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(54) **PLANTS WITH ALTERED PHYTOCHROMES**

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C12N 15/82 (2006.01)
C07K 14/415 (2006.01)

(52) **U.S. Cl.**
CPC **C12N 15/8269** (2013.01); **C07K 14/415** (2013.01); **C12N 15/8261** (2013.01)

(58) **Field of Classification Search**
None
See application file for complete search history.

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(57) **ABSTRACT**

Polynucleotides encoding polypeptides that increase the light sensitivity of plants were identified. Introduction of the polynucleotides into plants produces plants having altered characteristics, such as decreased height, decreased diameter, decreased petiole length, decreased internode length, decreased hypocotyl length, increased hyponasty or enhanced germination.

21 Claims, 18 Drawing Sheets

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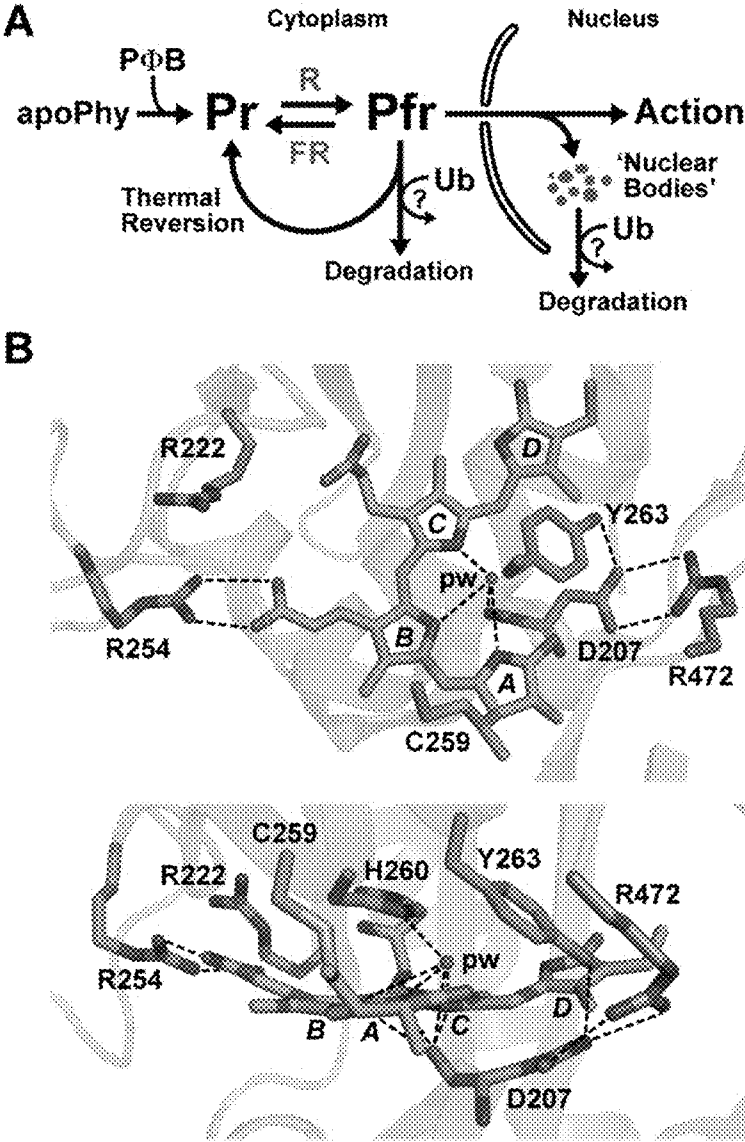


FIGURE 1

C

	Seq Id #	Seq Id #	Seq Id #	Seq Id #	Seq Id #	Seq Id #
Syn Cph1	D207	R222	R254	Y263	R472	
Dr BphP	ESDIPQ	PIEVI	IFRSA	LTYEK	HPRQSF	46
Pa BphP	NSDIPA	LIRIT	VFRAT	MDYER	GPRSSF	51
Rp BphP3	ASDIPA	PIRIT	VFRSV	CEVLT	TPRCSSF	56
SyB Cph1	SSDIPA	PVRIT	VFRSV	KEYNV	QTRASSF	61
At phyA	AGDIPE	QVRVI	LQRPV	VHIEK	LPLISF	66
At phyB	ADDIPO	KVRMI	ELRAP	LOYEA	HPRSSF	76
At phyC	ADDIPO	RVRMI	ELRAP	SOYEA	HPRSSF	76
At phyD	ADDIPO	KVRMI	ELRAP	AOYEA	HPRSSF	76
At phyE	ADDIPO	RVRMI	ELRAP	TOYEA	NPRSSF	77
At phyB	D307	R322	R352	Y361	R582	
At phyA	D273	R288	R318	Y327	R551	

