggg atgtgcttgg gacatttgat 1860
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cagttctcct catttactgt gaatgtgagg ggatcaggaa tgagaatact tgtaaggggc 1980
aattctcctg tattcaacta caacaaggcc acgaagagac tcacagttct cggaaaggat 2040
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gtgttggtaa tgaacgaaa acgggactct agcatactta ctgacagcca gacagcgacc 2280
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t 2341

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<210> SEQ ID NO 12
<211> LENGTH: 2233
<212> TYPE: DNA
<213> ORGANISM: Influenza virus

<400> SEQUENCE: 12

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aacaatttg cagcaatatg cactcacttg gaagtatgct tcatgtattc agatttccac 180
ttcatcaatg agcaaggcga gtcaataatc gtagaacttg gtgatcctaa tgcacttttg 240
aagcacagat ttgaataaat cgagggaaga gatcgacaaa tggcctggac agtagtaaac 300
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aagggaaaata gattcatcga aattggagta acaaggagag aagttcacat atactatctg	420
gaaaaggcca ataaaattaa atctgagaaa acacacatcc acattttctc gttcactggg	480
gaagaaatgg ccacaagggc cgactacact ctcgatgaag aaagcagggc taggatcaaa	540
accaggctat tcaccataag acaagaaatg gccagcagag gcctctggga ttctttctgt	600
cagtccgaga gaggagaaga gacaattgaa gaaaggtttg aaatcacagg aacaatgcgc	660
aagcttgccg accaaagtct cccgccgaac ttctccagcc ttgaaaattt tagagcctat	720
gtggatggat tcgaaccgaa cggctacatt gagggcaagc tgtctcaaat gtccaaagaa	780
gtaaatgcta gaattgaacc ttttttgaaa acaacaccac gaccacttag acttccgaat	840
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gaggacccaa gtcataagg agagggata ccgctatatg atgcaatcaa atgcatgaga	960
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aaaattccaa agactaaaaa tatgaaaaa acaagtcagc taaagtgggc acttggtgag	1140
aacatggcac cagaaaagg agactttgac gactgtaaag atgtaggaga tttgaagcaa	1200
tatgatagtg atgaaccaga attgaggtcg cttgcaagtt ggattcagaa tgagttcaac	1260
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gtccaattg aacacattgc aagcatgaga aggaattatt tcacatcaga ggtgtctcac	1380
tcgagagcca cagaatacat aatgaagggg gtgtacatca atactgcctt acttaatgca	1440
tcttgtgcag caatggatga tttccaatta attccaatga taagcaagtg tagaactaag	1500
gaggggaagg gaaagaccaa cttgtatggt ttcatacataa aaggaagatc ccacttaagg	1560
aatgacacgg acgtggtaaa ctttgtgagc atggagtttt ctctcactga cccaagactt	1620
gaaccacaca aatgggagaa gtactgtggt cttgagatag gagatatgct tctaagaagt	1680
gccataggcc aggtttcaag gccatgttc ttgtatgtga ggacaaatgg aacctcaaaa	1740
attaaaaatga aatggggaat ggagatgagg cgttgtctcc tcagtcact tcaacaaatt	1800
gagagtatga ttgaagctga gtcctctgtc aaagagaaag acatgaccaa agagttcttt	1860
gagaacaaat cagaacatg gccattgga gagtctccca aaggagtgga ggaaagtctc	1920
attgggaagg tctgcaggac tttattagca aagtcggtat ttaacagctt gtatgcatct	1980
ccacaactag aaggattttc agctgaatca agaaaactgc ttcttatcgt tcaggctctt	2040
agggacaatc tggaacctgg gacctttgat cttggggggc tatatgaagc aattgaggag	2100
tgctaatta atgatccctg ggttttgctt aatgcttctt ggttcaactc cttccttaca	2160
catgcattga gttagtgtg gcagtgtac tatttgctat ccatactgtc caaaaaagta	2220
ccttgtttct act	2233

<210> SEQ ID NO 13

<211> LENGTH: 1565

<212> TYPE: DNA

<213> ORGANISM: Influenza virus

<400> SEQUENCE: 13

agcaaaagca gggtagataa tcactcactg agtgacatca aaatcatggc gtcccaaggc	60
accaaacggt cttacgaaca gatggagact gatggagaac gccagaatgc cactgaaatc	120
agagcatccg tcggaaaaat gattggtgga attggacgat tctacatcca aatgtgcaca	180
gaacttaaac tcagtgatta tgagggacgg ttgatccaaa acagcttaac aatagagaga	240

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atggtgctct ctgcttttga cgaaaggaga aataaatacc tggaagaaca tcccagtgcg	300
gggaaagatc ctaagaaaac tggaggacct atatacagaa gagtaaacgg aaagtggatg	360
agagaactca tcctttatga caaagaagaa ataaggcgaa tctggcgcca agctaataat	420
ggtgacgatg caacggctgg tctgactcac atgatgatct ggcattecaa tttgaatgat	480
gcaacttata agaggacaag ggctcttgtt cgcaccggaa tggatcccag gatgtgctct	540
ctgatgcaag gttcaactct ccttaggagg tctggagccg caggtgctgc agtcaaagga	600
gttggaacaa tggatgatga attggtcagg atgatcaaac gtgggatcaa tgatcggaac	660
ttctggagggt gtgagaatgg acgaaaaaca agaattgctt atgaaagaat gtgcaacatt	720
ctcaaaggga aatttcaaac tgtgacacaa aaagcaatga tggatcaagt gagagagagc	780
cggaaaccag ggaatgctga gttcgaagat ctcaactttc tagcacggtc tgcactcata	840
ttgagagggt cggttgctca caagtctgc ctgcctgcct gtgtgtatgg acctgccgta	900
gccagtgggt acgactttga aagagaggga tactctctag tcggaataga ccctttcaga	960
ctgcttcaaa acagccaagt gtacagccta atcagaccaa atgagaatcc agcacacaag	1020
agtcaactgg tgtggatggc atgccattct gccgcatttg aagatctaag agtattgagc	1080
ttcatcaaa ggacgaaggt ggtcccaaga ggaagcctt ccactagagg agttcaaatt	1140
gcttccaatg aaaatatgga gactatggaa tcaagtacac ttgaactgag aagcaggtac	1200
tgggccataa ggaccagaag tggaggaaac accaatcaac agagggcatc tgcgggcca	1260
atcagcatac aacctacgtt ctcagtacag agaaatctcc cttttgacag aacaaccgtt	1320
atggcagcat tcaactggaa tacagagggg agaacatctg acatgaggac cgaaatcata	1380
aggatgatgg aaagtgaag accagaagat gtgtctttcc aggggcgggg agtcttcgag	1440
ctctcggacg aaaaggcagc gagcccgatc gtgccttctt ttgacatgag taatgaagga	1500
tcttatctct tcggagacaa tgcagaggag tacgacaatt aaagaaaaat acccttgttt	1560
ctact	1565