FIGURE 1

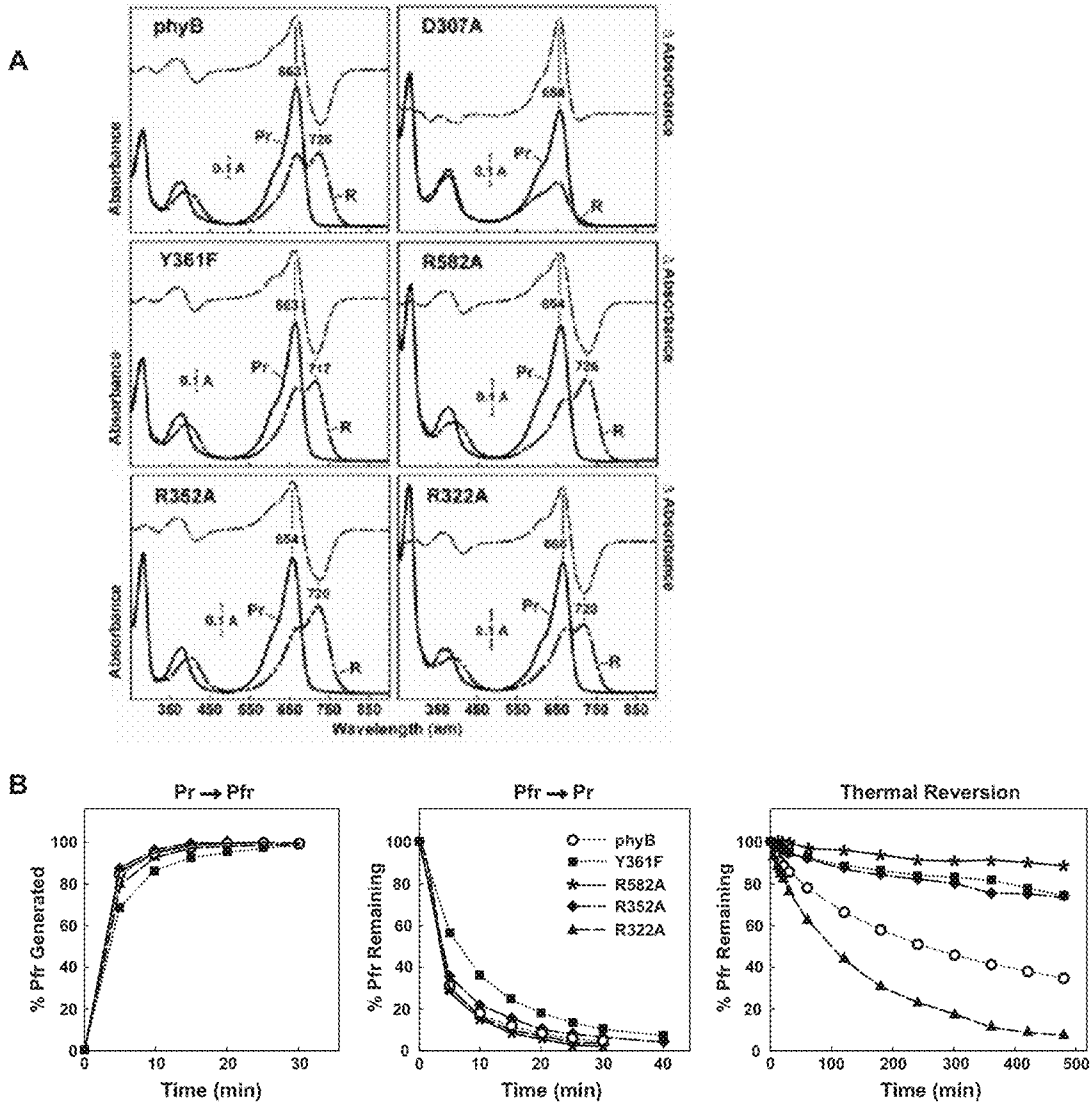


FIGURE 2

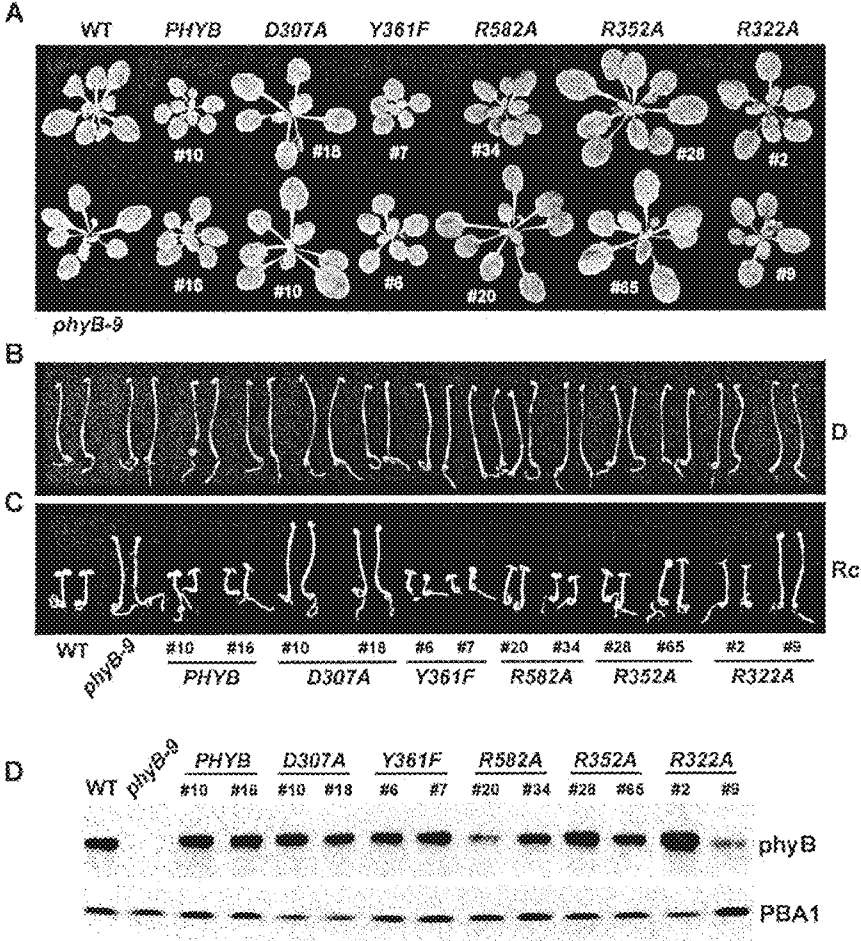


FIGURE 3

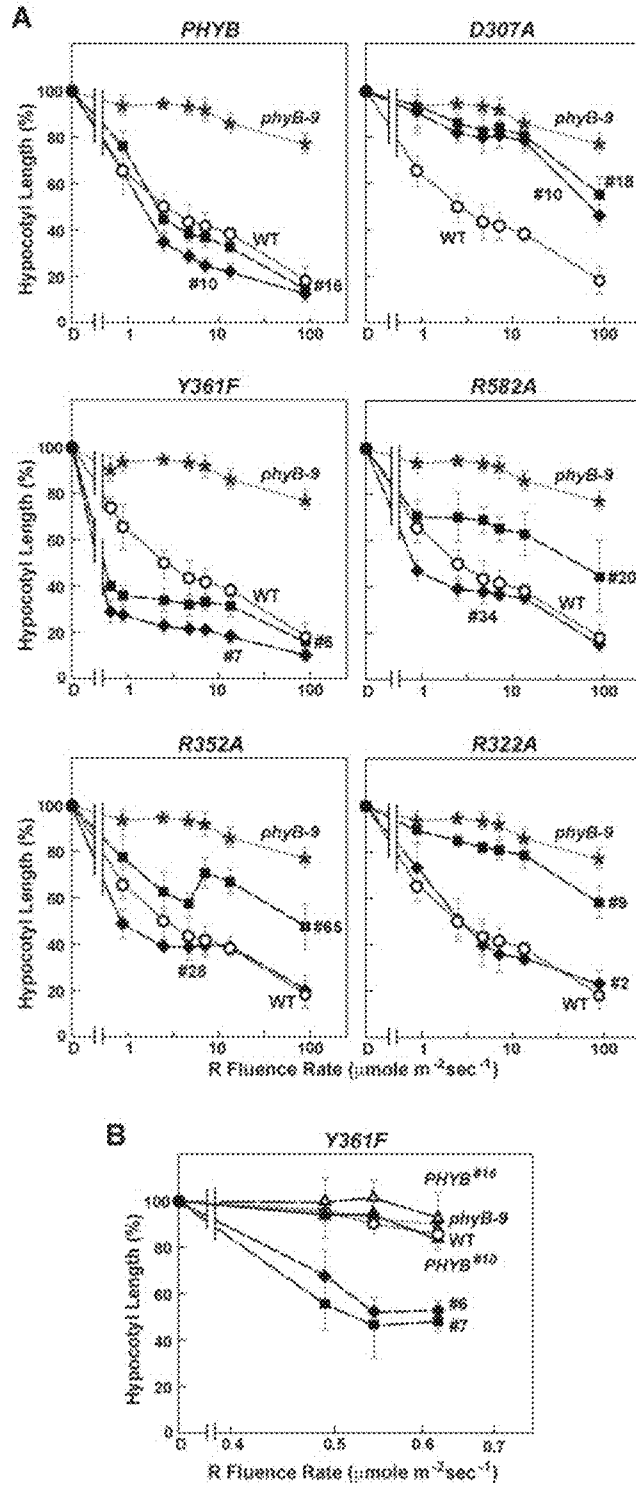


FIGURE 4

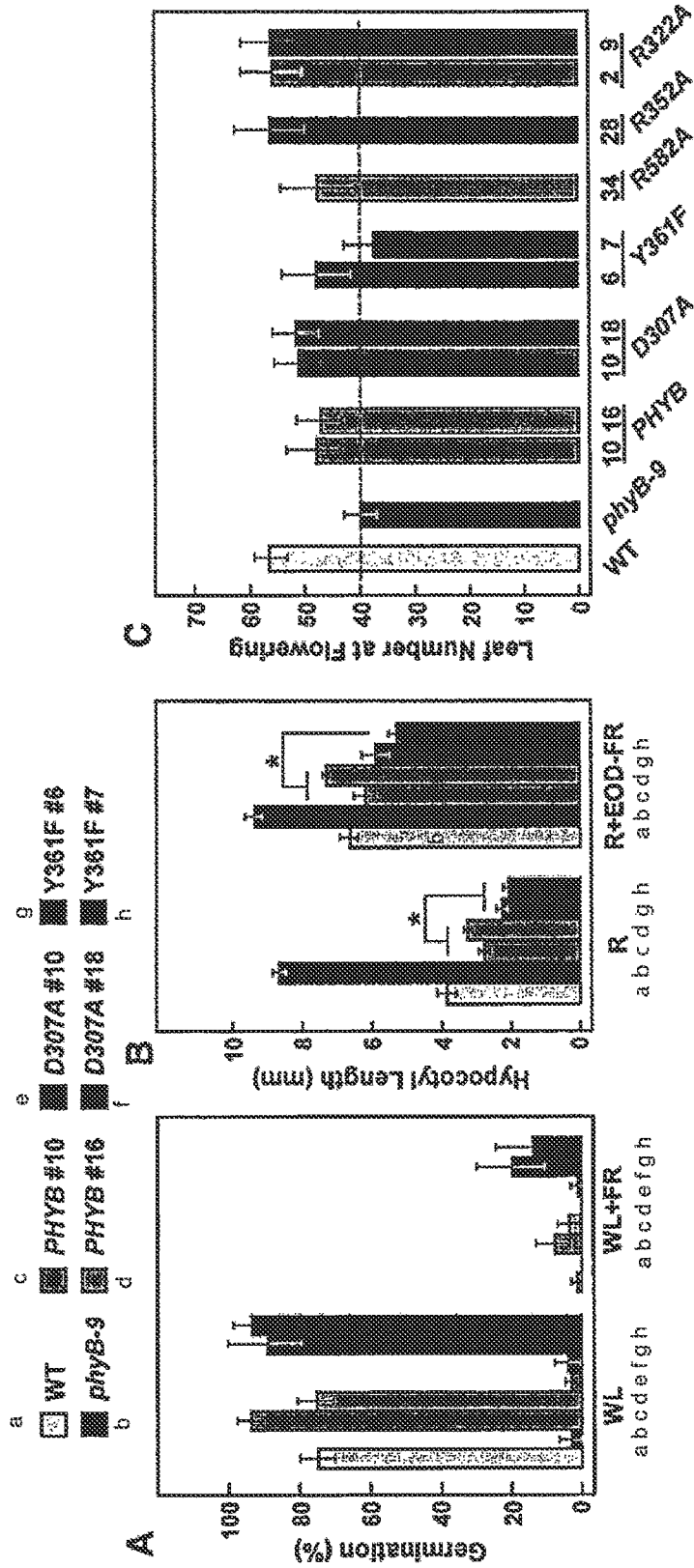


FIGURE 5

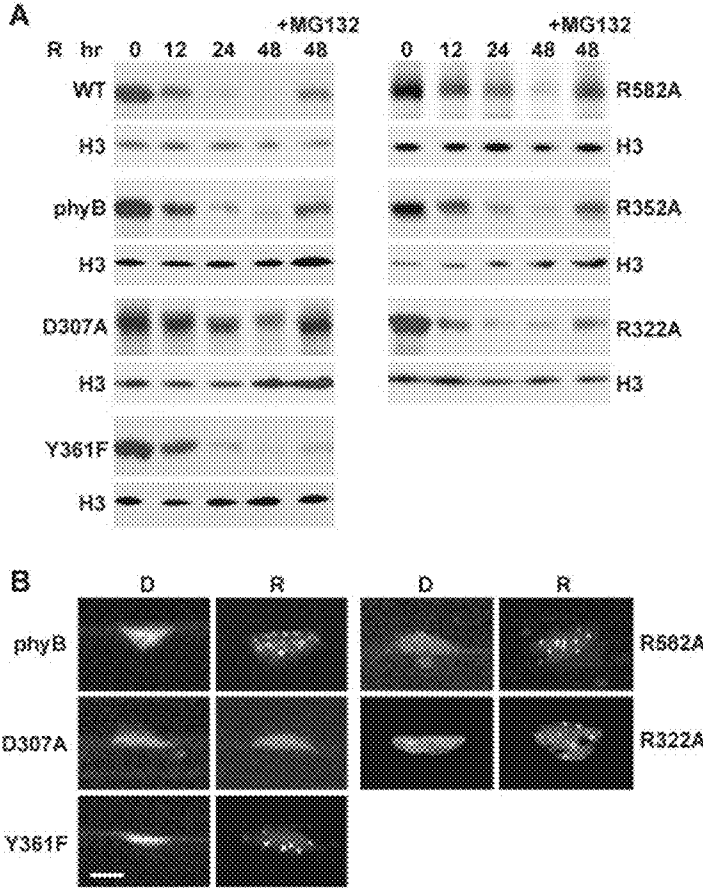


FIGURE 6

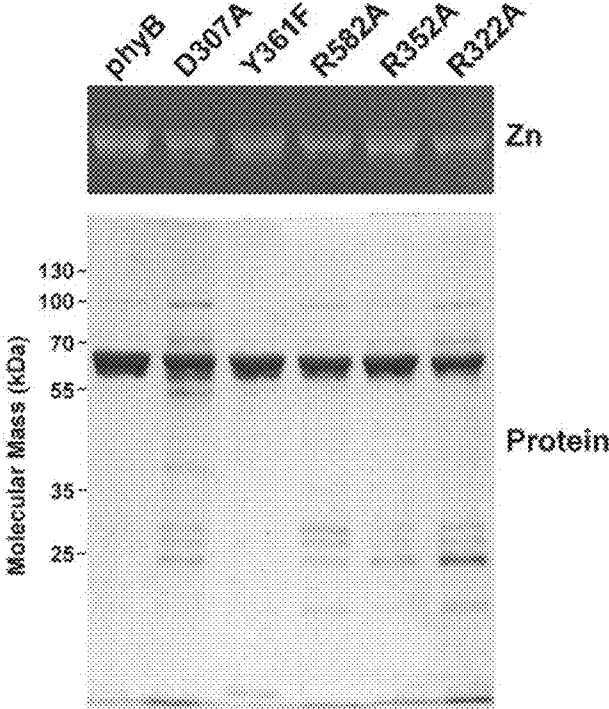


FIGURE 7

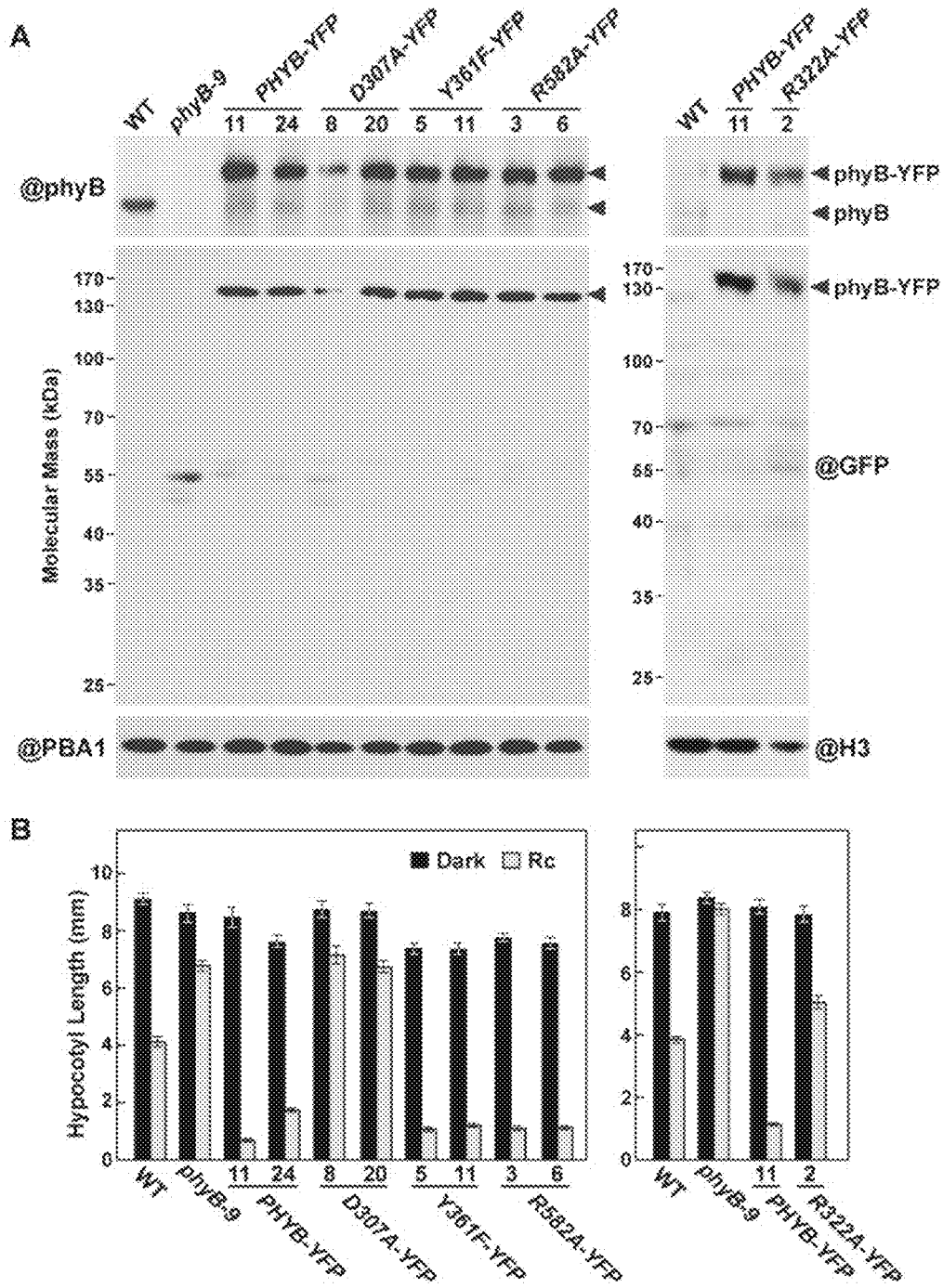


FIGURE 8

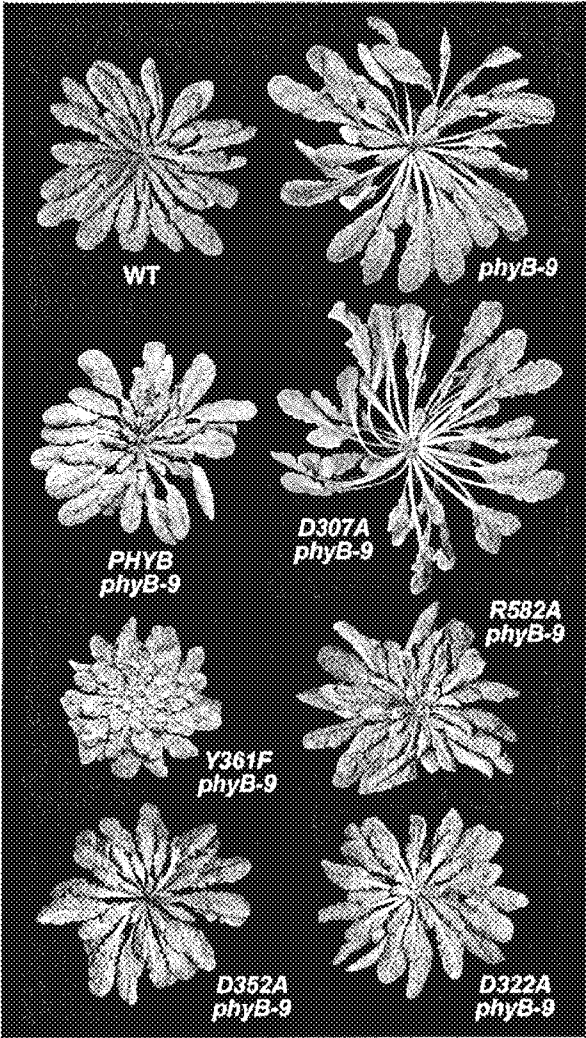


FIGURE 9

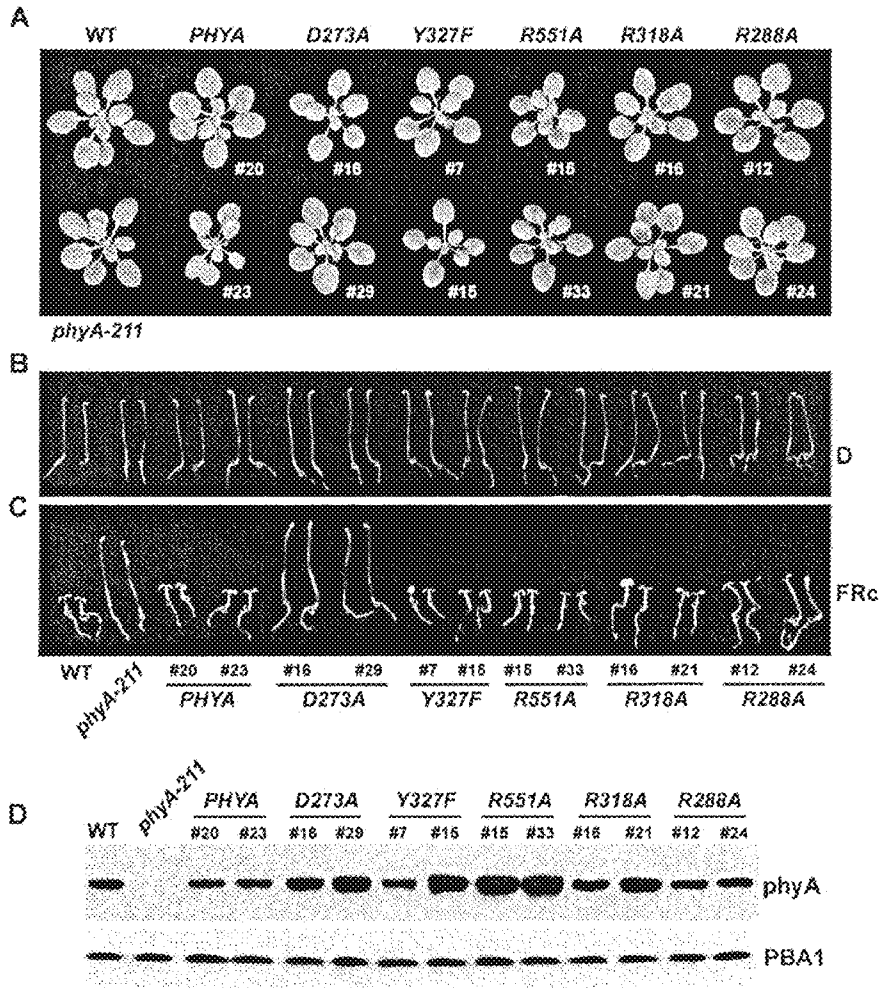


FIGURE 10

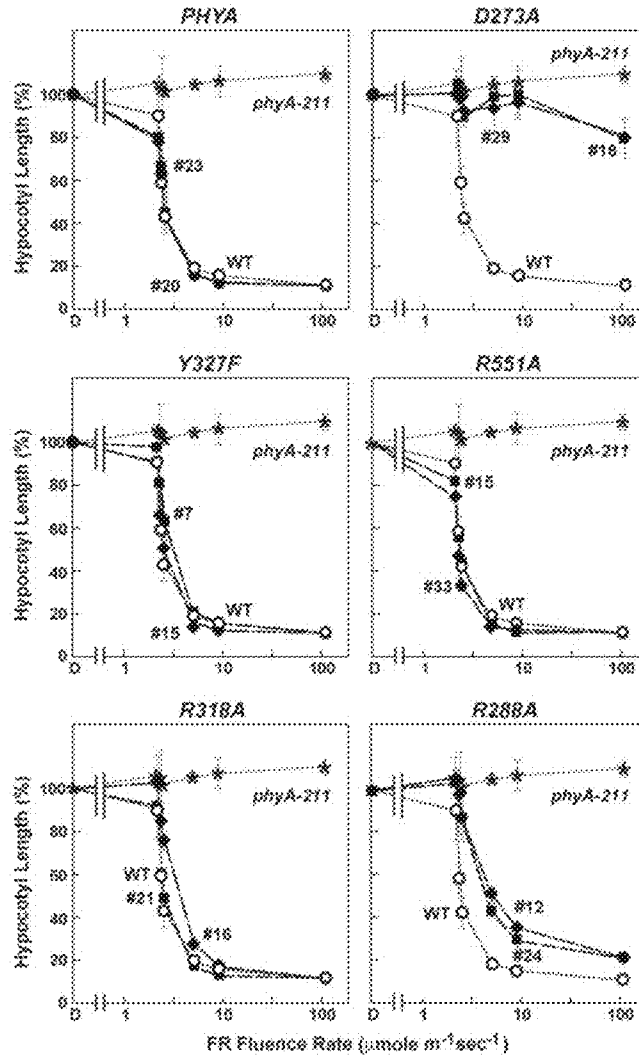


FIGURE 11

At phyB 1 MVSGVGGSGGGRGGGRGGEEPPSSSHTPNNRRGGEQAQSSGTSKSLRPRS.....
 ZmphyB 1MASGSRATPTRSPSSARPEAPRHAHHHHHSQS..SGGTSRAGGG.....
 Os phyB 1MGSGRATPTRSPSSARP AAPRHQHHSQS5G...GSTRAGGGGGGGGGGGG.
 Sb phyB 1MASGSRATPTRSPSSARPEAPRHAHHHHHHHSQS5GSTRAGGGGGGGGGGGT A
 GmphyB1 1MASASGAANSVPP.....PQIHSTRKLSHHSSNNNNN.....
 GmphyB2 1MASASGAENSSVPPSP LPPPPPPQIHSTRKLSHHHHNNNNNNNN.....
 GmphyB3 1
 GmphyB4 1
 St phyB 1MASGSRTKHS HHNSQAQSSGTSNVN.....
 Ps phyB 1SNNNNNRN IKR.....
 Vv phyB 1MSSGNRGTQS HHQAQSS.GTSNLRVY.....