<210> SEQ ID NO 14

<211> LENGTH: 1027

<212> TYPE: DNA

<213> ORGANISM: Influenza virus

<400> SEQUENCE: 14

agcaaaagca ggtagatatt gaaagatgag tcttctaacc gaggtcgaaa cgtacgttct	60
ctctatcatc ccgtcaggcc ccctcaaagc cgagatcgca cagagacttg aagatgtctt	120
tgcagggaag aacaccgatc ttgaggttct catggaatgg ctaaagacaa gaccaatcct	180
gtcacctctg actaagggga ttttaggatt tgtgttcacg ctcaaccgtc ccagtgagcg	240
aggactgcag cgtagacgct ttgtccaaaa tgcccttaat gggaacgggg atccaaataa	300
catggacaaa gcagttaaac tgtataggaa gctcaagagg gagataacat tccatggggc	360
caaagaaatc tcaactcagtt attctgctgg tgcacttgc agttgtatgg gcctcatata	420
caacaggatg ggggctgtga ccactgaagt ggcatttggc ctggtatgtg caacctgtga	480
acagattgct gactcccagc atcggctctca taggcaaatg gtgacaacaa ccaaccact	540
aatcagacat gagaacagaa tggtttttagc cagcactaca gctaaggcta tggagcaaat	600
ggctggatcg agtgagcaag cagcagaggc catggagggt gctagttagg ctaggcaaat	660
ggtgcaagcg atgagaacca ttgggactca tctagctcc agtgctggtc tgaaaaatga	720

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tcttcttgaa aatttgcagg cctatcagaa acgaatgggg gtgcagatgc aacgggttcaa	780
gtgatcctct cgctattgcc gcaaatatca ttgggatctt gcaacttgata ttgtggattc	840
ttgatcgtct ttttttcaaa tgcatttacc gtgcgtttta atacggactg aaaggagggc	900
cttctacgga aggagtgcc aagtctatga gggaagaata tcgaaaggaa cagcagagtg	960
ctgtggatgc tgacgatggt cattttgtca gcatagagct ggagtaaaaa actaccttgt	1020
ttctact	1027

<210> SEQ ID NO 15
 <211> LENGTH: 890
 <212> TYPE: DNA
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 15

agcaaaagca gggtgacaaa gacataatgg atccaaacac tgtgtcaagc tttcaggtag	60
attgctttct ttggcatgac cgcaaacgag ttgcagacca agaactaggt gatgccccat	120
tccttgatcg gcttcgccga gatcagaaat ccctaagagg aaggggcagc actcttggtc	180
tggacatcga gacagccaca cgtgctggaa agcagatagt ggagcggatt ctgaaagaag	240
aatccgatga ggcacttaaa atgaccatgg cctctgtacc tgcgtcgcgt tacctaaccg	300
acatgactct tgaggaaatg tcaagggaat ggtccatgct catacccaag cagaaagtgg	360
caggccctct ttgtatcaga atggaccagg cgatcatgga taaaaacatc atactgaaag	420
cgaacttcag tgtgattttt gaccggctgg agactctaatt attgctaagg gctttcaccg	480
aagagggagc aattgttggc gaaatttcac cattgccttc tcttcagga catactgtg	540
aggatgtcaa aaatgcagtt ggagtctca tcggaggact tgaatggaat gataacacag	600
ttcgagtctc tgaacttcta cagagattcg cttggagaag cagtaatgag aatgggagac	660
ctccactcac tccaaaacag aaacgagaaa tggcgggaac aattaggtca gaagtttgaa	720
gaaataagat ggttgattga agaagtgaga cacaaactga aggtaacaga gaatagtttt	780
gagcaataaa catttatgca agccttacat ctattgcttg aagtggagca agagataaga	840
actttctcat ttcagcttat ttaataataa aaaacaccct tgtttctact	890

<210> SEQ ID NO 16
 <211> LENGTH: 319
 <212> TYPE: PRT
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 16

Ala Thr Leu Cys Leu Gly His His Ala Val Pro Asn Gly Thr Leu Val	
1 5 10 15	
Lys Thr Ile Thr Asp Asp Gln Ile Glu Val Thr Asn Ala Thr Glu Leu	
20 25 30	
Val Gln Ser Ser Ser Thr Gly Lys Ile Cys Asn Asn Pro His Arg Ile	
35 40 45	
Leu Asp Gly Ile Asp Cys Thr Leu Ile Asp Ala Leu Leu Gly Asp Pro	
50 55 60	
His Cys Asp Val Phe Gln Asn Glu Thr Trp Asp Leu Phe Val Glu Arg	
65 70 75 80	
Ser Lys Ala Phe Ser Asn Cys Tyr Pro Tyr Asp Val Pro Asp Tyr Ala	
85 90 95	
Ser Leu Arg Ser Leu Val Ala Ser Ser Gly Thr Leu Glu Phe Ile Thr	
100 105 110	

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Glu Gly Phe Thr Trp Thr Gly Val Thr Gln Asn Gly Gly Ser Asn Ala
 115 120 125
 Cys Lys Arg Gly Pro Gly Ser Gly Phe Phe Ser Arg Leu Asn Trp Leu
 130 135 140
 Thr Lys Ser Gly Ser Thr Tyr Pro Val Leu Asn Val Thr Met Pro Asn
 145 150 155 160
 Asn Asp Asn Phe Asp Lys Leu Tyr Ile Trp Gly Ile His His Pro Ser
 165 170 175
 Thr Asn Gln Glu Gln Thr Ser Leu Tyr Val Gln Ala Ser Gly Arg Val
 180 185 190
 Thr Val Ser Thr Arg Arg Ser Gln Gln Thr Ile Ile Pro Asn Ile Gly
 195 200 205
 Ser Arg Pro Trp Val Arg Gly Leu Ser Ser Arg Ile Ser Ile Tyr Trp
 210 215 220
 Thr Ile Val Lys Pro Gly Asp Val Leu Val Ile Asn Ser Asn Gly Asn
 225 230 235 240
 Leu Ile Ala Pro Arg Gly Tyr Phe Lys Met Arg Thr Gly Lys Ser Ser
 245 250 255
 Ile Met Arg Ser Asp Ala Pro Ile Asp Thr Cys Ile Ser Glu Cys Ile
 260 265 270
 Thr Pro Asn Gly Ser Ile Pro Asn Asp Lys Pro Phe Gln Asn Val Asn
 275 280 285
 Lys Ile Thr Tyr Gly Ala Cys Pro Lys Tyr Val Lys Gln Asn Thr Leu
 290 295 300
 Lys Leu Ala Thr Gly Met Arg Asn Val Pro Glu Lys Gln Thr Arg
 305 310 315

<210> SEQ ID NO 17
 <211> LENGTH: 326
 <212> TYPE: PRT
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 17

Asp Gln Ile Cys Ile Gly Tyr His Ala Asn Asn Ser Thr Glu Gln Val
 1 5 10 15
 Asp Thr Ile Met Glu Lys Asn Ile Thr Val Thr His Ala Gln Asp Ile
 20 25 30
 Leu Glu Lys Thr His Asn Gly Lys Leu Cys Asp Leu Asn Gly Val Lys
 35 40 45
 Pro Leu Ile Leu Arg Asp Cys Ser Val Ala Gly Trp Leu Leu Gly Asn
 50 55 60
 Pro Met Cys Asp Glu Phe Leu Asn Val Pro Glu Trp Ser Tyr Ile Val
 65 70 75 80
 Glu Lys Asp Asn Pro Val Asn Gly Leu Cys Tyr Pro Glu Asn Phe Asn
 85 90 95
 Asp Tyr Glu Glu Leu Lys His Leu Leu Ser Ser Thr Asn His Phe Glu
 100 105 110
 Lys Ile Arg Ile Ile Pro Arg Ser Ser Trp Ser Asn His Asp Ala Ser
 115 120 125
 Ser Gly Val Ser Ser Ala Cys Pro Tyr Asn Gly Arg Ser Ser Phe Phe
 130 135 140
 Arg Asn Val Val Trp Leu Ile Lys Lys Asn Asn Ala Tyr Pro Thr Ile
 145 150 155 160
 Lys Arg Ser Tyr Asn Asn Thr Asn Gln Glu Asp Leu Leu Ile Leu Trp
 165 170 175

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Gly Ile His His Pro Asn Asp Ala Ala Glu Gln Thr Lys Leu Tyr Gln
 180 185 190
 Asn Pro Thr Thr Tyr Val Ser Val Gly Thr Ser Thr Leu Asn Gln Arg
 195 200 205
 Ser Ile Pro Glu Ile Ala Thr Arg Pro Lys Val Asn Gly Gln Ser Gly
 210 215 220
 Arg Met Glu Phe Tyr Trp Thr Ile Leu Lys Pro Asn Asp Ala Ile Asn
 225 230 235 240
 Phe Glu Ser Asn Gly Asn Phe Ile Ala Pro Glu Tyr Ala Tyr Lys Ile
 245 250 255
 Val Lys Lys Gly Gly Ser Ala Ile Met Lys Ser Gly Leu Glu Tyr Gly
 260 265 270
 Asn Cys Asn Thr Lys Cys Gln Thr Pro Met Gly Ala Ile Asn Ser Ser
 275 280 285
 Met Pro Phe His Asn Val His Pro Leu Thr Ile Gly Glu Cys Pro Lys
 290 295 300
 Tyr Val Lys Ser Gly Arg Leu Val Leu Ala Thr Gly Leu Arg Asn Val
 305 310 315 320
 Pro Gln Arg Glu Thr Arg
 325

<210> SEQ ID NO 18
 <211> LENGTH: 330
 <212> TYPE: PRT
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 18

Asp Gln Ile Cys Ile Gly Tyr His Ala Asn Asn Ser Thr Glu Gln Val
 1 5 10 15
 Asp Thr Ile Met Glu Lys Asn Ile Thr Val Thr His Ala Gln Asp Ile
 20 25 30
 Leu Glu Arg Thr His Asn Gly Lys Leu Cys Asp Leu Asn Gly Val Lys
 35 40 45
 Pro Leu Ile Leu Arg Asp Cys Ser Val Ala Gly Trp Leu Leu Gly Asn
 50 55 60
 Pro Met Cys Asp Glu Phe Ile Asn Val Pro Glu Trp Ser Tyr Ile Val
 65 70 75 80
 Glu Lys Ala Ser Pro Ala Asn Asp Leu Cys Tyr Pro Gly Asn Phe Asn
 85 90 95
 Asp Tyr Glu Glu Leu Lys His Leu Leu Ser Arg Ile Asn His Phe Glu
 100 105 110
 Lys Ile Gln Ile Ile Pro Lys Ser Ser Trp Ser Asn His Asp Ala Ser
 115 120 125
 Ser Gly Val Ser Ser Ala Cys Pro Tyr Leu Gly Arg Ser Ser Phe Phe
 130 135 140
 Arg Asn Val Val Trp Leu Ile Lys Lys Asn Ser Ala Tyr Pro Thr Ile
 145 150 155 160
 Lys Arg Ser Tyr Asn Asn Thr Asn Gln Glu Asp Leu Leu Val Leu Trp
 165 170 175
 Gly Ile His His Pro Asn Asp Ala Ala Glu Gln Thr Lys Leu Tyr Gln
 180 185 190
 Asn Pro Thr Thr Tyr Val Ser Val Gly Thr Ser Thr Leu Asn Gln Arg
 195 200 205
 Leu Ile Pro Glu Ile Ala Thr Arg Pro Lys Val Asn Gly Gln Ser Gly

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210	215	220
Arg Met Glu Phe Tyr Trp Thr Ile Leu Lys Pro Asn Asp Ala Ile Asn		
225	230	235 240
Phe Glu Ser Asn Gly Asn Phe Ile Ala Pro Glu Tyr Ala Tyr Lys Ile		
	245	250 255
Val Lys Lys Gly Asp Ser Thr Ile Met Lys Ser Glu Leu Glu Tyr Gly		
	260	265 270
Asn Cys Asn Thr Lys Cys Gln Thr Pro Met Gly Ala Ile Asn Ser Ser		
	275	280 285
Met Pro Phe His Asn Val His Pro Leu Thr Ile Gly Glu Cys Pro Lys		
	290	295 300
Tyr Val Lys Ser Asn Arg Leu Val Leu Ala Thr Gly Leu Arg Asn Thr		
305	310	315 320
Pro Gln Arg Glu Arg Arg Arg Lys Lys Arg		
	325	330

<210> SEQ ID NO 19

<211> LENGTH: 325

<212> TYPE: PRT

<213> ORGANISM: Influenza virus

<400> SEQUENCE: 19

Asp Lys Ile Cys Ile Gly Tyr Gln Ser Thr Asn Ser Thr Glu Thr Val		
1	5	10 15
Asp Thr Leu Met Glu Thr Asn Ile Pro Val Thr His Ala Lys Asp Ile		
	20	25 30
Leu His Thr Glu His Asn Gly Met Leu Cys Ala Thr Asn Leu Gly His		
	35	40 45
Pro Leu Ile Leu Asp Thr Cys Ser Ile Glu Gly Leu Ile Tyr Gly Asn		
	50	55 60
Pro Ser Cys Asp Leu Leu Leu Gly Gly Arg Glu Trp Ser Tyr Ile Val		
65	70	75 80
Glu Lys Pro Ser Pro Val Asn Gly Met Cys Tyr Pro Gly Asn Phe Glu		
	85	90 95
Asn Leu Glu Glu Leu Lys His Leu Phe Ser Arg Ala Ser Ser Tyr Gln		
	100	105 110
Arg Ile Gln Ile Ile Pro Asp Thr Ile Trp Asn His Ser Tyr Ser Ser		
	115	120 125
Gly Thr Ser Arg Ala Cys Ser Asp Ser Phe Phe Arg Ser Met Arg Trp		
	130	135 140
Leu Ile Gln Lys Asn Asn Ala Tyr Pro Thr Gln Asp Ala Gln Tyr Thr		
145	150	155 160
Asn Thr Arg Gly Lys Ser Ile Leu Val Met Trp Gly Ile Asn His Pro		
	165	170 175
Pro Asp Asp Thr Val Gln Thr Asn Leu Tyr Thr Arg Thr Asp Thr Thr		
	180	185 190
Thr Ser Val Thr Thr Glu Asp Ile Asn Arg Arg Phe Lys Pro Val Ile		
	195	200 205
Ala Pro Arg Pro Leu Val Asn Gly Gln His Gly Arg Met Asp Tyr Tyr		
	210	215 220
Trp Ser Ile Leu Lys Pro Asn Gln Thr Ile Arg Phe Arg Ser Asn Gly		
225	230	235 240
Asn Phe Ile Ala Pro Trp Tyr Ala His Ile Leu Ser Gly Glu Ser His		
	245	250 255