At phyB 50NTESMSKAIQOYTDARLHAVFEQSGESGKSF DYSQSLKTTTYGSSVPEQQIT A
 ZmphyB 44AAATESVSKAVAQYTL DARLHAVFEQSGASGRSFDYSQSLRAPPTP..SSEQQIAA
 Os phyB 52AAAAESVSKAVAQYTL DARLHAVFEQSGASGRSFDYTOSLRAPPTP..SSEQQIAA
 Sb phyB 57 ATATATATESVSKAVAQYTL DARLHAVFEQSGASGRSFDYSQSLRAPPTP..SSEQQIAA
 GmphyB1 35IDSMSKAIQOYTE DARLHAVFEQSGESGRSFDYSQSLRAPPTP..SSEQQIT A
 GmphyB2 46NIDSTS KAIQOYTE DARLHAVFEQSGESGRSFDYSQSLRAPPTP..SSEQQIT A
 GmphyB3 1MSKAIQOYTE DARLHAVFEQSGESGRSFDYSQSLRAPPTP..SSEQQIT A
 GmphyB4 1
 St phyB 27YKDSMSKAIQOYTDARLHAVFEQSGESGKSF DYSQSLKTTTYGSSVPEQQIT A
 Ps phyB 12ESLSMRKAIQOYTDARLHAVFEQSGESGRSFDYSQSLRAPPTP..SSEQQIT A
 Vv phyB 26HTDSMSKAIQOYTDARLHAVFEQSGESGKSF DYSQSLKTTTYGSSVPEQQIT A

At phyB 104 YLSRIQRGGV IQPFGCMI AVDESS FRVLAFFSENAREMILGLTPQSVPSLEKPE...ILL
 ZmphyB 98 YLSRIQRGGH IQPFGCTLAVADDSS FRVLAFFSENSPDL DLSPHHSVPSLDS.SAPPHVS
 Os phyB 106 YLSRIQRGGH IQPFGCTLAVADDSS FRVLAFFSENTADL DLSPHHSVPSLDSAVPPPVS
 Sb phyB 115 YLSRIQRGGH IQPFGCTLAVADDSS FRVLAFFSENADL DLSPHHSVPSLDS.AAPPPVS
 GmphyB1 86 YLVKIQRGGF IQPFGSMI AVDEPS FRVLAFFSDNARDMLGITPQSVPSLDDKN..DAafa
 GmphyB2 98 YLVKIQRGGF IQPFGSMI AVDEPS FRVLAFFSDNARDMLGITPQSVPSLDDKN..DAafa
 GmphyB3 49 YLVKIQRGGF IQPFGSMI AVDEPS FRVLAFFSDNARDMLGITPQSVPSLDDKN..DAafa
 GmphyB4 1MI AVDEPS FRVLAFFSDNARDMLGITPQSVPSLDDKN..DAafa
 St phyB 79 YLVKIQRGGH IQPFGCMI AVDEAS FRVLAFFSENAREMILGLTPQSVPSLEKCE...ILL
 Ps phyB 63 YLAKIQRGGF IQPFGSMI AVDETS FRVLAFFSENARDMLGITAPQSVPSME DSSSSSFFS
 Vv phyB 78 YLSKIQRGGH IQPFGCMI AVDEAT FRVLAFFSENAREMILGLTPQSVPSLEKPE...ILL

At phyB 159 MGTDVRS LFTSSSSILERAFVAREITL LNPVWIHSKNTGKPFYA I LHRIDVGVV IDLEP
 ZmphyB 157 LGADARL LFPSSAVLLEAF AAREISLLNP IWIHSRVSSKPFYA I LHRIDVGVV IDLEP
 Os phyB 166 LGADARL LFPSSAVLLEAF AAREISLLNP IWIHSRVSSKPFYA I LHRIDVGVV IDLEP
 Sb phyB 174 LGADARL LFPSSAVLLEAF AAREISLLNP IWIHSRVSSKPFYA I LHRIDVGVV IDLEP
 GmphyB1 143 LGTDVRA LFTSSALLLEKAFSAREISL MNP IWIHSRTSGKPFYGI LHRIDVGI V IDLEP
 GmphyB2 155 LGTDVRT LFTSSAVLLEKAFSAREISL MNP IWIHSRTSGKPFYGI LHRIDVGI V IDLEP
 GmphyB3 106 LGTDVRA LFTSSALLLEKAFSAREISL MNP IWIHSRTSGKPFYGI LHRIDVGI V IDLEP
 GmphyB4 42 LGTDVRT LFTSSAVLLEKAFSAREISL MNP IWIHSRTSGKPFYGI LHRIDVGI V IDLEP
 St phyB 134 LGTDVRT LFTSSSVLLEAF AAREITL LNP IWIHSKNSGKPFYA I LHRIDVGI V IDLEP
 Ps phyB 122 LGTDVRS LFPSSSVLLEKAFSAREISL MNP IWIHSRTSGKPFYGI LHRIDVGI V IDLEP
 Vv phyB 133 VGTDVRT LFTSSAVLLEKAFRAREITL LNPVWIHSKNSGKPFYA I LHRIDVGI V IDLEP

At phyB 219 ARTEDPALS IAGAVQSKLAVRAISQL QAL PGGDIKLL CDTVVESVRELTGYDRVMVYKF
 ZmphyB 217 ARTEDPALS IAGAVQSKLAVRAISRL QAL PGGDVKLL CDTVVEHVRELTGYDRVMVYRF
 Os phyB 226 ARTEDPALS IAGAVQSKLAVRAISRL QAL PGGDVKLL CDTVVEHVRELTGYDRVMVYRF
 Sb phyB 234 ARTEDPALS IAGAVQSKLAVRAISRL QAL PGGDIKLL CDTVVEHVRELTGYDRVMVYKF
 GmphyB1 203 ARTEDPALS IAGAVQSKLAVRAISQL QSL PGGDVKLL CDTVVESVRELTGYDRVMVYKF
 GmphyB2 215 ARTEDPALS IAGAVQSKLAVRAISQL QSL PGGDVKLL CDTVVESVRELTGYDRVMVYRF
 GmphyB3 166 ARTEDPALS IAGAVQSKLAVRAISQL QSL PGGDVKLL CDTVVESVRELTGYDRVMVYKF
 GmphyB4 102 ARTEDPALS IAGAVQSKLAVRAISQL QSL PGGDVKLL CDTVVESVRELTGYDRVMVYRF
 St phyB 194 ARTEDPALS IAGAVQSKLAVRAISQL QAL PGGDIKLL CDTVVESVRELTGYDRVMVYKF
 Ps phyB 182 ARTEDPALS IAGAVQSKLAVRAISQL QAL PGGDVKLL CDTVVESVRELTGYDRVMVYKF
 Vv phyB 193 ARTEDPALS IAGAVQSKLAVRAISQL QSL PGGDINLL CDTVVENVRELTGYDRVMVYKF

FIG. 12A

			▼ 307	▼ 322
<i>At phyB</i>	279	HEDEHGEVVAESKR	DDLEPYIGLHYPATDIPQASRFLFKQNRVRMVDGNATPVLV	VVQDD
<i>ZmphyB</i>	277	HEDEHGEVVAESRR	DNLEPYIGLHYPATDIPQASRFLFKQNRVRMADCHATPVRV	IQDP
<i>Os phyB</i>	286	HEDEHGEVVAESRR	SNLEPYIGLHYPATDIPQASRFLFKQNRVRMADCHAAPVRV	IQDP
<i>Sb phyB</i>	294	HEDEHGEVVAESRR	DNLEPYIGLHYPATDIPQASRFLFKQNRVRMADCHATPVRV	IQDP
<i>GmphyB1</i>	263	HEDEHGEVVAESKR	DDLEPYIGLHYPATDIPQASRFLFKQNRVRMVDCHASAVRV	VQDE
<i>GmphyB2</i>	275	HEDEHGEVVAETKR	DDLEPYIGLHYPATDIPQASRFLFKQNRVRMVDCHASAVRV	VQDE
<i>GmphyB3</i>	226	HEDEHGEVVAESKR	DDLEPYIGLHYPATDIPQASRFLFKQNRVRMVDCHASAVRV	VQDE
<i>GmphyB4</i>	162	HEDEHGEVVAETKR	DDLEPYIGLHYPATDIPQASRFLFKQNRVRMVDCHASAVRV	VQDE
<i>St phyB</i>	254	HEDEHGEVVAESKRS	DDLEPYIGLHYPATDIPQASRFLFKQNRVRMVDCHATPVRV	VQDE
<i>Ps phyB</i>	242	HEDEHGEVVAESKR	VDLEPYIGLHYPATDIPQASRFLFKQNRVRMVDCHASAVRV	VQDE
<i>Vv phyB</i>	253	HEDEHGEVVAESKRS	DDLEPYIGLHYPATDIPQASRFLFKQNRVRMVDCHATPVLV	VQDE
			▼ 352	▼ 361
<i>At phyB</i>	339	RLTQSMCLVGS	TLRAPHGCHSQYMANMGSIASLAMAVI	NGN.EDDGSNVASG.RSSMRL
<i>ZmphyB</i>	337	GLSQPLCLVGS	TLRAPHGCHAQYMANMGSIASLVMAVI	SSG.GDDEQTGRGGISSAVKLL
<i>Os phyB</i>	346	ALTQPLCLVGS	TLRAPHGCHAQYMANMGSIASLVMAVI	SSGGDDDHNIARGSPISAVKLL
<i>Sb phyB</i>	354	GMSQPLCLVGS	TLRAPHGCHAQYMANMGSIASLVMAVI	SSG.GDDEQTGRGGISSAVKLL
<i>GmphyB1</i>	323	ALVQPLCLVGS	TLRAPHGCHAQYMANMGSIASLVMAVI	NGN.DEEGVGG...RSSMRL
<i>GmphyB2</i>	335	ALVQPLCLVGS	TLRAPHGCHAQYMANMGSIASLVMAVI	NGN.DEEGVGG...RSSMRL
<i>GmphyB3</i>	286	ALVQPLCLVGS	TLRAPHGCHAQYMANMGSIASLVMAVI	NGN.DEEGVGG...RSSMRL
<i>GmphyB4</i>	222	ALVQPLCLVGS	TLRAPHGCHAQYMANMGSIASLVMAVI	NGN.DEEGVGG...RSSMRL
<i>St phyB</i>	314	SLMQPLCLVGS	TLRAPHGCHAQYMANMGSIASLVMAVI	NGN.DEEGVGG...RSSMRL
<i>Ps phyB</i>	302	GLVQPLCLVGS	TLRAPHGCHAQYMANMGSIASLAMAVI	NGN.DEDGGIGGAARGSMRL
<i>Vv phyB</i>	313	GLMQPLCLVGS	TLRAPHGCHAQYMANMGSIASLAMAVI	INGS.DEBATGG...RNLMLL
<i>At phyB</i>	397	WGLVVCCHHTS	ARCIPFPLRYACEFLMQAFGLQLNMELQLA	QNSEKRVLRQTQLCDMLL
<i>ZmphyB</i>	396	WGLVVCCHHTS	PRCIPFPLRYACEFLMQAFGLQLNMELQLA	HQSEKHLRQTQLCDMLL
<i>Os phyB</i>	406	WGLVVCCHHTS	PRCIPFPLRYACEFLMQAFGLQLNMELQLA	HQSEKHLRQTQLCDMLL
<i>Sb phyB</i>	413	WGLVVCCHHTS	PRCIPFPLRYACEFLMQAFGLQLNMELQLA	HQSEKHLRQTQLCDMLL
<i>GmphyB1</i>	378	WGLVVCCHHTS	ARCIPFPLRYACEFLMQAFGLQLNMELQLA	AGSEKRVLRQTQLCDMLL
<i>GmphyB2</i>	390	WGLVVCCHHTS	ARCIPFPLRYACEFLMQAFGLQLNMELQLA	AGSEKRVLRQTQLCDMLL
<i>GmphyB3</i>	341	WGLVVCCHHTS	ARCIPFPLRYACEFLMQAFGLQLNMELQLA	AGSEKRVLRQTQLCDMLL
<i>GmphyB4</i>	277	WGLVVCCHHTS	ARCIPFPLRYACEFLMQAFGLQLNMELQLA	AGSEKRVLRQTQLCDMLL
<i>St phyB</i>	370	WGLVVCCHHTS	VRSIPFPLRYACEFLMQAFGLQLNMELQLA	AGSEKRVLRQTQLCDMLL
<i>Ps phyB</i>	361	WGLVVCCHHTS	ARCIPFPLRYACEFLMQAFGLQLNMELQLA	AGSEKRVLRQTQLCDMLL
<i>Vv phyB</i>	368	WGLVVCCHHTS	ARCIPFPLRYACEFLMQAFGLQLNMELQLA	AGSEKRVLRQTQLCDMLL
<i>At phyB</i>	457	RDSPAGIVTQSPS	IMDLVKCDGAALYYHGKYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>ZmphyB</i>	456	RDSPAGIVTQSPS	IMDLVKCDGAALYYHGKYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>Os phyB</i>	466	RDSPAGIVTQSPS	IMDLVKCDGAALYYHGKYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>Sb phyB</i>	473	RDSPAGIVTQSPS	IMDLVKCDGAALYYHGKYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>GmphyB1</i>	438	RDSPAGIVTQSPS	IMDLVKCDGAALYYOGNYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>GmphyB2</i>	450	RDSPAGIVTQSPS	IMDLVKCDGAALYYOGNYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>GmphyB3</i>	401	RDSPAGIVTQSPS	IMDLVKCDGAALYYOGNYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>GmphyB4</i>	337	RDSPAGIVTQSPS	IMDLVKCDGAALYYOGNYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>St phyB</i>	430	RDSPAGIVTQSPS	IMDLVKCDGAALYYOGKYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>Ps phyB</i>	421	RDSHTGIVTQSPS	IMDLVKCDGAALYYOGNYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>Vv phyB</i>	428	RDSPAGIVTQSPS	IMDLVKCDGAALYYOGKYYPLGVTPTESQ	IKDIEWLLAFHGSDTGL
<i>At phyB</i>	517	STDLSLGDAGYPGAASL	GDAVCGMAVAYITKRDFLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>ZmphyB</i>	516	STDLSLADAGYLGAASL	GDAVCGMAVAYITPSDYLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>Os phyB</i>	526	STDLSLADAGYSGAASL	GDAVCGMAVAYITPSDYLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>Sb phyB</i>	533	STDLSLADAGYLGAAAL	GDAVCGMAVAYITPSDYLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>GmphyB1</i>	498	STDLSLGDAGYPGAASL	GDAVCGMAVAYITEKDFLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>GmphyB2</i>	510	STDLSLADAGYPGAASL	GDAVCGMAVAYITEKDFLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>GmphyB3</i>	461	STDLSLGDAGYPGAASL	GDAVCGMAVAYITEKDFLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>GmphyB4</i>	397	STDLSLADAGYPGAASL	GDAVCGMAVAYITEKDFLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>St phyB</i>	490	STDLSLADAGYPGAASL	GDAVCGMAVAYISSKDFLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>Ps phyB</i>	481	STDLSLADAGYPGAASL	GDAVCGMAVAYITEKDFLFWFRSHTAKEIKWGGAKHHPEDKDDG	
<i>Vv phyB</i>	488	STDLSLADAGYPGAASL	GDAVCGMAVAYITSRDFLFWFRSHTAKEIKWGGAKHHPEDKDDG	

FIG. 12B

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At phyB 577 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 ZmphyB 576 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 Os phyB 586 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 Sb phyB 593 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 GmphyB1 558 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 GmphyB2 570 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 GmphyB3 521 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 GmphyB4 457 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 St phyB 550 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 Ps phyB 541 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV
 Vv phyB 548 QRMHPRSSFQAFLEVVKSRSLPWENAEMDAIHS LQL ILRDSFKDAEHR. NSKAVLDPHV

At phyB 636 PCDMAQEGQIDELGAVAREMVRLETATVPF AVDAGGCINGWNAKIAELTGLSVEEAM
 ZmphyB 636 QLRELELRGIDELSSVAREMVRLETATVPF AVDTDGCINGWNAKIAELTGLSVEEAM
 Os phyB 646 QLRELELRGIDELSSVAREMVRLETATVPF AVDTDGCINGWNAKIAELTGLSVEEAM
 Sb phyB 653 QLRELELRGIDELSSVAREMVRLETATVPF AVDTDGCINGWNAKIAELTGLSVEEAM
 GmphyB1 616 SELELQGGIDELSSVAREMVRLETATVPF AVDVGHVINGWNAKIAELTGLSVEEAM
 GmphyB2 628 SELELQGGIDELSSVAREMVRLETATVPF AVDVGHVINGWNAKIAELTGLSVEEAM
 GmphyB3 579 SELELQGGIDELSSVAREMVRLETATVPF AVDVGHVINGWNAKIAELTGLSVEEAM
 GmphyB4 515 SELELQGGIDELSSVAREMVRLETATVPF AVDVGHVINGWNAKIAELTGLSVEEAM
 St phyB 608 SELELQGGIDELSSVAREMVRLETATVPF AVDVGHVINGWNAKIAELTGLSVEEAM
 Ps phyB 599 SELELQGGIDELSSVAREMVRLETATVPF AVDVGHVINGWNAKIAELTGLSVEEAM
 Vv phyB 607 SELELQGGIDELSSVAREMVRLETATVPF AVDVGHVINGWNAKIAELTGLSVEEAM