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Gly Arg Ile Leu Lys Thr Glu Leu Asn Ser Gly Asn Cys Asn Val Gln
 260 265 270

Cys Gln Thr Glu Arg Gly Gly Leu Asn Thr Thr Leu Pro Phe His Asn
 275 280 285

Val Ser Pro Tyr Ala Ile Gly Asn Cys Pro Lys Tyr Val Gly Val Lys
 290 295 300

Ser Leu Val Leu Ala Val Gly Leu Arg Asn Thr Pro Ala Arg Ser Ser
 305 310 315 320

Arg Arg Lys Lys Arg
 325

<210> SEQ ID NO 20
 <211> LENGTH: 325
 <212> TYPE: PRT
 <213> ORGANISM: Influenza virus
 <400> SEQUENCE: 20

Asp Lys Ile Cys Ile Gly Tyr Gln Ser Thr Asn Ser Thr Glu Thr Val
 1 5 10 15

Asp Thr Leu Met Glu Thr Asn Ile Pro Val Thr His Ala Lys Asp Ile
 20 25 30

Leu His Thr Glu His Asn Gly Met Leu Cys Ala Thr Ser Leu Gly His
 35 40 45

Pro Leu Ile Leu Asp Thr Cys Ser Ile Glu Gly Leu Val Tyr Gly Asn
 50 55 60

Pro Ser Cys Asp Leu Leu Leu Gly Gly Arg Glu Trp Ser Tyr Ile Val
 65 70 75 80

Glu Lys Pro Ser Pro Val Asn Gly Thr Cys Tyr Pro Gly Asn Phe Glu
 85 90 95

Asn Leu Glu Glu Leu Lys Thr Leu Phe Ser Arg Ala Ser Ser Tyr Gln
 100 105 110

Arg Ile Gln Ile Ile Pro Asp Thr Ile Trp Asn His Ser Tyr Thr Ser
 115 120 125

Gly Thr Ser Arg Ala Cys Ser Gly Ser Phe Phe Arg Ser Met Arg Trp
 130 135 140

Leu Ile Gln Lys Ser Gly Phe Tyr Pro Thr Gln Asp Ala Gln Tyr Thr
 145 150 155 160

Asn Thr Arg Gly Lys Ser Ile Leu Val Met Trp Gly Ile Asn His Pro
 165 170 175

Pro Asp Tyr Thr Val Gln Thr Asn Leu Tyr Thr Arg Asn Asp Thr Thr
 180 185 190

Thr Ser Val Thr Thr Glu Asp Leu Asn Arg Arg Phe Lys Pro Val Ile
 195 200 205

Ala Pro Arg Pro Leu Val Asn Gly Gln Gln Gly Arg Met Asp Tyr Tyr
 210 215 220

Trp Ser Ile Leu Lys Pro Asn Gln Thr Ile Arg Phe Arg Ser Asn Gly
 225 230 235 240

Asn Phe Ile Ala Pro Trp Tyr Ala His Val Leu Ser Gly Gly Ser His
 245 250 255

Gly Arg Ile Leu Lys Thr Glu Leu Lys Gly Gly Asn Cys Asn Val Gln
 260 265 270

Cys Gln Thr Glu Lys Gly Gly Leu Asn Ser Thr Leu Pro Phe His Asn
 275 280 285

Val Ser Pro Tyr Ala Ile Gly Thr Cys Pro Lys Tyr Val Arg Val Lys
 290 295 300

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Ser Leu Val Leu Ala Val Gly Leu Arg Asn Thr Pro Ala Arg Ser Ser
305 310 315 320

Arg Arg Lys Lys Arg
325

<210> SEQ ID NO 21
<211> LENGTH: 566
<212> TYPE: PRT
<213> ORGANISM: Influenza virus

<400> SEQUENCE: 21

Met Lys Ala Ile Leu Val Val Leu Leu Tyr Thr Phe Ala Thr Ala Asn
1 5 10 15

Ala Asp Thr Leu Cys Ile Gly Tyr His Ala Asn Asn Ser Thr Asp Thr
20 25 30

Val Asp Thr Val Leu Glu Lys Asn Val Thr Val Thr His Ser Val Asn
35 40 45

Leu Leu Glu Asp Lys His Asn Gly Lys Leu Cys Lys Leu Arg Gly Val
50 55 60

Ala Pro Leu His Leu Gly Lys Cys Asn Ile Ala Gly Trp Ile Leu Gly
65 70 75 80

Asn Pro Glu Cys Glu Ser Leu Ser Thr Ala Ser Ser Trp Ser Tyr Ile
85 90 95

Val Glu Thr Pro Ser Ser Asp Asn Gly Thr Cys Tyr Pro Gly Asp Phe
100 105 110

Ile Asp Tyr Glu Glu Leu Arg Glu Gln Leu Ser Ser Val Ser Ser Phe
115 120 125

Glu Arg Phe Glu Ile Phe Pro Lys Thr Ser Ser Trp Pro Asn His Asp
130 135 140

Ser Asn Lys Gly Val Thr Ala Ala Cys Pro His Ala Gly Ala Lys Ser
145 150 155 160

Phe Tyr Lys Asn Leu Ile Trp Leu Val Lys Lys Gly Asn Ser Tyr Pro
165 170 175

Lys Leu Ser Lys Ser Tyr Ile Asn Asp Lys Gly Lys Glu Val Leu Val
180 185 190

Leu Trp Gly Ile His His Pro Ser Thr Ser Ala Asp Gln Gln Ser Leu
195 200 205

Tyr Gln Asn Ala Asp Ala Tyr Val Phe Val Gly Ser Ser Arg Tyr Ser
210 215 220

Lys Lys Phe Lys Pro Glu Ile Ala Ile Arg Pro Lys Val Arg Asp Gln
225 230 235 240

Glu Gly Arg Met Asn Tyr Tyr Trp Thr Leu Val Glu Pro Gly Asp Lys
245 250 255

Ile Thr Phe Glu Ala Thr Gly Asn Leu Val Val Pro Arg Tyr Ala Phe
260 265 270

Ala Met Glu Arg Asn Ala Gly Ser Gly Ile Ile Ile Ser Asp Thr Pro
275 280 285

Val His Asp Cys Asn Thr Thr Cys Gln Thr Pro Lys Gly Ala Ile Asn
290 295 300

Thr Ser Leu Pro Phe Gln Asn Ile His Pro Ile Thr Ile Gly Lys Cys
305 310 315 320

Pro Lys Tyr Val Lys Ser Thr Lys Leu Arg Leu Ala Thr Gly Leu Arg
325 330 335

Asn Ile Pro Ser Ile Gln Ser Arg Gly Leu Phe Gly Ala Ile Ala Gly

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340					345					350					
Phe	Ile	Glu	Gly	Gly	Trp	Thr	Gly	Met	Val	Asp	Gly	Trp	Tyr	Gly	Tyr
		355					360					365			
His	His	Gln	Asn	Glu	Gln	Gly	Ser	Gly	Tyr	Ala	Ala	Asp	Leu	Lys	Ser
	370					375					380				
Thr	Gln	Asn	Ala	Ile	Asp	Glu	Ile	Thr	Asn	Lys	Val	Asn	Ser	Val	Ile
385					390					395					400
Glu	Lys	Met	Asn	Thr	Gln	Phe	Thr	Ala	Val	Gly	Lys	Glu	Phe	Asn	His
			405						410					415	
Leu	Glu	Lys	Arg	Ile	Glu	Asn	Leu	Asn	Lys	Lys	Val	Asp	Asp	Gly	Phe
			420						425				430		
Leu	Asp	Ile	Trp	Thr	Tyr	Asn	Ala	Glu	Leu	Leu	Val	Leu	Leu	Glu	Asn
	435						440					445			
Glu	Arg	Thr	Leu	Asp	Tyr	His	Asp	Ser	Asn	Val	Lys	Asn	Leu	Tyr	Glu
	450					455					460				
Lys	Val	Arg	Ser	Gln	Leu	Lys	Asn	Asn	Ala	Lys	Glu	Ile	Gly	Asn	Gly
465					470					475					480
Cys	Phe	Glu	Phe	Tyr	His	Lys	Cys	Asp	Asn	Thr	Cys	Met	Glu	Ser	Val
			485					490					495		
Lys	Asn	Gly	Thr	Tyr	Asp	Tyr	Pro	Lys	Tyr	Ser	Glu	Glu	Ala	Lys	Leu
		500						505					510		
Asn	Arg	Glu	Glu	Ile	Asp	Gly	Val	Lys	Leu	Glu	Ser	Thr	Arg	Ile	Tyr
		515					520					525			
Gln	Ile	Leu	Ala	Ile	Tyr	Ser	Thr	Val	Ala	Ser	Ser	Leu	Val	Leu	Val
	530					535					540				
Val	Ser	Leu	Gly	Ala	Ile	Ser	Phe	Trp	Met	Cys	Ser	Asn	Gly	Ser	Leu
545					550					555					560
Gln	Cys	Arg	Ile	Cys	Ile										
				565											

<210> SEQ ID NO 22

<211> LENGTH: 231

<212> TYPE: PRT

<213> ORGANISM: Influenza virus

<400> SEQUENCE: 22

Arg	Asn	Ile	Pro	Ser	Ile	Gln	Ser	Arg	Gly	Leu	Phe	Gly	Ala	Ile	Ala
1				5					10					15	
Gly	Phe	Ile	Glu	Gly	Gly	Trp	Thr	Gly	Met	Val	Asp	Gly	Trp	Tyr	Gly
		20						25					30		
Tyr	His	His	Gln	Asn	Glu	Gln	Gly	Ser	Gly	Tyr	Ala	Ala	Asp	Leu	Lys
	35					40						45			
Ser	Thr	Gln	Asn	Ala	Ile	Asp	Glu	Ile	Thr	Asn	Lys	Val	Asn	Ser	Val
	50					55					60				
Ile	Glu	Lys	Met	Asn	Thr	Gln	Phe	Thr	Ala	Val	Gly	Lys	Glu	Phe	Asn
65					70					75				80	
His	Leu	Glu	Lys	Arg	Ile	Glu	Asn	Leu	Asn	Lys	Lys	Val	Asp	Asp	Gly
			85					90						95	
Phe	Leu	Asp	Ile	Trp	Thr	Tyr	Asn	Ala	Glu	Leu	Leu	Val	Leu	Leu	Glu
		100						105					110		
Asn	Glu	Arg	Thr	Leu	Asp	Tyr	His	Asp	Ser	Asn	Val	Lys	Asn	Leu	Tyr
	115						120					125			
Glu	Lys	Val	Arg	Ser	Gln	Leu	Lys	Asn	Asn	Ala	Lys	Glu	Ile	Gly	Asn
	130					135						140			

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Gly Cys Phe Glu Phe Tyr His Lys Cys Asp Asn Thr Cys Met Glu Ser
145 150 155 160

Val Lys Asn Gly Thr Tyr Asp Tyr Pro Lys Tyr Ser Glu Glu Ala Lys
165 170 175

Leu Asn Arg Glu Glu Ile Asp Gly Val Lys Leu Glu Ser Thr Arg Ile
180 185 190

Tyr Gln Ile Leu Ala Ile Tyr Ser Thr Val Ala Ser Ser Leu Val Leu
195 200 205

Val Val Ser Leu Gly Ala Ile Ser Phe Trp Met Cys Ser Asn Gly Ser
210 215 220

Leu Gln Cys Arg Ile Cys Ile
225 230

<210> SEQ ID NO 23
 <211> LENGTH: 221
 <212> TYPE: PRT
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 23

Gly Leu Phe Gly Ala Ile Ala Gly Phe Ile Glu Asn Gly Trp Glu Gly
1 5 10 15

Met Ile Asp Gly Trp Tyr Gly Phe Arg His Gln Asn Ser Glu Gly Thr
20 25 30

Gly Gln Ala Ala Asp Leu Lys Ser Thr Gln Ala Ala Ile Asp Gln Ile
35 40 45

Asn Gly Lys Leu Asn Arg Val Ile Glu Lys Thr Asn Glu Lys Phe His
50 55 60

Gln Ile Glu Lys Glu Phe Ser Glu Val Glu Gly Arg Ile Gln Asp Leu
65 70 75 80

Glu Lys Tyr Val Glu Asp Thr Lys Ile Asp Leu Trp Ser Tyr Asn Ala
85 90 95

Glu Leu Leu Val Ala Leu Glu Asn Gln His Thr Ile Asp Leu Thr Asp
100 105 110

Ser Glu Met Asn Lys Leu Phe Glu Lys Thr Arg Arg Gln Leu Arg Glu
115 120 125

Asn Ala Glu Glu Met Gly Asn Gly Cys Phe Lys Ile Tyr His Lys Cys
130 135 140

Asp Asn Ala Cys Ile Glu Ser Ile Arg Asn Gly Thr Tyr Asp His Asp
145 150 155 160

Val Tyr Arg Asp Glu Ala Leu Asn Asn Arg Phe Gln Ile Lys Gly Val
165 170 175

Glu Leu Lys Ser Gly Tyr Lys Asp Trp Ile Leu Trp Ile Ser Phe Ala
180 185 190

Ile Ser Cys Phe Leu Leu Cys Val Val Leu Leu Gly Phe Ile Met Trp
195 200 205

Ala Cys Gln Arg Gly Asn Ile Arg Cys Asn Ile Cys Ile
210 215 220

<210> SEQ ID NO 24
 <211> LENGTH: 222
 <212> TYPE: PRT
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 24