At phyB 696 GKSLVSDLVYKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 ZmphyB 695 GKSLVNDLIFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 Os phyB 705 GKSLVNDLIFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 Sb phyB 712 GKSLVNDLIFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 GmphyB1 673 GKSLVHDLVFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 GmphyB2 685 GKSLVHDLVFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 GmphyB3 636 GKSLVHDLVFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 GmphyB4 572 GKSLVHDLVFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 St phyB 665 GKSLVHDLVFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 Ps phyB 656 GKSLVHDLVFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD
 Vv phyB 664 GKSLVHDLVFKESAEATVKKLLSRALRGEEDKNVEIKLRTFGSEQNGKAVFVVVNACSSKD

At phyB 756 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 ZmphyB 755 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 Os phyB 765 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 Sb phyB 772 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 GmphyB1 733 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 GmphyB2 745 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 GmphyB3 696 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 GmphyB4 632 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 St phyB 725 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 Ps phyB 716 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT
 Vv phyB 724 YLNNIVGVCFVGQDVTGQKVMDKFINIQGDYKAVHSPNPLIPPFAASDNTCCLEWNT

At phyB 816 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 ZmphyB 815 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 Os phyB 825 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 Sb phyB 793 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 GmphyB1 832 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 GmphyB2 805 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 GmphyB3 756 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 GmphyB4 692 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 St phyB 785 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 Ps phyB 776 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD
 Vv phyB 784 AMEKL TGWSRGEVIGKMLVGEVFGSCCMLKGPDALTKFMIVLHNAIGGQDTRKFPFSFDD

FIG. 12C

At phyB	876	RNGKYEVOAL TANKRVSLGKVI GAFCF LQI PSPELQOALAVORRQDTE CFTKAKELAY I
ZmphyB	875	KNGKYVOAL TANTRSKMDGKS IGAFCF LQI ASPELQOAFE IQRQOQEK KCYARMKELAY I
Os phyB	885	KNGKYVOAL TANTRSRVDGEA IGAFCF LQI ASPELQOAFE IQRHHEK KCYARMKELAY I
Sb phyB	892	KNGKYVOAL TANTRSKMDGKS IGAFCF LQI ASAEIQOAFE IQRQOQEK KCYARMKELAY I
GmphyB1	853	RHGKYVOTFL TANKRVNMGQT IGAFCF LQI MSPELQOAL KAQRQOQEKNSF GRMKELAY I
GmphyB2	865	RYGKHVQAF L TANKRVNMDGQI GAFCF LQI VSPPELQOAL KAQRQOQEKNS FARMKELAY I
GmphyB3	816	RHGKYVOTFL TANKRVNMGQT IGAFCF LQI MSPELQOAL KAQRQOQEKNSF GRMKELAY I
GmphyB4	752	RYGKHVQAF L TANKRVNMDGQI GAFCF LQI VSPPELQOAL KAQRQOQEKNS FARMKELAY I
St phyB	845	RNGKYVOAL TANKRVNMGQT IGAFCF LQI ASPELQOAL RVORQOQEK KGYSQMKELAY I
Ps phyB	836	RHGKYVHTFL TANKRVNMDGQI GAFCF LQI VNPPELQOAL TVQRQOQES S LARMKELAY I
Vv phyB	844	QNGKYVOAL TANKRVNMGQT IGAFCF LQI ASPELQOAL KVQRQOQEK KCFARMKELAY I

At phyB	936	CQV IKNPLSGVRFANSLLEAT D LNEEDQKQL LETSVSCEKQISRIVGDV DLES IEDGSEFVL
ZmphyB	935	COE IKNPLSGIRETNSLLEAT D LNDDDRQQL LETSSACEKQMSKIVK DALS OS IEDGSEFVL
Os phyB	945	YQE IKNPLSGIRFTNSLLEAT D LKDDDRQQL LETSTAGEKQMSKIVK DALS OS IEDGSEFVL
Sb phyB	952	COE IKNPLSGIRFTNSLLEAT D LNDDDRQQL LETSSACEKQMSKIVK DALS OS IEDGSEFVL
GmphyB1	913	CQGVKNPLSGIRFTNSLLEAT S LITNEQKQF LETSVACEKQMLKI IRVDLES IEDGSEFEL
GmphyB2	925	CQGVKNPLSGIRFTNSLLEAT S LITNEQKQF LETSAACEKQMLKI IRVDLES IEDGSEFEL
GmphyB3	876	CQGVKNPLSGIRFTNSLLEAT S LITNEQKQF LETSVACEKQMLKI IRVDLES IEDGSEFEL
GmphyB4	812	CQGVKNPLSGIRFTNSLLEAT S LITNEQKQF LETSAACEKQMLKI IRVDLES IEDGSEFEL
St phyB	905	COE IKSPLNGIRFTNSLLEAT N LITENQKQF LETSAACEKQMSK I IRVDLEN IEDGSEFEL
Ps phyB	896	COE VKNPLSGIRFTNSLLEAT S LITDEQKQL LETSVACEKQMLKI IRVDLES IEDGSEFEL
Vv phyB	904	COE IKNPLSGIRFTNSLLEAT D LITEDQKQF LETSAACEKQMSK I IRVDLES IEDGSEFEL

At phyB	996	KREFFFLGS V INAVVSQAMFL LRDRGLQL IRDIPEEIKS IEVFGDQIR IQQLAEFLS M
ZmphyB	995	EKGEFSLG V INAVVSQAMFL LRDRDLQL IRDIPDEIKDASAYGDDQRIQQVLA DFLLSM
Os phyB	1005	EKGEFSLG V INAVVSQVMFL LRERDLQL IRDIPDEIKEASAYGDDQRIQQVLCDFLLSM
Sb phyB	1012	EKSEFSLG V INAVVSQAMFL LRDRDLQL IRDIPDEIKDASAYGDDQRIQQVLA DFLLSM
GmphyB1	973	EKGEFLLGNV INAVVSQVMLLRERNLQL IRDIPEEIKTAVYGDQIRIQQVLSDFLLNM
GmphyB2	985	EKGEFLLGNV INAVVSQVMLLRERNLQL IRDIPEEIKTAVYGDQIRIQQVLSDFLLNM
GmphyB3	936	EKGEFLLGNV INAVVSQVMLLRERNLQL IRDIPEEIKTAVYGDQIRIQQVLSDFLLNM
GmphyB4		
St phyB	965	EKREDFFLGS V INAVVSQVMLLRKGVQL IRDIPEEIKTAVHGDQIRIQQVLA DFLLSM
Ps phyB	956	EKGEFLLGNV INAVVSQVMLLRDRKLQL IRDIPEEIKAVYGDQIRIQQVLA DFLLSM
Vv phyB	964	EKAEFLLGS V INAVVSQVMFL LRERDLQL IRDIPEEIKTAVYGDQIRIQQVLA DFLLSM

At phyB	1056	I RYAPSQE WVE HLSQLSKQADGFAAIRTE FRMACPGEGLPPELVRDMFHSSRWTSPE
ZmphyB	1055	VRSAPSENGWVE I QVRPNVKONSDGTNTELF IFRFACPGEGLPADVVDQMFNSNSQWSTQE
Os phyB	1065	VRSAPAEENGWVE I QVRPNIKONSDGTDMLFLFRFACPGEGLPPELVQDMFNSNSRWTTQE
Sb phyB	1072	VRSAPSENGWVE I QVRPNVKONSDGTDTELF IFRFACPGEGLPADIVQDMFNSNSQWSTQE
GmphyB1	1033	VRYAPSPDGWVE I HVVPRIKQISDGLTLLHAEFRMVCPGEGLPPEL IQDMFNNSRWGTQE
GmphyB2	1045	VRYAPSPDGWVE I HVVPRIKQISDGLTLLHAEFRMVCPGEGLPPEL IQDMFNNSRWGTQE
GmphyB3	996	VRYAPSPDGWVE I HVVPRIKQISDGLTLLHAEFRMVCPGEGLPPEL IQDMFNNSRWGTQE
GmphyB4		
St phyB	1025	VRYAPSPDGWVE I QLRPSMMPISDGVTVGVHIELRIICPGEGLPPELVQDMFHSSRWVTQE
Ps phyB	1016	VRYAPSPDGWVE I HVVPRIKQISEGLTLLHAEFRMVCPGEGLPPEL IQDMFNNSRWVTQE
Vv phyB	1024	VRYAPSPDGWVE I QVRPCLKQISEEVKLMHTEFRMVCPGEGLPPEL IQDMFHSSRWMTQE

At phyB	1115	GLGLSVCRKILKLMNGEVQY IRESERSYF L ILELPVPRKRPLSTASGS GDMMLMMPY D
ZmphyB	1115	GVGLSTCRKILKLMNGEVQY IRESERSFF L VLEQPQPPAAGREIV D
Os phyB	1125	GLGLSVCRKILKLMNGEVQY IRESERSFF L H VLELPQPPQAASRGTS D
Sb phyB	1132	GVGLSTCRKILKLMNGEVQY IRESERSFF L VLELPQPPPAADREIS D
GmphyB1	1093	GLGLSMSRKILKLMNGEVQY IREAEERCYFVLLLELPVTRRSSKCC D
GmphyB2	1105	GLGLSMSRKILKLMNGEVQY IREAEERCYFVLLLELPVTRRSSKCC D
GmphyB3	1056	GLGLSMSRKILKLMNGEVQY IREAEERCYFVLLLELPVTRRSSKCC D
GmphyB4		
St phyB	1085	GLGLSVCRKILKLMNGEVQY IRESERCYF L VLDLPMTKGPKSVG D
Ps phyB	1076	GLGLSMSRKILKLMNGEVQY IREAEERCYFVLLLELPVTRRSSKCC D
Vv phyB	1084	GLGLSMCRKILKLMNGEVQY IRESERCYF L SLELPVTRRSSKCC D

At phyB = SEQ ID NO.1
 Zm phyB = SEQ ID NO.2
 Os phyB = SEQ ID NO.3
 Sb phyB = SEQ ID NO.4
 GmphyB1 = SEQ ID NO.5
 GmphyB2 = SEQ ID NO.6
 GmphyB3 = SEQ ID NO.7
 GmphyB4 = SEQ ID NO.8
 St phyB = SEQ ID NO.9
 Ps phyB = SEQ ID NO.10
 Vv phyB = SEQ ID NO.11

FIG. 12D

<i>At phyB_GAF</i>	234	SQKLAVRAISQLQALPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>ZmphyB_GAF</i>	232	SQKLAVRAISRLQALPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>Os phyB_GAF</i>	241	SQKLAVRAISRLQALPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>Sb phyB_GAF</i>	249	SQKLAVRAISRLQALPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>GmphyB1_GAF</i>	218	SQKLAVRAISQLQSLPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>GmphyB2_GAF</i>	230	SQKLAVRAISQLQSLPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>GmphyB3_GAF</i>	181	SQKLAVRAISQLQSLPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>GmphyB4_GAF</i>	117	SQKLAVRAISQLQSLPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>St phyB_GAF</i>	209	SQKLAVRAISMLQSLPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>Ps phyB_GAF</i>	197	SQKLAVRAISQLQALPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>Vv phyB_GAF</i>	208	SQKLAVRAISHLQSLPGGDVKKLCDTVVESVRDLTGYDRVMVYRFHEDEHGEVVAESRR
<i>Syn Cph1_GAF</i>	134	LGFYHMANAALNRLRQANLRDFYDVIVVEVRRMTGSDRVMVYRFDENNHGDVDAEDKR
<i>Dr BphP_GAF</i>	134	STGPHALRNAMFALASAPNLRALAEVATQTVRELTGSDRVMVYRFAPDATGVAEARR
<i>Po BphP_GAF</i>	121	T.SFTLNAQRIFAQVQLHNDTASL5MVTDEVRMTGSDRVMVYRFHSDSGEVVAESRR
<i>Rp BphP3_GAF</i>	143	NEFFRSRVVAIRRLQTAADLPTAIAAASEVRRITGSDRVMVYRFADDSGVVAESRR
<i>SyB Cph1_GAF</i>	13	.SRDALINRITHQIROSLELDQILRATVVEVRAFLGTDRVMVYRFDPFGHGTVAEARRGG

▼ 307

▼ 322

<i>At phyB_GAF</i>	293	DPLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...PVLVVDQDRITQSMCLVGS
<i>ZmphyB_GAF</i>	291	DPLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...PVRVVDQDRITQSMCLVGS
<i>Os phyB_GAF</i>	300	SNLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...PVRVVDQDRITQSMCLVGS
<i>Sb phyB_GAF</i>	308	DNLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...PVRVVDQDRITQSMCLVGS
<i>GmphyB1_GAF</i>	277	PDLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...AVRVVQDEALVQPLCLVGS
<i>GmphyB2_GAF</i>	289	PDLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...AVRVVQDEALVQPLCLVGS
<i>GmphyB3_GAF</i>	240	PDLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...AVRVVQDEALVQPLCLVGS
<i>GmphyB4_GAF</i>	176	PDLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...AVRVVQDEALVQPLCLVGS
<i>St phyB_GAF</i>	268	SDLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...PVRVVDQDRITQSMCLVGS
<i>Ps phyB_GAF</i>	256	VDLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...PVRVVDQDRITQSMCLVGS
<i>Vv phyB_GAF</i>	267	SDLEPYIGLHYPATDIPQASRFLKQNRVRMIVDCHAS...PVLVVDQDRITQSMCLVGS
<i>Syn Cph1_GAF</i>	193	DNLEPYIGLHYVESDIPQARRFLHMPTRVIVDVGVAV...LTPAVNPFSTRAMDITESTI
<i>Dr BphP_GAF</i>	193	EGLHAFLLHRFPASDIPQARRFLYTRHLRRTADTRAAAV...DPPILNPTNAPVPPGGAV
<i>Po BphP_GAF</i>	180	EGLHAFLLHRFPASDIPQARRFLYTRHLRRTADTRAAAV...DPPILNPTNAPVPPGGAV
<i>Rp BphP3_GAF</i>	202	SGTSSLDFRFPESDIPQARRFLYTRHLRRTADTRAAAV...DPPILNPTNAPVPPGGAV
<i>SyB Cph1_GAF</i>	72	ERTPSLLGRTFPAGDIPQARRFLRLAQVRVIVDVEAQSRS...SQPESWGLSARMPGEPL

▼ 352

▼ 361

<i>At phyB_GAF</i>	351	RAPHGCHSQYMANMGSIASLVMAYINGN...EDDGSNVASG...SSMRLWGLVVCCHHTSS
<i>ZmphyB_GAF</i>	349	RAPHGCHSQYMANMGSIASLVMAYINGN...SSG.GDDEQTGRGG...ISSAMRLWGLVVCCHHTSS
<i>Os phyB_GAF</i>	358	RNPHGCHSQYMANMGSIASLVMAYINGN...SSGGDDHNTARGG...IPSAAMRLWGLVVCCHHTSS
<i>Sb phyB_GAF</i>	366	RAPHGCHSQYMANMGSIASLVMAYINGN...SSG.GDDEQTGRGG...ISSAMRLWGLVVCCHHTSS
<i>GmphyB1_GAF</i>	335	RAPHGCHSQYMANMGSIASLVMAYINGN...DEEGVGG...RSMRLWGLVVCCHHTSA
<i>GmphyB2_GAF</i>	347	RAPHGCHSQYMANMGSIASLVMAYINGN...DEEGVGG...RSMRLWGLVVCCHHTSA
<i>GmphyB3_GAF</i>	298	RAPHGCHSQYMANMGSIASLVMAYINGN...DEEGVGG...RSMRLWGLVVCCHHTSA
<i>GmphyB4_GAF</i>	234	RAPHGCHSQYMANMGSIASLVMAYINGN...DEEGVGG...RSMRLWGLVVCCHHTSA
<i>St phyB_GAF</i>	326	RAPHGCHSQYMANMGSIASLVMAYINGN...DEEGVGG...RSMRLWGLVVCCHHTSV
<i>Ps phyB_GAF</i>	314	RAPHGCHSQYMANMGSIASLVMAYINGN...DEEGVGG...RSMRLWGLVVCCHHTSA
<i>Vv phyB_GAF</i>	325	RAPHGCHSQYMANMGSIASLVMAYINGN...DEEGVGG...RSMRLWGLVVCCHHTSA
<i>Syn Cph1_GAF</i>	253	RSAYHCHLTVLNMGVGLISLTKDGH...L...MGLTACHHOTP
<i>Dr BphP_GAF</i>	253	RATSPMHMQLRNMGVGLISLTVVGGQ...L...MGLTACHHOTP
<i>Po BphP_GAF</i>	240	RSVSPHCEVLTNMGVGLISLTVVGGQ...L...MGLTACHHOTP
<i>Rp BphP3_GAF</i>	262	RSVSPHCEVLTNMGVGLISLTVVGGQ...L...MGLTACHHOTP
<i>SyB Cph1_GAF</i>	132	QRVPDPCHVHLKSMGVGLISLTVVGGQ...L...MGLTACHHOTP