Gly Leu Phe Gly Ala Ile Ala Gly Phe Ile Glu Gly Gly Trp Gln Gly
1 5 10 15

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Met	Ile	Asp	Gly	Trp	Tyr	Gly	Phe	His	His	Ser	Asn	Glu	Gln	Gly	Ser
			20					25					30		
Gly	Tyr	Ala	Ala	Asp	Lys	Glu	Ser	Thr	Gln	Lys	Ala	Ile	Asp	Gly	Thr
		35					40					45			
Thr	Asn	Lys	Val	Asn	Ser	Val	Ile	Asp	Lys	Met	Asn	Thr	Gln	Phe	Glu
		50				55					60				
Ala	Ile	Gly	Lys	Glu	Phe	Asn	Asn	Leu	Glu	Arg	Arg	Ile	Glu	Asn	Leu
65					70					75				80	
Asn	Lys	Lys	Met	Glu	Asp	Gly	Phe	Leu	Asp	Val	Trp	Thr	Tyr	Asn	Ala
				85					90					95	
Glu	Leu	Leu	Val	Leu	Met	Glu	Asn	Glu	Arg	Thr	Leu	Asp	Phe	His	Asp
			100					105					110		
Ser	Asn	Val	Lys	Asn	Leu	Phe	Asp	Lys	Val	Arg	Leu	Gln	Leu	Arg	Asp
		115					120					125			
Asn	Ala	Lys	Glu	Leu	Gly	Asn	Gly	Cys	Phe	Glu	Phe	Tyr	His	Lys	Cys
	130					135					140				
Asp	Asn	Glu	Cys	Met	Glu	Ser	Ile	Lys	Asn	Gly	Thr	Tyr	Asp	Tyr	Pro
145					150					155					160
Gln	Tyr	Ser	Glu	Glu	Ala	Arg	Leu	Asn	Arg	Glu	Glu	Ile	Ser	Gly	Val
			165					170						175	
Lys	Leu	Glu	Ser	Met	Gly	Ile	Tyr	Gln	Ile	Leu	Ser	Ile	Tyr	Ser	Thr
			180					185					190		
Val	Ala	Ser	Ser	Leu	Ala	Leu	Ala	Val	Met	Ile	Ala	Gly	Leu	Ser	Leu
		195					200					205			
Trp	Met	Cys	Ser	Asn	Gly	Ser	Leu	Gln	Cys	Arg	Ile	Cys	Ile		
	210					215					220				

<210> SEQ ID NO 25
 <211> LENGTH: 222
 <212> TYPE: PRT
 <213> ORGANISM: Influenza virus

 <400> SEQUENCE: 25

Gly	Leu	Phe	Gly	Ala	Ile	Ala	Gly	Phe	Ile	Glu	Gly	Gly	Trp	Gln	Gly
1				5					10					15	
Met	Ile	Asp	Gly	Trp	Tyr	Gly	Phe	His	His	Ser	Asn	Glu	Gln	Gly	Ser
		20						25					30		
Gly	Tyr	Ala	Ala	Asp	Lys	Glu	Ser	Thr	Gln	Lys	Ala	Ile	Asp	Gly	Thr
		35					40					45			
Thr	Asn	Lys	Val	Asn	Ser	Val	Ile	Asn	Lys	Met	Asn	Thr	Gln	Phe	Glu
		50				55					60				
Ala	Ile	Gly	Arg	Glu	Phe	Asn	Asn	Leu	Glu	Arg	Arg	Ile	Glu	Asn	Leu
65					70					75				80	
Asn	Lys	Lys	Met	Glu	Asp	Gly	Phe	Leu	Asp	Val	Trp	Thr	Tyr	Asn	Ala
				85					90					95	
Glu	Leu	Leu	Val	Leu	Met	Glu	Asn	Glu	Arg	Thr	Leu	Asp	Phe	His	Asp
			100					105					110		
Ser	Asn	Val	Lys	Asn	Leu	Phe	Asp	Lys	Val	Arg	Leu	Gln	Leu	Arg	Asp
		115					120					125			
Asn	Ala	Lys	Glu	Leu	Gly	Asn	Gly	Cys	Phe	Glu	Phe	Tyr	His	Lys	Cys
	130					135					140				
Asp	Asn	Glu	Cys	Met	Glu	Ser	Ile	Lys	Asn	Gly	Thr	Tyr	Asp	Tyr	Pro
145					150					155					160
Gln	Tyr	Ser	Glu	Glu	Ala	Arg	Leu	Asn	Arg	Glu	Glu	Ile	Ser	Gly	Val
			165					170						175	

Lys	Leu	Glu	Ser	Met	Gly	Thr	Tyr	Gln	Ile	Leu	Ser	Ile	Tyr	Ser	Thr
			180					185					190		
Val	Ala	Ser	Ser	Leu	Ala	Leu	Ala	Val	Met	Val	Ala	Gly	Leu	Ser	Leu
		195					200					205			
Trp	Met	Cys	Ser	Asn	Gly	Ser	Leu	Gln	Cys	Arg	Ile	Cys	Ile		
	210					215					220				

<400> SEQUENCE: 26

[illegible]

<400> SEQUENCE: 27

Gly	Leu	Phe	Gly	Ala	Ile	Ala	Gly	Phe	Ile	Glu	Gly	Gly	Trp	Pro	Gly
1				5					10					15	
Leu	Ile	Ala	Gly	Trp	Tyr	Gly	Phe	Gln	His	Ser	Asn	Asp	Gln	Gly	Val
			20					25					30		
Gly	Met	Ala	Ala	Asp	Arg	Asp	Ser	Thr	Gln	Lys	Ala	Ile	Asp	Lys	Thr
		35					40					45			
Thr	Ser	Lys	Val	Asn	Asn	Val	Ile	Asp	Lys	Met	Asn	Lys	Gln	Phe	Glu
	50					55					60				

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Ile Ile Asp His Glu Phe Asn Asn Leu Glu Thr Arg Leu Asn Met Ile
65              70              75              80

Asn Asn Lys Met Asp Asp Gln Ile Gln Asp Val Trp Ala Tyr Asn Ala
85              90              95

Glu Leu Leu Val Leu Met Glu Asn Glu Lys Thr Leu Asp Glu His Asp
100            105            110

Ala Asn Val Lys Asn Leu Phe Asn Lys Val Lys Leu Ala Leu Gly Ser
115            120            125

Asn Ala Met Glu Asp Gly Lys Gly Cys Phe Glu Leu Tyr His Lys Cys
130            135            140

Asp Asp Gln Cys Met Glu Thr Ile Lys Asn Gly Thr Tyr Asn Arg Arg
145            150            155            160

Lys Tyr Lys Glu Glu Ser Arg Leu Glu Arg Gln Lys Ile Glu Gly
165            170            175

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<210> SEQ ID NO 28
<211> LENGTH: 222
<212> TYPE: PRT
<213> ORGANISM: Influenza virus

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<400> SEQUENCE: 28

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Gly Leu Phe Gly Ala Ile Ala Gly Phe Ile Glu Gly Gly Trp Thr Gly
1      5      10      15

Met Val Asp Gly Trp Tyr Gly Tyr His His Gln Asn Glu Gln Gly Ser
20     25     30

Gly Tyr Ala Ala Asp Gln Lys Ser Thr Gln Asn Ala Ile Asn Gly Ile
35     40     45

Thr Asn Lys Val Asn Ser Val Ile Glu Lys Met Asn Thr Gln Phe Thr
50     55     60

Ala Val Gly Lys Glu Phe Asn Lys Leu Glu Arg Arg Met Glu Asn Leu
65     70     75     80

Asn Lys Lys Val Asp Asp Gly Phe Leu Asp Ile Trp Thr Tyr Asn Ala
85     90     95

Glu Leu Leu Val Leu Leu Glu Asn Glu Arg Thr Leu Asp Phe His Asp
100    105    110

Ser Asn Val Lys Asp Leu Tyr Glu Lys Val Lys Ser Gln Leu Lys Asn
115    120    125

Asn Ala Lys Glu Ile Gly Asn Gly Cys Phe Glu Phe Tyr His Lys Cys
130    135    140

Asn Asn Glu Cys Met Glu Ser Val Lys Asn Gly Thr Tyr Asp Tyr Pro
145    150    155    160