<i>At phyB_GAF</i>	408	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.12
<i>ZmphyB_GAF</i>	407	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.13
<i>Os phyB_GAF</i>	417	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.14
<i>Sb phyB_GAF</i>	424	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.15
<i>GmphyB1_GAF</i>	389	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.16
<i>GmphyB2_GAF</i>	401	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.17
<i>GmphyB3_GAF</i>	352	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.18
<i>GmphyB4_GAF</i>	288	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.19
<i>St phyB_GAF</i>	381	RSIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.20
<i>Ps phyB_GAF</i>	372	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.21
<i>Vv phyB_GAF</i>	379	RCIPFPFLRYACEFLMQAFGLQLNMEI	SEQ ID NO.22
<i>Syn Cph1_GAF</i>	295	KVLEHEIRKACEFFGRVVFNSISAQE	SEQ ID NO.34
<i>Dr BphP_GAF</i>	295	YVLEPDARTLISYIGRLLSIQVQVKE	SEQ ID NO.35
<i>Po BphP_GAF</i>	282	XLITVIVRMSFQIFSOVCSAIVERLE	SEQ ID NO.36
<i>Rp BphP3_GAF</i>	304	RFVSYEVROACQLIAQVLTWQIGVLE	SEQ ID NO.37
<i>SyB Cph1_GAF</i>	174	RPYSQEELQVVQLQADQVSIATQAE	SEQ ID NO.38

FIGURE 13

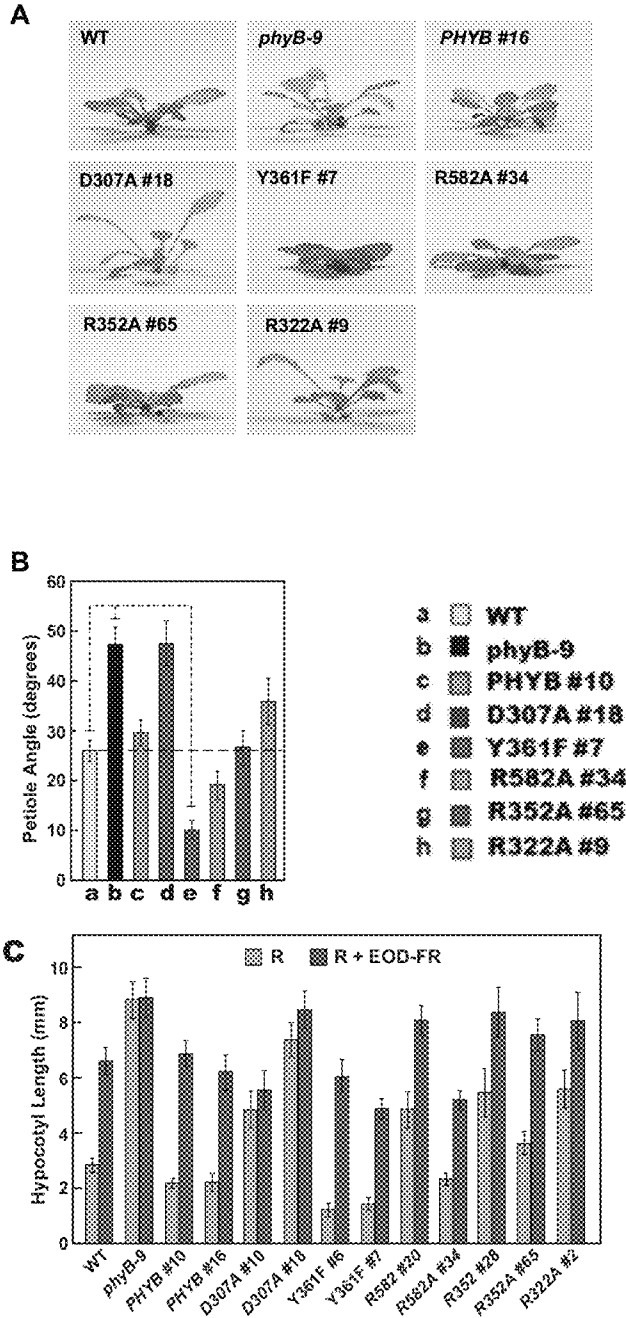


FIGURE 14

PLANTS WITH ALTERED PHYTOCHROMES

GOVERNMENT SUPPORT

This invention was made with government support under 07191530 awarded by the National Science Foundation and 13-CRHF-0-6055 awarded by the USDA/NIFA. The government has certain rights in the invention.

INCORPORATION OF SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted in ASCII format via EFS-Web and is hereby incorporated by reference in its entirety. Said ASCII copy, created on Oct. 1, 2013, is named ASFILED_Substitute SequenceListing-Text.txt and is 188,936 bytes in size.

BACKGROUND

The rise of the global population and demands for carbon-neutral biofuels have accelerated the need to improve agricultural productivity. An emerging strategy is to control plant reproduction and architecture to better fit specific environments and to increase crop densities. Increasing crop densities may be achieved by producing plants that perform well in more competitive environments. Plant architecture, timing of reproduction, and plant responses to competition may be manipulated to produce plants adapted to growing in crowded conditions.

Phytochromes encompass a diverse collection of biliproteins that enable cellular light perception by photoconverting between a red-light (R)-absorbing ground state—Pr and a far-red light (FR)-absorbing active state—Pfr. In *Arabidopsis thaliana* there are five phytochromes, designated phytochrome A (phyA) to phytochrome E (phyE). Phytochrome B (phyB) is the predominant phytochrome regulating de-etiolation responses in R light and shade avoidance. Phytochromes are synthesized in the cytosol as an inactive Pr form, and are converted to the biologically active Pfr form by light irradiation which then is translocated into the nucleus. Phytochromes play fundamental roles in photoperception by a plant and adaptation of its growth to the ambient light environment.

SUMMARY

An isolated polynucleotide comprising a contiguous coding sequence encoding a polypeptide comprising an amino acid sequence having at least 80% identity to at least one sequence selected from SEQ ID NOs: 1-22 and containing an amino acid other than tyrosine at the position corresponding to Y361 of SEQ ID NO. 1, and plants and plant cells containing such polynucleotides are provided. In certain embodiments, a plant comprising the isolated polynucleotide exhibits increased expression of the polypeptide, relative to a control plant, and, relative to the control plant, may exhibit increased light sensitivity, decreased height, decreased diameter, decreased petiole length, decreased internode length, decreased stem diameter, decreased hypocotyl length under an R (red light) fluence rate of less than $1 \mu\text{mole m}^{-2} \text{sec}^{-1}$, modified hyponasty, or enhanced germination. In certain embodiments, increased light sensitivity results in a smaller plant adapted to provide an increased yield in shaded or competitive conditions.

In another embodiment the invention provides a method of producing a transgenic plant by introducing into a plant cell a polynucleotide encoding a polypeptide comprising an

amino acid sequence having at least 80% identity to at least one amino acid sequence selected from SEQ ID NOs: 1-22 and having an amino acid other than tyrosine at the position corresponding to Y361 of SEQ ID NO:1, and regenerating the transformed cell to produce a transgenic plant.

In another embodiment, an isolated polypeptide comprising an amino acid sequence having at least 80% identity to at least one amino acid sequence selected from SEQ ID NOs: 1-22, and having an amino acid other than tyrosine at the position corresponding to Y361 of SEQ ID NO:1 is provided.

In another embodiment, an isolated polynucleotide is provided which comprises a contiguous coding sequence encoding a polypeptide having at least 80% identity to at least one amino acid sequence selected from SEQ ID NOs: 1-22 and which has at least one different amino acid at a select position. The different amino acid may be (i) an amino acid other than aspartate (D) at the position corresponding to 307 of SEQ ID NO:1, (ii) an amino acid other than arginine (R) at the position corresponding to 322 of SEQ ID NO: 1, (iii) an amino acid other than arginine (R) at the position corresponding to 352 of SEQ ID NO: 1, (iv) an amino acid other than arginine (R) at the position corresponding to 582 of SEQ ID NO: 1, or a combination thereof.

Other aspects of the invention will become apparent by consideration of the detailed description and accompanying drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1. Drawings depicting the scheme of phy action and the three-dimensional relationships of key amino acids within the bilin-binding photosensory module (PSM). (A) Scheme depicting the main steps involved in phy assembly, Pr/Pfr photointerconversion, stability, and action in higher plants. (B) Top (upper) and side (lower) three-dimensional views of the PSM from Syn-Cph1 (PDB code 2VEA [3]) assembled with phycocyanobilin (PCB) highlighting the positions of key conserved amino acids surrounding the bilin and the cysteine involved in bilin attachment (C259). The residue numbers are those for the homolog Syn-Cph1 from the cyanobacterium *Synechocystis* PCC6803. The GAF domain and PHY hairpin are colored in green and orange, respectively. PCB is colored in cyan with the individual pyrrole rings labeled. Sulfur, oxygen and nitrogen atoms are colored yellow, red, and deep blue, respectively. Important contacts are indicated by dashed lines. pw, pyrrole water. (C) Alignment of the GAF domain protein sequences among bacterial phys with available structures with those from the phyB-E family in *Arabidopsis*. Residues pertinent to this study are indicated by red arrowhead; their sequence positions are shown either above for Syn-Cph1 or below the alignment for *A. thaliana* phyA and phyB. At, *Arabidopsis thaliana*; Dr, *Deinococcus radiodurans*; Pa, *Pseudomonas aeruginosa*; Rp, *Rhodospseudomonas palustris*; SyB, *Synechococcus* OS-B¹; Syn, *Synechocystis* PCC6803

FIG. 2. Graphs depicting spectral properties, photochemistry, and thermal reversion rates of wild-type and mutant versions of *Arabidopsis* phyB. The PSM (PAS-GAF-PHY) of each phyB protein was synthesized recombinantly with a C-terminal 6His tag, assembled with PΦB in vivo, and purified. See FIG. 7 for SDS-PAGE analysis. (A) UV-vis absorption spectra of Pr (solid lines) or following its excitation with saturating R (dashed lines). Difference maxima and minima (Pr minus R) are indicated. (B) Rates of Pr→Pfr photoconversion (left), Pfr→Pr photoconversion (middle), and thermal reversion of Pfr back to Pr (right). All rates were

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expressed as the percent of Pfr in the sample using the absorption maximum of Pfr near 725 nm for quantification.

FIG. 3. Photographs showing the phenotypes of an *Arabidopsis* phyB null mutant rescued with transgenes expressing wild-type or mutant versions of full-length phyB. Shown are wild type (WT), the phyB-9 null mutant, and two independent transgenic lines expressing either the wild-type or mutant PHYB cDNAs in the phyB-9 background. (A) Representative 3-week-old plants grown in long days (LDs). (B) Representative 4-d-old seedlings either grown in the dark (D) or under continuous 13 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ R. (C) Levels of the phyB protein in each of the lines examined in panels A and B as determined by immunoblot analysis of crude extracts from 5-d-old dark grown seedlings with an anti-phyB monoclonal antibody. Near equal protein loading was confirmed with anti-PBA1 antibodies.

FIG. 4. Graphs depicting R sensitivity of hypocotyl elongation for a phyB null mutant rescued with transgenes expressing wild-type or mutant versions of full-length phyB. Shown are wild type (WT), the phyB-9 null mutant, and two independent transgenic lines expressing either the wild-type or mutant PHYB cDNAs in the phyB-9 background. Hypocotyl length of each line was expressed relative to that measured for dark-grown seedlings. Each data point represents the mean (\pm SE) from four independent experiments. (A) Sensitivity to a broad range of R fluence rates. (B) Sensitivity of the Y361F variant to very low fluence rates of R.

FIG. 5. Graphs depicting sensitivity of phyB-9 plants rescued with various phyB mutants to a collection of photomorphogenic processes controlled by phyB. Shown are wild type (WT), the phyB-9 null mutant, and two independent transgenic lines expressing either the wild-type or mutant PHYB cDNAs in the phyB-9 background. (A) Germination efficiency of seeds either treated with 2-hr pulse of WL (white light) alone or followed by a pulse of FR (far-red light). Germination was assessed after a subsequent 5-d incubation in darkness. Each bar represents the average (\pm SE) of 5 experiments involving at least 40 seeds each. (B) EOD-FR effect on hypocotyl growth. Etiolated seedlings were subject over 4 d to a light regime of continuous R (90 $\mu\text{mole}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$) for 8 hr, followed by either darkness or a 10-min pulse of FR (R+EODFR, 100 $\mu\text{mole}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$), and then 16-hr of darkness. Each bar represents the average (\pm SE) of 4 experiments involving at least 15 seedlings each. The two phyB^{Y361F} lines were significantly more sensitive to R and line #7 was more derepressed by R+EOD-FR than the two phyB^{WT} lines (*, Student's t test: $P < 0.05$). (C) Flowering time in short days (SDs) (8-hr light/16-hr dark). Each bar represents the average number of leaves generated before emergence of the inflorescence stem for >20 plants (\pm SE).

FIG. 6. Photographs showing the effect of the phyB mutations on the nuclear distribution and R-induced degradation of the photoreceptor. (A) Loss of phyB protein during continuous R irradiation of etiolated seedlings. phyB levels in 4-d-old dark-adapted *Arabidopsis* were measured after various length exposures to 90 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ R by immunoblot analysis using an anti-phyB monoclonal antibody. The seedlings were exposed to 100 μM MG132 or an equivalent volume of DMSO 12 hr before irradiation. Near equal protein loading was confirmed with anti-histone H3 antibodies. (B) Subcellular partitioning of wild-type and mutant phyB in continuous R. Wild-type phyB and the various mutants were expressed as fusions to the N-terminus of YFP in the phyB-9 background. Regions surrounding the nucleus were imaged by fluorescence confocal microscopy from hypocotyl cells either kept in the dark or irradiated for 12 hr

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with continuous 90 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ R. Scale bar represents 20 μm . Expression levels of the fusions and their ability to rescue the phyB-9 phenotype with respect to hypocotyl elongation in R can be found in FIG. 8.

FIG. 7. Photographs of protein gels showing the purification and assembly of the PSM from wild-type phyB and the various site-directed mutants. The 6His-tagged polypeptides were co-expressed recombinantly with the dual enzyme system that synthesizes the higher plant phy chromophore phytochromobilin (P Φ B). The purified chromoproteins were subjected to SDS-PAGE and either stained for protein with Coomassie blue or for the bound bilin by zinc-induced fluorescence (Zn).

FIG. 8. Photographs (A) and graphs (B) showing the accumulation of YFP fusions of phyB in *Arabidopsis* seedlings. Full-length wild-type phyB or the various mutants (R322A, D307A, Y361F, and R582A) were expressed in the phyB-9 background as fusions to the N-terminus of YFP. Total crude extracts from 5-d-old dark grown seedlings were subjected to immunoblot analysis using a monoclonal antibody against either phyB or GFP (Sigma). Near equal loading was confirmed with anti-PBA1 or anti-histone H3 antibodies. The arrowheads locate phyB and the phyB-YFP fusions.

FIG. 9. Photographs showing the morphology of phyB null mutant *Arabidopsis* rescued with transgenes expressing wild-type or mutant versions of full length phyB. Shown are wild type (WT), the phyB-9 null mutant, and representative transgenic lines expressing either the wild-type or mutant PHYB cDNAs in the phyB-9 background grown under SD (short day) until bolting.

FIG. 10. Photographs showing the phenotype of an *Arabidopsis* phyA null mutant rescued with transgenes expressing wild-type or mutant versions of full-length phyA. Shown are wild-type (WT), the phyA-211 null mutant, and two independent transgenic lines expressing either the wild-type or mutant PHYA cDNAs in the phyA-211 background. (A) Representative 3-week-old plants grown under LD. (B) Representative 4-d-old seedlings either grown in the dark (D) or under continuous 5 $\mu\text{mole}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ FR. (C) Levels of the phyA protein in each of the lines examined in panels A and B as determined by immunoblot analysis of crude extracts from 5-d-old dark grown seedlings with an anti-phyA monoclonal antibody. Near equal protein loading was confirmed with anti-PBA1 antibodies.

FIG. 11. Graphs depicting FR sensitivity of hypocotyl elongation for a phyA null mutant rescued with transgenes expressing wild-type or mutant versions of full-length phyA. Shown are wild type (WT), the phyA-211 null mutant, and two independent transgenic lines expressing either the wild-type or mutant PHYA cDNAs in the phyA-211 background. Hypocotyl length of each line was expressed relative to that measured for dark-grown seedlings. Each data point represents the mean (\pm SE) from four independent experiments.