Lys Tyr Ser Glu Glu Ser Lys Leu Asn Arg Glu Lys Ile Asp Gly Val
165    170    175

Lys Leu Glu Ser Met Gly Val Tyr Gln Ile Leu Ala Ile Tyr Ser Thr
180    185    190

Val Ala Ser Ser Leu Val Leu Leu Val Ser Leu Gly Ala Ile Ser Phe
195    200    205

Trp Met Cys Ser Asn Gly Ser Leu Gln Cys Arg Ile Cys Ile
210    215    220

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<210> SEQ ID NO 29
<211> LENGTH: 222
<212> TYPE: PRT
<213> ORGANISM: Influenza virus

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<400> SEQUENCE: 29

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Gly Leu Phe Gly Ala Ile Ala Gly Phe Ile Glu Gly Gly Trp Thr Gly
 1 5 10 15
 Met Val Asp Gly Trp Tyr Gly Tyr His His Gln Asn Glu Gln Gly Ser
 20 25 30
 Gly Tyr Ala Ala Asp Gln Lys Ser Thr Gln Asn Ala Ile Asn Gly Ile
 35 40 45
 Thr Asn Lys Val Asn Ser Val Ile Glu Lys Met Asn Thr Gln Phe Thr
 50 55 60
 Ala Val Gly Lys Glu Phe Asn Lys Leu Glu Arg Arg Met Glu Asn Leu
 65 70 75 80
 Asn Lys Lys Val Asp Asp Gly Phe Ile Asp Ile Trp Thr Tyr Asn Ala
 85 90 95
 Glu Leu Leu Val Leu Leu Glu Asn Glu Arg Thr Leu Asp Phe His Asp
 100 105 110
 Ser Asn Val Lys Asp Leu Tyr Glu Lys Val Lys Ser Gln Leu Lys Asn
 115 120 125
 Asn Ala Lys Glu Ile Gly Asn Gly Cys Phe Glu Phe Tyr His Lys Cys
 130 135 140
 Asn Asp Glu Cys Met Glu Ser Val Lys Asn Gly Thr Tyr Asp Tyr Pro
 145 150 155 160
 Lys Tyr Ser Glu Glu Ser Lys Leu Asn Arg Glu Lys Ile Asp Gly Val
 165 170 175
 Lys Leu Glu Ser Met Gly Val Tyr Gln Ile Leu Ala Ile Tyr Ser Thr
 180 185 190
 Val Ala Ser Ser Leu Val Leu Leu Val Ser Leu Gly Ala Ile Ser Phe
 195 200 205
 Trp Met Cys Ser Asn Gly Ser Leu Gln Cys Arg Ile Cys Ile
 210 215 220

<210> SEQ ID NO 30
 <211> LENGTH: 221
 <212> TYPE: PRT
 <213> ORGANISM: Influenza virus

<400> SEQUENCE: 30

Gly Ile Phe Gly Ala Ile Ala Gly Phe Ile Glu Asn Gly Trp Glu Gly
 1 5 10 15
 Met Val Asp Gly Trp Tyr Gly Phe Arg His Gln Asn Ser Glu Gly Thr
 20 25 30
 Gly Gln Ala Ala Asp Leu Lys Ser Thr Gln Ala Ala Ile Asn Gln Ile
 35 40 45
 Asn Gly Lys Leu Asn Arg Leu Ile Gly Lys Thr Asn Glu Lys Phe His
 50 55 60
 Gln Ile Glu Lys Glu Phe Ser Glu Val Glu Gly Arg Ile Gln Asp Leu
 65 70 75 80
 Glu Lys Tyr Val Glu Asp Thr Lys Ile Asp Leu Trp Ser Tyr Asn Ala
 85 90 95
 Glu Leu Leu Val Ala Leu Glu Asn Gln His Thr Ile Asp Leu Thr Asp
 100 105 110
 Ser Glu Met Asp Lys Leu Phe Glu Arg Thr Lys Lys Gln Leu Arg Glu
 115 120 125
 Asn Ala Glu Asp Met Gly Asn Gly Cys Phe Lys Ile Tyr His Lys Cys
 130 135 140
 Asp Asn Ala Cys Ile Glu Ser Ile Arg Asn Gly Thr Tyr Asp His Asp

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145	150	155	160
Val Tyr Arg Asp Glu Ala Leu Asn Asn Arg Phe Gln Ile Lys Gly Val			
	165	170	175
Glu Leu Lys Ser Gly Tyr Lys Asp Trp Ile Leu Trp Ile Ser Phe Ala			
	180	185	190
Ile Ser Cys Phe Leu Leu Cys Val Ala Leu Leu Gly Phe Ile Met Trp			
	195	200	205
Ala Cys Gln Lys Gly Asn Ile Arg Cys Asn Ile Cys Ile			
	210	215	220

What is claimed is:

1. An isolated Vero cell infected with a recombinant reassortant influenza virus having PA, PB1, PB2, NP, NS, and M gene segments from a first influenza vaccine virus isolate, an influenza virus HA gene segment selected to encode an aspartic acid or glutamic acid at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA, and a NA gene segment that encodes a full-length NA protein that has a tyrosine at position 255, wherein the numbering for NA residues is that for N1.

2. The isolated cell of claim 1, wherein the NA gene segment and the HA gene segment in the reassortant virus are from the same influenza virus isolate.

3. The isolated cell of claim 1, wherein the HA gene segment in the reassortant virus is mutated to encode the aspartic acid or glutamic acid at position 117.

4. The isolated cell of claim 1, wherein the PA, PB1, PB2, NP, NS, and M gene segments in the reassortant virus are from the same influenza virus isolate.

5. The isolated cell of claim 1, wherein the PA, PB1, PB2, NP, NS, and M gene segments in the reassortant virus comprise sequences for at least one of the following: a PB1 having the amino acid sequence encoded by SEQ ID NO:2 or PB1 with at least 95% amino acid sequence identity to the PB1 encoded by SEQ ID NO:2; a PB2 having the amino acid sequence encoded by SEQ ID NO:3 or PB2 with at least 95% amino acid sequence identity to the PB2 encoded by SEQ ID NO:3; a PA having the amino acid sequence encoded by SEQ ID NO:1 or PA with at least 95% amino acid sequence identity to the PA encoded by SEQ ID NO:1; a NP having the amino acid sequence encoded by SEQ ID NO:4 or NP with at least 95% amino acid sequence identity to the NP encoded by SEQ ID NO:4; a M having the amino acid sequence encoded by SEQ ID NO:5 or M with at least 95% amino acid sequence identity to the M encoded by SEQ ID NO:5; or a NS having the amino acid sequence encoded by SEQ ID NO:6 or NS with at least 95% amino acid sequence identity to the NS encoded by SEQ ID NO:6.

6. The isolated cell of claim 1, wherein the PA, PB1, PB2, NP, NS, and M gene segments in the reassortant virus comprise sequences for at least one of the following: a PB1 having the amino acid sequence encoded by SEQ ID NO:10 or PB1 with at least 95% amino acid sequence identity to the PB1 encoded by SEQ ID NO:10; a PB2 having the amino acid sequence encoded by SEQ ID NO:11 or PB2 with at least 95% amino acid sequence identity to the PB2 encoded by SEQ ID NO:11; a PA having the amino acid sequence encoded by SEQ ID NO:12 or PA with at least 95% amino acid sequence identity to the PA encoded by SEQ ID NO:12; a NP having the amino acid sequence encoded by SEQ ID NO:13 or NP with at least 95% amino acid sequence identity to the NP encoded by SEQ ID NO:13; a M having the amino

15 acid sequence encoded by SEQ ID NO:14 or M with at least 95% amino acid sequence identity to the M encoded by SEQ ID NO:14; or a NS having the amino acid sequence encoded by SEQ ID NO:15 or NS with at least 95% amino acid sequence identity to the NS encoded by SEQ ID NO:15.

7. An isolated recombinant reassortant influenza virus comprising a HA gene segment selected to encode an aspartic acid or glutamic acid at position 117 in HA2 and comprising a NA gene segment that encodes a full-length NA protein that has a tyrosine at position 255, wherein the numbering for NA residues is that for N1, wherein the recombinant influenza virus has enhanced replication in Vero cells relative to a corresponding influenza virus that does not have aspartic acid or glutamic acid at position 117 in HA2, wherein the numbering for HA2 residues is that for H1 HA2.

8. The isolated recombinant virus of claim 7, wherein the corresponding virus has an alanine, asparagine, arginine or lysine at position 117 in HA2.

9. The isolated recombinant virus of claim 7, which has a NA gene segment that is from the same influenza isolate as the HA gene segment.

10. The isolated recombinant virus of claim 7, wherein the HA gene segment is mutated to encode the aspartic acid or glutamic acid at position 117.

11. The isolated recombinant virus of claim 7, wherein the PA, PB1, PB2, NP, NS, and M gene segments are from the same influenza virus isolate.

12. The isolated recombinant virus of claim 7, wherein the PA, PB1, PB2, NP, NS, and M gene segments comprise sequences for a PB1 having the amino acid sequence encoded by SEQ ID NO:2 or PB1 with at least 95% amino acid sequence identity to the PB1 encoded by SEQ ID NO:2; a PB2 having the amino acid sequence encoded by SEQ ID NO:3 or PB2 with at least 95% amino acid sequence identity to the PB2 encoded by SEQ ID NO:3; a PA having the amino acid sequence encoded by SEQ ID NO:1 or PA with at least 95% amino acid sequence identity to the PA encoded by SEQ ID NO:1; a NP having the amino acid sequence encoded by SEQ ID NO:4 or NP with at least 95% amino acid sequence identity to the NP encoded by SEQ ID NO:4; a M having the amino acid sequence encoded by SEQ ID NO:5 or M with at least 95% amino acid sequence identity to the M encoded by SEQ ID NO:5; or a NS having the amino acid sequence encoded by SEQ ID NO:6 or NS with at least 95% amino acid sequence identity to the NS encoded by SEQ ID NO:6.

13. The isolated recombinant virus of claim 7, wherein the PA, PB1, PB2, NP, NS, and M gene segments comprise sequences for a PB1 having the amino acid sequence encoded by SEQ ID NO:10 or PB1 with at least 95% amino acid sequence identity to the PB1 encoded by SEQ ID NO:10; a PB2 having the amino acid sequence encoded by SEQ ID NO:11 or PB2 with at least 95% amino acid sequence identity

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to the PB2 encoded by SEQ ID NO:11; a PA having the amino acid sequence encoded by SEQ ID NO:12 or PA with at least 95% amino acid sequence identity to the PA encoded by SEQ ID NO:12; a NP having the amino acid sequence encoded by SEQ ID NO:13 or NP with at least 95% amino acid sequence identity to the NP encoded by SEQ ID NO:13; a M having the amino acid sequence encoded by SEQ ID NO:14 or M with at least 95% amino acid sequence identity to the M encoded by SEQ ID NO:14; or a NS having the amino acid sequence encoded by SEQ ID NO:15 or NS with at least 95% amino acid sequence identity to the NS encoded by SEQ ID NO:15.

14. The isolated recombinant virus of claim 7, wherein the HA2 that has an aspartic acid or glutamic acid at position 117 in HA2 has at least 80% amino acid sequence identity to one of SEQ ID Nos. 22-27.

15. The isolated recombinant virus of claim 7, wherein the HA gene segment is a H1, H2, H3, H5, H7, or H9 gene segment.

16. A vaccine having the isolated recombinant virus of claim 7.

17. A method to prepare the recombinant reassortant influenza virus of claim 7, comprising: transfecting a cell with:

a vector for vRNA production comprising a promoter operably linked to an influenza virus PA DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus PB1 DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus PB2 DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus HA DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus NP DNA linked to a transcription termination sequence, a vector for vRNA production comprising a promoter operably linked to an influenza virus NA DNA linked to a transcription termination sequence, and a vector for vRNA production comprising a promoter operably linked to an influenza virus NS DNA linked to a transcription termination sequence, and wherein the HA DNA in the vector for vRNA production of HA is selected to encode an aspartic acid or glutamic acid at position 117 in HA2, and wherein the NA DNA in the vector for vRNA production of NA encodes for a tyrosine at position 255, wherein the numbering for NA residues is that for N1, wherein the numbering for HA2 residues is that for H1 HA2; and

a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus PA, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding

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influenza virus PB1, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus PB2, and a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus NP, and optionally a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus HA, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus NA, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus M1, a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus M2, or a vector for mRNA production comprising a promoter operably linked to a DNA segment encoding influenza virus NS2;

in an amount effective to yield infectious influenza virus; and isolating the recombinant reassortant influenza virus.

18. The method of claim 17, wherein the vRNA vectors include sequences for a PB1 polypeptide having the amino acid sequence encoded by SEQ ID NO:2 or a PB1 polypeptide with at least 95% amino acid sequence identity to the PB1 polypeptide encoded by SEQ ID NO:2, a PB2 polypeptide having the amino acid sequence encoded by SEQ ID NO:3 or a PB2 polypeptide with at least 95% amino acid sequence identity to the PB2 polypeptide encoded by SEQ ID NO:3, a PA polypeptide having the amino acid sequence encoded by SEQ ID NO:1 or a PA polypeptide with at least 95% amino acid sequence identity to the PA polypeptide encoded by SEQ ID NO:1, a NP polypeptide having the amino acid sequence encoded by SEQ ID NO:4 or a NP polypeptide with at least 95% amino acid sequence identity to the NP polypeptide encoded by SEQ ID NO:4, a M polypeptide having the amino acid sequence encoded by SEQ ID NO:5 or a M polypeptide with at least 95% amino acid sequence identity to the M encoded by SEQ ID NO:5, and a NS polypeptide having the amino acid sequence encoded by SEQ ID NO:6 or 15 or a NS polypeptide with at least 95% amino acid sequence identity to the NS encoded by SEQ ID NO:6 or 15.

19. The method of claim 17, wherein the cell is an isolated avian cell.

20. The method of claim 17, wherein the cell is an isolated mammalian cell.

21. The method of claim 20, wherein the isolated mammalian cell is a Vero cell, an isolated human cell or an isolated hamster cell.

22. The method of claim 17, wherein the wherein the PB1, PB2, PA, NP, NS, and M DNAs in the vectors for vRNA productions have a sequence that corresponds to one that encodes a polypeptide having at least 95% amino acid sequence identity to a corresponding polypeptide encoded by SEQ ID NOs:1-6 or 10-15.

* * * * *