FIG. 12A, FIG. 12B, FIG. 12C, and FIG. 12D. Chart depicting the alignment of the full-length polypeptide sequences of phyB from *Arabidopsis* and crop species. Residues corresponding to Tyr361 in *Arabidopsis* phyB are indicated by the arrowhead at 361. At, *Arabidopsis thaliana*; Zm, *Zea mays*; Os, *Oryza sativa*; Sb, *Sorghum bicolor*, Gm, *Glycine max*; St, *Solanum tuberosum* L.; Ps, *Pisum sativum*; Vv, *Vitis vinifera*. The protein sequences were obtained from National Center for Biotechnology Information except ZmphyB sequence which was from the Phytozome resource. Alignment was performed using ClustalW (*Nucleic Acids Res.* 22 (22); 4673-80).

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FIG. 13. Chart depicting the alignment of the polypeptide sequences of GAF domains from microbial phys with available structures and phyB among *Arabidopsis* and crop species. Residues corresponding to Asp307, Arg322, Arg352, and Tyr361 in *Arabidopsis* phyB are indicated by the arrowhead. At, *Arabidopsis thaliana*; Zm, *Zea mays*; Os, *Oryza sativa*; Sb, *Sorghum bicolor*; Gm, *Glycine max*; St, *Solanum tuberosum* L.; Ps, *Pisum sativum*; Vv, *Vitis vinifera*; Dr, *Deinococcus radiodurans*; Pa, *Pseudomonas aeruginosa*; Rp, *Rhodospseudomonas palustris*; SyB, *Synechococcus* OS-B'; Syn, *Synechocystis* PCC6803. The protein sequences were obtained from National Center for Biotechnology Information except ZmphyB sequence which was from the Phytozome resource. Alignment was performed using ClustalW (*Nucleic Acids Res.* 22 (22); 4673-80).

FIG. 14. Photograph and graphs depicting the sensitivity of phyB-9 plants rescued with various phyB mutants to photomorphogenic processes controlled by phyB. Shown are wild type (WT), the phyB-9 null mutant, and one or two independent transgenic lines expressing either the wild-type or mutant PHYB cDNAs in the phyB-9 background. See FIG. 3D for the description of the mutant lines. (A), Photograph showing the side view of 45-d-old seedlings grown in white light under SD illustrating the influence of phyB on leaf epinasty. (B), Quantification of leaf epinasty for seedlings in panel A. Each bar represents the average angle between the soil surface and the petiole for the 4th and 5th leaves of 10 plants (20 total angles). The 95% confidence interval for each average is shown. The values for WT, phyB-9, and Y361F lines are significantly different from each other by Student's t test ($p < 0.05$). (C), Effect of R and EOD-FR on hypocotyl growth. Etiolated seedlings were subjected over 4 d to a 24-hr light regime of continuous R ($90 \mu\text{mole m}^{-2} \text{s}^{-1}$) for 8 hr, followed by either darkness (R) or a 10-min pulse of $100 \mu\text{mole m}^{-2} \text{s}^{-1}$ FR(R+EOD-FR) and then 16-hr of darkness. Each bar represents the average (\pm SE) of 4 experiments involving at least 15 seedlings each. The Y361F #7 line was significantly different from WT and PHYB for both R and R+EOD-FR by Student's t-test ($p < 0.05$).

DETAILED DESCRIPTION

The present disclosure relates to polynucleotides and polypeptides and use of the polynucleotides and polypeptides for modifying the phenotypes of plants or plant cells. Modified plants or plant cells comprising the polynucleotides and/or polypeptides are also provided. In certain embodiments, the modified plants or plant cells exhibit one or more of an altered light sensitivity, an improved or enhanced germination efficiency of seeds, such as in low light, a hypersensitivity to white and red light with respect to hypocotyl and stem growth, improved shade tolerance, and a smaller plant size.

The polypeptides discussed herein are phytochromes and show homology to certain phytochrome sequences from *Arabidopsis thaliana*. The term "phytochrome" is used generically to refer to a phytochrome from any plant species. Plant phytochromes include phyA, phyB, phyC, phyD and phyE.

Phytochrome domains from a variety of organisms may be used as starting points for modifications that will generate the modified phytochromes of the present invention, and isolated polynucleotides encoding the modified phy domains. In certain embodiments the phytochrome is a modified phyB plant phytochrome, or a modified cGMP

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phosphodiesterase/adenylyl cyclase/FhIA (GAF) domain or modified chromophore binding domain (CBD) of phyB. Modification of phytochromes and/or phytochrome domains can be performed by methods known in the art, e.g., site-directed mutations, additions, deletions, and/or substitutions of one or more amino acid residues of existing phytochromes and/or phytochrome domains. Alternatively, modified phytochromes and/or phytochrome domains can be synthesized de novo, for example by synthesis of novel genes that would encode phytochrome domains with desired modifications.

In certain embodiments, expression in plants of a modified phytochrome having an amino acid sequence with at least 80%, or at least 95% identity to at least one of SEQ ID NOs: 1-22 and having an amino acid other than tyrosine at the position corresponding to Y361 of SEQ ID NO: 1 (for example, by introducing a polynucleotide sequence having at least 95% identity to (i) a sequence selected from SEQ ID NOs 23-33 into the plant, or (ii) a GAF-encoding domain of a sequence selected from SEQ ID NOs: 23-33, and encoding an amino acid other than tyrosine at the position corresponding to Y361 of SEQ ID NO: 1) results in plants that have altered light sensitivity, including, but not limited to, an improved germination efficiency of seeds in low light, a hypersensitivity to white and red light with respect to hypocotyl and stem growth, improved shade tolerance, reduced leaf surface area and combinations thereof, relative to control plants that do not express the modified phytochrome. The shared sequence identity of the nucleotides encoding phyB from a variety of species is shown in Table 1.

TABLE 1

Percent Identities of phyB from a variety of species with <i>Arabidopsis</i> phyB			
Species phyB	Accession Number	Nucleotide SEQ ID NO:	Percent Identity to SEQ ID NO: 23
<i>Arabidopsis</i>	NM_127435	SEQ ID NO: 23	100%
<i>Zea mays</i> (maize)	GRMZM2G124532	SEQ ID NO: 24	70.2%
<i>Oryza sativa</i> (rice)	JN594210	SEQ ID NO: 25	70.3%
<i>Sorghum bicolor</i> (<i>sorghum</i>)	Y466089	SEQ ID NO: 26	69.6%
<i>Glycine max</i> (soybean) phyB1	EU428749	SEQ ID NO: 27	73.1%
<i>G. max</i> phyB2	EU428750	SEQ ID NO: 28	72.6%
<i>G. max</i> phyB3	EU428751	SEQ ID NO: 29	72.5%
<i>G. max</i> phyB4	EU428752	SEQ ID NO: 30	58.6%
<i>Solanum tuberosum</i> L. (potato)	DQ342235	SEQ ID NO: 31	75.1%
<i>Pisum sativum</i> (pea)	AF069305	SEQ ID NO: 32	70.8%
<i>Vitis vinifera</i> (grape):	EU436650	SEQ ID NO: 33	76.1%

The terms "isolated," "purified," or "biologically pure" refer to material that is substantially or essentially free from components that normally accompany it as found in its native state. Purity and homogeneity are typically determined using analytical chemistry techniques such as polyacrylamide gel electrophoresis or high performance liquid chromatography. A protein that is the predominant species present in a preparation is substantially purified. In particular, an isolated nucleic acid of the present invention is separated from open reading frames that flank the desired gene and encode proteins other than the desired protein. The term "purified" denotes that a nucleic acid or protein gives rise to essentially one band in an electrophoretic gel. Par-

ticularly, it means that the nucleic acid or protein is at least 85% pure, more preferably at least 95% pure, and most preferably at least 99% pure.

Two nucleic acid sequences or polypeptides are said to be “identical” if the sequence of nucleotides or amino acid residues, respectively, in the two sequences are the same when aligned for maximum correspondence as described below. The term “complementary to” is used herein to mean that the sequence is complementary to all or a portion of a reference polynucleotide sequence. In the case of both expression of transgenes and inhibition of endogenous genes (e.g., by antisense or sense suppression) the inserted polynucleotide sequence need not be identical and may be “substantially identical” to a sequence of the gene from which it was derived.

In the case of polynucleotides used to inhibit expression of an endogenous gene, the introduced sequence need not be perfectly identical to a sequence of the target endogenous gene. The introduced polynucleotide sequence will typically be at least substantially identical (as determined below) to the target endogenous sequence.

In the case where the inserted polynucleotide sequence is transcribed and translated to produce a functional polypeptide, because of codon degeneracy, a number of polynucleotide sequences will encode the same polypeptide. These variants are specifically covered by the term “polynucleotide sequence from” a particular gene. In addition, the term specifically includes sequences (e.g., full length sequences) that are substantially identical (determined as described below) with a gene sequence encoding a polypeptide of the present invention and that encode polypeptides or functional polypeptide fragments that retain the function of a polypeptide of the present invention, e.g., a modified bacterial phytochrome with increased fluorescence.

Optimal alignment of sequences for comparison may be conducted by methods commonly known in the art, for example by the search for similarity method described by Pearson and Lipman 1988, Proc. Natl. Acad. Sci. USA 85: 2444-2448, by computerized implementations of algorithms such as GAP, BESTFIT, BLAST®, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), Madison, Wis., or by inspection. In a preferred embodiment, protein and nucleic acid sequence identities are evaluated using the Basic Local Alignment Search Tool (“BLAST®”), which is well known in the art (Karlin and Altschul, 1990, Proc. Natl. Acad. Sci. USA 87: 2267-2268; Altschul et al., 1997, Nucl. Acids Res. 25: 3389-3402), the disclosures of which are incorporated by reference in their entirety. The BLAST® programs identify homologous sequences by identifying similar segments, which are referred to herein as “high-scoring segment pairs,” between a query amino or nucleic acid sequence and a test sequence which is preferably obtained from a protein or nucleic acid sequence database. Preferably, the statistical significance of a high-scoring segment pair is evaluated using the statistical significance formula (Karlin and Altschul, 1990). The BLAST® programs can be used with the default parameters or with modified parameters provided by the user.

“Percentage of sequence identity” is determined by comparing two optimally aligned sequences over a comparison window, where the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical

nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

The term “substantial identity” of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 25% sequence identity compared to a reference sequence as determined using the programs described herein; preferably BLAST® using standard parameters, as described. Alternatively, percent identity can be any integer from 25% to 100%. More preferred embodiments include polynucleotide sequences that have at least about: 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% sequence identity compared to a reference sequence. These values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like. Accordingly, polynucleotides of the present invention encoding a protein of the present invention include nucleic acid sequences that have substantial identity to the nucleic acid sequences that encode the polypeptides of the present invention. Polynucleotides encoding a polypeptide comprising an amino acid sequence that has at least about: 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity compared to a reference polypeptide sequence are also preferred.

The term “substantial identity” of amino acid sequences (and of polypeptides having these amino acid sequences) normally means sequence identity of at least 40% compared to a reference sequence as determined using the programs described herein; preferably BLAST® using standard parameters, as described. Preferred percent identity of amino acids can be any integer from 40% to 100%. More preferred embodiments include amino acid sequences that have at least about: 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity compared to a reference sequence. Polypeptides that are “substantially identical” share amino acid sequences as noted above except that residue positions which are not identical may differ by conservative amino acid changes. Conservative amino acid substitutions refer to the interchangeability of residues having similar side chains. For example, a group of amino acids having aliphatic side chains is glycine, alanine, valine, leucine, and isoleucine; a group of amino acids having aliphatic-hydroxyl side chains is serine and threonine; a group of amino acids having amide-containing side chains is asparagine and glutamine; a group of amino acids having aromatic side chains is phenylalanine, tyrosine, and tryptophan; a group of amino acids having basic side chains is lysine, arginine, and histidine; and a group of amino acids having sulfur-containing side chains is cysteine and methionine. Preferred conservative amino acid substitution groups are: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, aspartic acid-glutamic acid, and asparagine-glutamine. Accordingly, polypeptides or proteins of the present invention include amino acid sequences that have substantial identity to the amino acid sequences of the polypeptides of the present invention, which are modified phytochromes that result in plants having altered sensitivity compared with plants.

In one embodiment, a modified phyB phytochrome is created by substituting the tyrosine at the position corresponding to Tyr361 of SEQ ID NO:1 with a phenylalanine (Phe). It is contemplated that various modifications of this Tyr361 residue (and its functional equivalents in other phytochromes) will result in phytochromes useful for practicing the present invention. Some examples of modified phytochromes useful for the practice of this invention include modifications of the tyrosine domain corresponding to Y361 of SEQ ID NO: 1, for example: Tyr to Phe (i.e., Y to F); Tyr to Trp (i.e., Y to W); Tyr to Ile (i.e., Y to I); Tyr to Leu (i.e., Y to L); Tyr to Val (i.e., Y to V); Tyr to Ala (i.e., Y to A); Tyr to Pro (i.e., Y to P); Tyr to Asn (i.e., Y to N); Tyr to Glu (i.e., Y to E); and Tyr to Thr (i.e., Y to T), Tyr to Gly (i.e., Y to G); Tyr to Ser (i.e., Y to S); Tyr to Cys (i.e., Y to C); Tyr to Lys (i.e., Y to K); Tyr to Arg (i.e., Y to R); Tyr to His (i.e., Y to H); Tyr to Met (i.e., Y to M); Tyr to Asp (i.e., Y to D); or Tyr to Gln (i.e., Y to Q).

The modified phyB phytochrome may contain amino acid substitutions described herein or known in the art at other locations in the phyB or phyB domain. In certain embodiments, the modified phytochrome may contain an amino acid substitution at the residue corresponding to D307, R322, R352, or R582 of the *Arabidopsis* phytochrome shown in SEQ ID NO: 1. For example, the modified phytochrome may contain at least one of (i) an amino acid other than aspartate (D) at the position corresponding to 307 of SEQ ID NO:1, (ii) an amino acid other than arginine (R) at the position corresponding to 322 of SEQ ID NO: 1, (iii) an amino acid other than arginine (R) at the position corresponding to 352 of SEQ ID NO: 1, and (iv) an amino acid other than arginine (R) at the position corresponding to 582 of SEQ ID NO: 1, or any combination thereof. These substitutions may be present alone, in any combination, including one or more substitutions in addition to a substitution at the position corresponding to Y361 of SEQ ID NO:1. The substitutions may include one or more of the following R352A, R582A, R322A and D307A.

The substitutions at R352 and R582, such as R352A, R582A produce a phytochrome phenotype that is slightly hyperactive with respect to signalling. For example, the substitution at R582 shows slightly stronger repression on hypocotyl growth at intermediate R fluence rates as compared to wild-type. The substitution at R322, such as R322A, produces a photochrome phenotype that is slightly hypoactive with respect to signalling.

As shown in the sequence alignment of FIG. 12, Tyr361 is conserved in plant phytochromes. Sequence identity of phyB from crop species compared with *Arabidopsis* phyB (SEQ ID NO: 1) is as follows: *Zea mays* (maize; SEQ ID NO: 2): 70.2%; *Oryza sativa* (rice; SEQ ID NO: 3): 70.3%; *Sorghum bicolor* (*sorghum*; SEQ ID NO: 4): 69.6%; *Glycine max* (soybean, phyB1 (SEQ ID NO: 5): 73.1%; phyB2 (SEQ ID NO: 6): 72.6%; phyB3 (SEQ ID NO: 7): 72.5%; phyB4 (SEQ ID NO: 8): 58.6%; *Solanum tuberosum* L. (potato; SEQ ID NO:9): 75.1%; *Pisum sativum* (pea; SEQ ID NO: 10): 70.8%; *Vitis vinifera* (grape; SEQ ID NO: 11): 76.1%.

As shown in FIG. 12, Tyr361 of SEQ ID NO: 1 (*Arabidopsis* phyB) corresponds to Tyr359 of SEQ ID NO: 2 (maize phyB), Tyr368 of SEQ ID NO: 3 (rice phyB), Tyr376 of SEQ ID NO: 4 (*sorghum* phyB), Tyr345 of SEQ ID NO: 5 (soybean phyB1), Tyr357 of SEQ ID NO: 6 (soybean phyB2), Tyr308 of SEQ ID NO: 7 (soybean phyB3), Tyr244 of SEQ ID NO: 8 (soybean phyB4), Tyr336 of SEQ ID NO: 9 (potato phyB), Tyr324 of SEQ ID NO: 10 (pea phyB), and Tyr335 of SEQ ID NO: 11 (grape phyB).

Tyr361 of SEQ ID NO: 1 (*Arabidopsis* phyB) also corresponds to Tyr 263 of the cyanobacteriophytochrome from *Synechocystis* PCC6803, to Tyr 263 of the bacteriophytochrome from *Deinococcus radiodurans*, to Tyr 250 of the bacteriophytochrome from *Pseudomonas aeruginosa*, to Tyr272 of the bacteriophytochrome from *Rhodospseudomonas palustris*, and to Tyr142 of the cyanobacteriophytochrome from *Synechococcus* OS-B'.

As shown in FIG. 13, the percent identity of the GAF domains to *Arabidopsis* phyB GAF domain (position 234 to 433 of SEQ ID NO: 1; SEQ ID NO: 12) are as follows: *Zea mays* (maize; position 232-432 of SEQ ID NO: 2; SEQ ID NO: 13): 82.1%; *Oryza sativa* (rice; position 241 to 442 of SEQ ID NO: 3; SEQ ID NO: 14): 82.7%; *Sorghum bicolor* (*sorghum*; position 249 to 449 of SEQ ID NO: 4; SEQ ID NO: 15): 82.1%; *Glycine max* (soybean), phyB1 (position 218 to 414 of SEQ ID NO: 5; SEQ ID NO: 16): 87.5%; phyB2 (position 230 to 426 of SEQ ID NO: 6; SEQ ID NO: 17): 85.5%; phyB3 (position 181 to 377 of SEQ ID NO: 18): 87.5%; phyB4 (position 117 to 313 of SEQ ID NO: 8; SEQ ID NO: 19): 85.5%; *Solanum tuberosum* L. (potato; position 209 to 406 of SEQ ID NO:9; SEQ ID NO: 20): 87.0%; *Pisum sativum* (pea; position 197 to 397 of SEQ ID NO: 10; SEQ ID NO: 21): 88.1%; *Vitis vinifera* (grape; position 208 to 404 of SEQ ID NO: 11; SEQ ID NO: 22): 85.0%.

As shown in FIG. 13, Tyr361 of SEQ ID NO: 1 (*Arabidopsis* phyB) corresponds to Tyr128 of SEQ ID NO: 12 (*Arabidopsis* phyB GAF domain); Tyr128 of SEQ ID NO: 13 (maize phyB GAF domain), Tyr128 of SEQ ID NO: 14 (rice phyB GAF domain), Tyr128 of SEQ ID NO: 16 (soybean phyB1 GAF domain), Tyr128 of SEQ ID NO: 17 (soybean phyB2 GAF domain), Tyr128 of SEQ ID NO: 18 (soybean phyB3 GAF domain), Tyr128 of SEQ ID NO: 19 (soybean phyB4 GAF domain), Tyr128 of SEQ ID NO: 20 (potato phyB GAF domain), Tyr128 of SEQ ID NO: 21 (pea phyB GAF domain), and Tyr128 of SEQ ID NO: 22 (grape phyB GAF domain).

As shown in FIG. 13, Tyr361 of SEQ ID NO: 1 (At phyB_GAF; *Arabidopsis* phyB) also corresponds to Tyr130 of the cyanobacteriophytochrome GAF domain from *Synechocystis* PCC6803 (Syn Cph_GAF; SEQ ID NO: 34), to Tyr 130 of the bacteriophytochrome GAF domain from *Deinococcus radiodurans* (Dr Bph_GAF; SEQ ID NO: 35), to Tyr 130 of the bacteriophytochrome GAF domain from *Pseudomonas aeruginosa* (Pa BphP_GAF; SEQ ID NO: 36), to Tyr130 of the bacteriophytochrome GAF domain from *Rhodospseudomonas palustris* (Rp BphP3_GAF; SEQ ID NO: 37), and to Tyr130 of the cyanobacteriophytochrome GAF domain from *Synechococcus* OS-B (SyB Cph_GAF; SEQ ID NO: 38).

The invention also relates to nucleic acids that selectively hybridize to the exemplified sequences, including hybridizing to the exact complements of these sequences. The specificity of single-stranded DNA to hybridize complementary fragments is determined by the "stringency" of the reaction conditions (Sambrook et al., 1989). Hybridization stringency increases as the propensity to form DNA duplexes decreases. In nucleic acid hybridization reactions, the stringency can be chosen to favor specific hybridizations (high stringency), which can be used to identify, for example, full-length clones from a library. Less-specific hybridizations (low stringency) can be used to identify related, but not exact (homologous, but not identical), DNA molecules or segments.

DNA duplexes are stabilized by: (1) the number of complementary base pairs; (2) the type of base pairs; (3) salt

concentration (ionic strength) of the reaction mixture; (4) the temperature of the reaction; and (5) the presence of certain organic solvents, such as formamide, which decrease DNA duplex stability. In general, the longer the probe, the higher the temperature required for proper annealing. A common approach is to vary the temperature; higher relative temperatures result in more stringent reaction conditions.

To hybridize under "stringent conditions" describes hybridization protocols in which nucleotide sequences at least 60% homologous to each other remain hybridized. Generally, stringent conditions are selected to be about 5° C. lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength, pH, and nucleic acid concentration) at which 50% of the probes complementary to the target sequence hybridize to the target sequence at equilibrium. Since the target sequences are generally present at excess, at T_m, 50% of the probes are occupied at equilibrium.

"Stringent hybridization conditions" are conditions that enable a probe, primer, or oligonucleotide to hybridize only to its target sequence (e.g., SEQ ID NO:1). Stringent conditions are sequence-dependent and will differ. Stringent conditions comprise: (1) low ionic strength and high temperature washes, for example 15 mM sodium chloride, 1.5 mM sodium citrate, 0.1% sodium dodecyl sulfate, at 50° C.; (2) a denaturing agent during hybridization, e.g. 50% (v/v) formamide, 0.1% bovine serum albumin, 0.1% Ficoll, 0.1% polyvinylpyrrolidone, 50 mM sodium phosphate buffer (750 mM sodium chloride, 75 mM sodium citrate; pH 6.5), at 42° C.; or (3) 50% formamide. Washes typically also comprise 5×SSC (0.75 M NaCl, 75 mM sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5×Denhardt's solution, sonicated salmon sperm DNA (50 µg/ml), 0.1% SDS, and 10% dextran sulfate at 42° C., with a wash at 42° C. in 0.2×SSC (sodium chloride/sodium citrate) and 50% formamide at 55° C., followed by a high-stringency wash consisting of 0.1×SSC containing EDTA at 55° C. Preferably, the conditions are such that sequences at least about 65%, 70%, 75%, 85%, 90%, 95%, 98%, or 99% homologous to each other typically remain hybridized to each other. These conditions are presented as examples and are not meant to be limiting.

"Moderately stringent conditions" use washing solutions and hybridization conditions that are less stringent, such that a polynucleotide will hybridize to the entire, fragments, derivatives, or analogs of the target sequence (e.g., SEQ ID NO:1). One example comprises hybridization in 6×SSC, 5×Denhardt's solution, 0.5% SDS and 100 µg/ml denatured salmon sperm DNA at 55° C., followed by one or more washes in 1×SSC, 0.1% SDS at 37° C. The temperature, ionic strength, etc., can be adjusted to accommodate experimental factors such as probe length. Other moderate stringency conditions have been described (Ausubel et al., 1993; Kriegler, 1990).

"Low stringent conditions" use washing solutions and hybridization conditions that are less stringent than those for moderate stringency, such that a polynucleotide will hybridize to the entire, fragments, derivatives, or analogs of the target sequence (e.g., SEQ ID NO:1). A nonlimiting example of low stringency hybridization conditions includes hybridization in 35% formamide, 5×SSC, 50 mM Tris HCl (pH 7.5), 5 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.2% BSA, 100 µg/ml denatured salmon sperm DNA, 10% (wt/vol) dextran sulfate at 40° C., followed by one or more washes in 2×SSC, 25 mM Tris HCl (pH 7.4), 5 mM EDTA, and 0.1% SDS at 50° C. Other conditions of low stringency, such

as those for cross-species hybridizations, are well-described (Ausubel et al., 1993; Kriegler, 1990).

A "functional homolog," "functional equivalent," or "functional fragment" of a polypeptide of the present invention is a polypeptide that is homologous to the specified polypeptide but has one or more amino acid differences from the specified polypeptide. A functional fragment or equivalent of a polypeptide retains at least some, if not all, of the activity of the specified polypeptide.

Transgenic plants and methods of producing transgenic plants are provided. Such transgenic plants are produced, in certain embodiments, by introducing into a plant or plant cell a polynucleotide encoding a polypeptide comprising a sequence having at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, or at least about 99% identity to at least one amino acid sequence selected from SEQ ID NOs: 1-22, wherein the tyrosine corresponding to Y361 of SEQ ID NO:1 is replaced with a different amino acid. In certain embodiments, the polynucleotide is provided as a construct in which a promoter is operably linked to the polynucleotide. Such transgenic plants may also be produced, in certain embodiments, by introducing into a plant or plant cell a polynucleotide encoding a polypeptide comprising a sequence having at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, or at least about 99% identity to at least one sequence selected from the amino acid sequence selected from SEQ ID NOs: 1-22, wherein the tyrosine corresponding to Y361 of SEQ ID NO:1 is replaced with a different amino acid.

The polynucleotide sequences can be introduced into plants which do not express the corresponding native form of unmodified phyB, such as plants lacking the native gene, or containing a mutated, truncated or downregulated version of the native gene, such that little or no phyB polypeptide is expressed, or a phyB polypeptide is expressed that is partially or substantially inactive. The modified phyB replaces or substitutes for the native gene function. The polynucleotides can also be expressed in wild-type plants containing the corresponding native phyB gene sequence. In this case, the modified phyB over-rides the functions of the wild type endogenous gene in a dominant fashion, since it is hyperactive.

Plants expressing the modified phyB have surprisingly altered light sensitivity and altered photoresponses. Altered photoresponses relative to a control plant include, without limitation, at least one of an improved germination efficiency of seeds, such as in low light or following a pulse of white light, a hypersensitivity to white and red light with respect to hypocotyl and stem growth, improved shade tolerance, a smaller mature plant size, reduced plant height, smaller mature plant diameter, decreased petiole length, reduced internode length, shorter stems, smaller stem diameter, increased leaf chlorophyll concentration, decreased leaf length, increased root length, increased root branching, improved leaf unfolding, flatter leaves (increased hypostasy), reduced leaf surface area and combinations thereof.

Plants expressing modified phyB comprising a substitution at Tyr361 are smaller in size and more tolerant of low light conditions such as would be experienced in crowded field conditions. In one embodiment, plants expressing the modified phyB grow more effectively when planted in higher densities, permitting higher yields over a given planting area.

The Y361F substitution generates a hyperactive photoreceptor that still requires light for activation. As such, plants expressing the modified Y361F phytochrome display accen-

tuated phyB signaling, useful in agricultural settings with fewer side effects. The replacement of wild-type phyB with phyB^{1361F} in plants increases the sensitivity of hypocotyls to R, generates seeds with a stronger germination response in white light, and further accentuates the end-of-day far-red light (EOD-FR) response of seedlings, substantially without altering flowering time, such as in short days. The phyB-mediated responses to R and EOD-FR are connected to the shade avoidance response. Without wishing to be bound to any theory, it is possible that increased signaling by the phyB^{1361F} variant attenuates shade avoidance response by enabling the small amounts of Pfr generated by low fluence R, or the residual Pfr remaining after EOD-FR (or presumably in high FR/R light environments) to more effectively promote normal photomorphogenesis.

It is envisaged that a plant produced following the introduction of a polynucleotide disclosed herein exhibits altered or modified characteristics relative to the control plant. The modified characteristics include, but are not limited to, increased hyponasty, decreased height, decreased diameter, decreased petiole length, decreased internode length, decreased stem diameter, decreased hypocotyl length under an R fluence rate of less than 1 $\mu\text{mole m}^{-2} \text{sec}^{-1}$ (or less than 0.5 $\mu\text{mole m}^{-2} \text{sec}^{-1}$, less than 0.6 $\mu\text{mole m}^{-2} \text{sec}^{-1}$, less than 0.7 $\mu\text{mole m}^{-2} \text{sec}^{-1}$, or less than 0.8 $\mu\text{mole m}^{-2} \text{sec}^{-1}$), enhanced germination or any combination thereof. The altered characteristic may be decreased or enhanced by at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, at least about 100%, at least about 110%, at least about 120%, at least about 130%, at least about 140%, at least about 150%, at least about 175%, at least about 200%, at least about 250%, at least about 300%, or at least about 400% relative to a control plant.

As a nonlimiting example, such modified plants may have a compact size and have a height or diameter that is at least about 20%, at least about 30%, at least about 50%, at least about 75%, or at least about 100% smaller than the height or diameter of a control plant. As another nonlimiting example, such modified plants may provide an increased yield of seed, grain, forage, fruit, root, leaf, or combination thereof that is at least about 5%, at least about 10%, at least about 20%, at least about 25%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 75%, at least about 80%, at least about 90%, or at least about 100% increased over the yield from corresponding control plants. As used herein, "yield" refers to the maximum yield achievable per given planting area, and does not refer to the yield from an individual plant. Maximum or higher yields may be achieved by planting a higher number or density of plants in a given area.

As used herein, a "control plant" is a plant that is substantially equivalent to a test plant or modified plant in all parameters with the exception of the test parameters. For example, when referring to a plant into which a polynucleotide according to the present invention has been introduced, in certain embodiments, a control plant is an equivalent plant into which no such polynucleotide has been introduced. In certain embodiments, a control plant is an equivalent plant into which a control polynucleotide has been introduced. In such instances, the control polynucleotide is one that is expected to result in little or no phenotypic effect on the plant.

The polynucleotides of the present invention may be introduced into a plant cell to produce a transgenic plant. As used herein, "introduced into a plant" with respect to poly-

nucleotides encompasses the delivery of a polynucleotide into a plant, plant tissue, or plant cell using any suitable polynucleotide delivery method. Methods suitable for introducing polynucleotides into a plant useful in the practice of the present invention include, but are not limited to, freeze-thaw method, microparticle bombardment, direct DNA uptake, whisker-mediated transformation, electroporation, sonication, microinjection, plant virus-mediated, and *Agrobacterium*-mediated transfer to the plant. Any suitable *Agrobacterium* strain, vector, or vector system for transforming the plant may be employed according to the present invention. In certain embodiments, the polynucleotide is introduced using at least one of stable transformation methods, transient transformation methods, or virus-mediated methods.

By "stable transformation" is intended that the nucleotide construct introduced into a plant integrates into the genome of the plant and is capable of being inherited by progeny thereof. By "transient transformation" is intended that a nucleotide construct introduced into a plant does not integrate into the genome of the plant.

Transformation protocols as well as protocols for introducing nucleotide sequences into plants may vary depending on the type of plant or plant cell, i.e., monocot or dicot, targeted for transformation. Suitable methods of introducing nucleotide sequences into plant cells and subsequent insertion into the plant genome include microinjection (Crossway et al. (1986) *Biotechniques* 4:320-334), electroporation (Riggs et al. (1986) *Proc. Natl. Acad. Sci. USA* 83:5602-5606, *Agrobacterium*-mediated transformation (U.S. Pat. Nos. 5,981,840 and 5,563,055), direct gene transfer (Paszowski et al. (1984) *EMBO J.* 3:2717-2722), and ballistic particle acceleration (see, for example, U.S. Pat. Nos. 4,945,050; 5,879,918; 5,886,244; 5,932,782; Tomes et al. (1995) in *Plant Cell, Tissue, and Organ Culture: Fundamental Methods*, ed. Gamborg and Phillips (Springer-Verlag, Berlin); and McCabe et al. (1988) *Biotechnology* 6:923-926). Also see Weissinger et al. (1988) *Ann. Rev. Genet.* 22:421-477; Sanford et al. (1987) *Particulate Science and Technology* 5:27-37 (onion); Christou et al. (1988) *Plant Physiol.* 87:671-674 (soybean); McCabe et al. (1988) *Bio/Technology* 6:923-926 (soybean); Finer and McMullen (1991) *In Vitro Cell Dev. Biol.* 27P:175-182 (soybean); Singh et al. (1998) *Theor. Appl. Genet.* 96:319-324 (soybean); Datta et al. (1990) *Biotechnology* 8:736-740 (rice); Klein et al. (1988) *Proc. Natl. Acad. Sci. USA* 85:4305-4309 (maize); Klein et al. (1988) *Biotechnology* 6:559-563 (maize); U.S. Pat. Nos. 5,240,855; 5,322,783 and 5,324,646; Klein et al. (1988) *Plant Physiol.* 91:440-444 (maize); Fromm et al. (1990) *Biotechnology* 8:833-839 (maize); Hooykaas-Van Slogteren et al. (1984) *Nature (London)* 311:763-764; U.S. Pat. No. 5,736,369 (cereals); Bytebier et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:5345-5349 (Liliaceae); De Wet et al. (1985) in *The Experimental Manipulation of Ovule Tissues*, ed. Chapman et al. (Longman, N.Y.), pp. 197-209 (pollen); Kaeppler et al. (1990) *Plant Cell Reports* 9:415-418 and Kaeppler et al. (1992) *Theor. Appl. Genet.* 84:560-566 (whisker-mediated transformation); D'Halluin et al. (1992) *Plant Cell* 4:1495-1505 (electroporation); Li et al. (1993) *Plant Cell Reports* 12:250-255 and Christou and Ford (1995) *Annals of Botany* 75:407-413 (rice); Osjoda et al. (1996) *Nature Biotechnology* 14:745-750 (maize via *Agrobacterium tumefaciens*); all of which are herein incorporated by reference in their entireties.

In some embodiments, a plant may be regenerated or grown from the plant, plant tissue or plant cell. Any suitable methods for regenerating or growing a plant from a plant cell

or plant tissue may be used, such as, without limitation, tissue culture or regeneration from protoplasts. Suitably, plants may be regenerated by growing transformed plant cells on callus induction media, shoot induction media and/or root induction media. See, for example, McCormick et al. (1986) Plant Cell Reports 5:81-84. These plants may then be grown, and either pollinated with the same transformed strain or different strains, and the resulting hybrid having expression of the desired phenotypic characteristic identified. Two or more generations may be grown to ensure that expression of the desired phenotypic characteristic is stably maintained and inherited and then seeds harvested to ensure expression of the desired phenotypic characteristic has been achieved. Thus as used herein, "transformed seeds" refers to seeds that contain the nucleotide construct stably integrated into the plant genome.

In certain embodiments, the polynucleotides to be introduced into the plant are operably linked to a promoter sequence and may be provided as a construct. As used herein, a polynucleotide is "operably linked" when it is placed into a functional relationship with a second polynucleotide sequence. For instance, a promoter is operably linked to a coding sequence if the promoter is connected to the coding sequence such that it may effect transcription of the coding sequence. In various embodiments, the polynucleotides may be operably linked to at least one, at least two, at least three, at least four, at least five, or at least ten promoters.

Promoters useful in the practice of the present invention include, but are not limited to, constitutive, inducible, temporally-regulated, developmentally regulated, chemically regulated, tissue-preferred and tissue-specific promoters. Suitably, the promoter causes sufficient expression in the plant to produce the phenotypes described herein. Suitable promoters include, without limitation, the 35S promoter of the cauliflower mosaic virus, ubiquitin, tCUP cryptic constitutive promoter, the Rsyn7 promoter, pathogen-inducible promoters, the maize In2-2 promoter, the tobacco PR-1a promoter, glucocorticoid-inducible promoters, and tetracycline-inducible and tetracycline-repressible promoters.

It is envisaged that analogous substitutions of tyrosine at positions corresponding to Tyr361 of SEQ ID NO: 1 should elicit similar altered light sensitivity and photo responses when expressed in other plants. Plants that may express a modified phytochrome include, among others, crop plants and ornamental plants.

Suitable plant species include, without limitation, corn (*Zea mays*), soybean (*Glycine max*), *Brassica* sp. (e.g., *Brassica napus*, *B. rapa*, and *B. juncea*), alfalfa (*Medicago sativa*), rice (*Oryza sativa*), rye (*Secale cereale*), sorghum (*Sorghum bicolor*, *Sorghum vulgare*), millet (e.g., pearl millet (*Pennisetum glaucum*), proso millet (*Panicum miliaceum*), foxtail millet (*Setaria italica*), finger millet (*Eleusine coracana*), sunflower (*Helianthus annuus*), safflower (*Carthamus tinctorius*), wheat (*Triticum aestivum*), tobacco (*Nicotiana tabacum*), potato (*Solanum tuberosum*), peanuts (*Arachis hypogaea*), cotton (*Gossypium barbadense*, *Gossypium hirsutum*), sweet potato (*Ipomoea batatas*), cassava (*Manihot esculenta*), coffee (*Cofea* spp.), coconut (*Cocos nucifera*), pineapple (*Ananas comosus*), citrus trees (*Citrus* spp.), cocoa (*Theobroma cacao*), tea (*Camellia sinensis*), banana (*Musa* spp.), avocado (*Persea americana*), fig (*Ficus casica*), guava (*Psidium guajava*), mango (*Mangifera indica*), olive (*Olea europaea*), papaya (*Carica papaya*), cashew (*Anacardium occidentale*), macadamia (*Macadamia integrifolia*), almond (*Prunus amygdalus*), sugar beets (*Beta*

vulgaris), sugarcane (*Saccharum* spp.), oats (*Avena sativa*), barley (*Hordeum vulgare*), vegetables, ornamentals, and conifers.

Vegetables include, without limitation, tomatoes (*Lycopersicon esculentum*), lettuce (e.g., *Lactuca sativa*), green beans (*Phaseolus vulgaris*), lima beans (*Phaseolus limensis*), peas (*Lathyrus* spp.), and members of the genus *Cucumis* such as cucumber (*C. sativus*), cantaloupe (*C. cantalupensis*), and musk melon (*C. melo*).

Ornamental plants are plants that are grown for decorative purposes in gardens and landscapes, as houseplants, and for cut flowers. Suitable ornamentals include, without limitation, azalea (*Rhododendron* spp.), hydrangea (*Macrophylla hydrangea*), hibiscus (*Hibiscus rosasanensis*), roses (*Rosa* spp.), tulips (*Tulipa* spp.), daffodils (*Narcissus* spp.), petunias (*Petunia hybrida*), carnation (*Dianthus caryophyllus*), poinsettia (*Euphorbia pulcherrima*), and chrysanthemum (*Chrysanthemum* spp.).

As used herein, the term "plant" includes reference to whole plants, plant organs (e.g., leaves, stems, roots, etc.), seeds, plant cells, and progeny of same. Parts of transgenic plants comprise, for example, plant cells, protoplasts, tissues, callus, embryos as well as flowers, ovules, stems, fruits, leaves, roots originating in transgenic plants or their progeny previously transformed with a DNA. As used herein, the term "plant cell" includes, without limitation, protoplasts and cells of seeds, suspension cultures, embryos, meristematic regions, callus tissue, leaves, roots, shoots, gametophytes, sporophytes, pollen, and microspores.

Consequently, this invention encompasses transgenic crops and other plants with improved shade tolerance needed for increased planting density and increased yields.

It will be apparent to those of skill in the art that variations may be applied to the compositions and methods described herein and in the steps or in the sequence of steps of the method described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents which are both chemically and physiologically related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention.

It is to be understood that the invention is not limited in its application to the details of construction and the arrangement of components set forth in the following description. Also, it is to be understood that the phraseology and terminology used herein is for the purpose of description and should not be regarded as limiting. The use of "including," "comprising," or "having" and variations thereof herein is meant to encompass the items listed thereafter and equivalents thereof as well as additional items.

It also is understood that any numerical range recited herein includes all values from the lower value to the upper value. For example, if a concentration range is stated as 1% to 50%, it is intended that values such as 2% to 40%, 10% to 30%, or 1% to 3%, etc., are expressly enumerated in this specification. These are only examples of what is specifically intended, and all possible combinations of numerical values between and including the lowest value and the highest value enumerated are to be considered to be expressly stated in this application.

The following non-limiting examples are purely illustrative.

Materials and Methods

Recombinant phyB Protein Expression, Purification, and Analysis

All the site-directed mutations in PHYB were introduced into the cDNA by the Quikchange method (Stratagene). cDNA fragments encoding the photosensory modules (residues 1-624) were appended in-frame corresponding to the N-terminus of the 6His tag (KLHHHHHHH) (SEQ ID NO: 39) by introduction into the pBAD plasmid (Invitrogen), and then co-transformed into *Escherichia coli* BL21 (AI) cells (Invitrogen) with the pPL-PΦB plasmid expressing the *Synechocystis* PCC6803 HO1 heme oxygenase and *A. thaliana* HY2 PΦB synthase enzymes [40, 41] to direct apoprotein expression and chromophore assembly. Following sequential induction of the HO1/HY2 genes and PHYB genes with IPTG and arabinose, the cells were disrupted by sonication in extraction buffer (50 mM HEPES-NaOH (pH 7.8), 300 mM NaCl, 30 mM imidazole, 0.1% Tween-20, 10% glycerol, 1 mM 2-mercaptoethanol, and 1 mM PMSF) with the addition of 1 tablet of protease inhibitor cocktail (Roche) before use. The clarified supernatant was applied to a HisTrap HP column (GE) pre-equilibrated in extraction buffer, and the column was washed with extraction buffer followed by elution with a 30-300 mM imidazole gradient in extraction buffer. The phyB-containing fractions were pooled, dialyzed against 10 mM HEPES-NaOH (pH 7.8), 100 mM NaCl, 5 mM 2-mercaptoethanol, 5 mM Na₂EDTA, 50 mM imidazole, and 0.05% Tween-20 overnight, and subjected to size-exclusion chromatography using a 24-ml Superose 6 (GE) column pre-equilibrated with the same buffer. phyB-containing fractions were pooled and stored in 10 mM HEPES-NaOH (pH 7.8), 50 mM NaCl, 1 mM 2-mercaptoethanol, 0.05% Tween-20, and 10% glycerol.

Pr-Pfr photointerconversion and Pfr→Pr thermal-reversion of each phyB preparation were assayed by UV-vis absorption spectroscopy at 24° C., using white light filtered through 650- and 730-nm interference filters (Orion) to drive Pr→Pfr and Pfr→Pr phototransformation, respectively.

Plant Materials and Growth Conditions

All the plant lines were derived from *A. thaliana* Col-0 ecotype. The phyB-9 and phyA-211 alleles were as described [30, 32]. Seeds were surface-sterilized in chlorine gas, and stratified in water for 3 d at 4° C. before sowing. Unless otherwise noted, seedlings were grown at 22° C. under white light in LD (16-hr light/8-hr dark) on 0.7% (w/v) agar medium containing 1× Gamborg's (GM) salts, 2% (w/v) sucrose, 0.5 g/L MES (pH 5.7). After 10 d, seedlings were transferred to soil and grown at 22° C. under continuous white light in LD or SD (8-hr light/16-hr dark).

Plasmid Constructions for Plant Transformation

The full coding regions of PHYA and PHYB [39] were inserted into the pDONR221 plasmid via BP reactions (Invitrogen), and appended the coding sequence in-frame for the FLAG-epitope (GGGDYKDDDDK) (SEQ ID NO: 40) to their 3' ends. The PHYA/B promoter and 5' UTRs (2634- and 1983-bp upstream beginning at the ATG translation initiation codon), and 3' UTRs (242- and 279-bp downstream of the translation termination codon) were amplified by PCR from the Col-0 genomic DNA, and then sequentially inserted into the pDONR211 plasmids to appropriately flank the coding regions. The completed PHYB and PHYA transgenes were introduced into the pMDC123 plasmid (Invitrogen) via LR reactions. The PHYB-yellow fluorescent protein (YFP) constructions (WT-YFP, R322A-YFP,

Y361F-YFP, D307A-YFP, and R582A-YFP) were created by appending the UBQ10 promoter fragment (1986-bp fragment proximal to the ATG codon) and the cDNA encoding YFP, to the 5' and 3' ends of the PHYB cDNA in a pDONR211 plasmid, respectively. The complete transgenes were introduced into the pMDC123 plasmid via LR reactions.

Plant Transformation and Selection of Transgenic Lines

The PHYA and PHYB transgenes were introduced into the homozygous *Arabidopsis* phyA-211 or phyB-9 mutants, respectively, via the *Agrobacterium*-mediated floral dip method using the pMDC123-derived plasmids [42]. Transformed lines were selected by resistance to 10 μg/mL BASTA. T2 transgenic plants with a resistance segregation ratio of ~3:1 were used to obtain isogenic lines in the T4 or T5 generation for all the biochemical, phenotypic, and localization assays.

Protein Extraction and Immunoblot Analysis

Five-d-old, dark-grown *Arabidopsis* seedlings were frozen and pulverized at liquid nitrogen temperatures, and homogenized in 100 mM Tris-HCl (pH 8.5), 10 mM Na₂EDTA, 25% ethylene glycol, 2 mM PMSF, 10 mM N-ethylmaleimide, 5 μg/mL sodium metabisulfite, 2% (w/v) SDS, 10 μg/mL aprotinin, 10 μg/mL leupeptin and 0.5 μg/mL pepstatin [43]. The extracts were heated to 100° C. for 10 min and clarified by centrifugation at 13,000×g for 10 min. The supernatants were subjected to SDS-PAGE and immunoblot analysis with a monoclonal antibody against phyA (073D, [44]), phyB (B1-B7, [45]), or green fluorescent protein (GFP) (Sigma). Anti-PBA1 antiserum or anti-histone H3 antibodies were used to confirm equal protein loading [46].

To measure phyB degradation in response to Rc, seeds were sown in liquid medium containing half-strength Murashige and Skoog (MS) salts, 0.5 g/L MES (pH 5.7), and 10 g/L sucrose, and irradiated with white light (24 hr for seeds carrying the PHYB^{D307A} transgene and 12 hr for all others) to initiate germination before maintaining the seedlings in the dark for 4 d. Seedlings were collected after various exposure times to continuous 20 μmol·m⁻²·s⁻¹ R and subjected to immunoblot analysis as above. Seedlings were incubated for 12 hr in the dark with 100 μM MG132 or an equivalent volume of DMSO before R.

Phenotypic Assays

Germination efficiency was measured according to Oh et al. (Plant Cell 19, 1192-1208). The parental plants (5 per genotype) were grown side by side at 22° C. in LDs, and the resulting seeds were harvested as separate seed pools. At least 60 seeds from each pool were sown on 0.7% (w/v) water agar after 20-min FR irradiation (4 μmol m⁻² s⁻¹). The seeds were then exposed to white light for 2 hr, and either kept in dark or irradiated with 4 μmol m⁻² s⁻¹ FR for 5 min. The plates were kept in darkness for an additional 5 d before measurement of germination, which was scored as emergence of the radical from the seed coat. For hypocotyl elongation, seeds were sown on solid half-strength MS salts, 0.5 g/L MES (pH 5.7), and 0.7% (w/v) agar, and irradiated with 12-hr white light. The plates were exposed to either R or FR for 3.5 d using a bank of diodes (E-30LED-controlled environment chamber, Percival), before measurement of hypocotyl length. For measurement of the EOD-FR response, seedlings were irradiated over a 4-d cycle with 90 μmol·m⁻²·s⁻¹ R for 8 hr followed by either darkness or by a 10-min pulse of 100 μmol·m⁻²·s⁻¹ FR and then darkness for 16 hr. Effect on flowering time was measured for plants grown under white light in SD.

Confocal Microscopic Analysis

Transgenic seeds expressing wild-type and mutant versions of phyB-YFP were sown on solid medium containing half-strength MS salts, 0.5 g/L MES (pH 5.7), 2% (w/v) sucrose, and 0.7% (w/v) agar and irradiated for 12 hr at 22°C with white light before incubation in the dark for 5 d. Fluorescence of hypocotyl cells, either kept in the dark or irradiated with 90 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ R for 12 hr, was imaged using a Zeiss 510-Meta laser scanning confocal microscope. YFP fluorescence was visualized in the single-track mode by excitation with 488-nm light using the BP 500-530 IR filter. Images were processed with the LSM510 image browser.

Example 2

Rational Design of phyB Variants to Alter Light Signaling

Site-directed substitutions of certain amino acids based on the microbial scaffolds were introduced into the *Arabidopsis* phyB isoform. The photochemistry of the mutant photosensory modules was examined after recombinant assembly with the native chromophore phytylchromobilin (FOB), and the full-length versions were assessed for their phenotypic rescue of the phyB-9 null mutant using the native PHYB promoter to drive expression. The results collectively demonstrate that various aspects of phy dynamics and signaling can be adjusted (FIG. 1A), which in some cases generates plants with unique photobehavioral properties.

We examined five mutations predicted to compromise Pr to Pfr photoconversion, interaction of the bilin with its binding pocket, and/or possible signal transmission from the cGMP phosphodiesterase/adenylyl cyclase/FhlA (GAF) domain to the downstream phytochrome (PHY) domain in the photosensory module. As shown in FIGS. 1B and 1C, the D307A substitution removes a key aspartate (D207 in *Synechocystis* PCC6803 (Syn)-Cph1) that participates through its main chain carbonyl in a unique hydrogen-bonding lattice involving the A-C pyrrole rings of P Φ B and the centrally positioned pyrrole water. This invariant aspartate is essential for the Pr to Pfr photoconversion of bacterial phys as most, if not all, substitutions are stalled in the deprotonation/protonation cycle following R irradiation and become highly fluorescent [18-20]. Two substitutions alter the hydrogen-bond contacts with D307. The relatively mild Y361F substitution (Y263 in SynCph1) maintains the aromatic character but is expected to eliminate the hydrogen bond that helps hold the side chain carboxyl group of D307 in place [18, 22], whereas the R582A substitution (R472 in SynCph1) removes a potential salt bridge between D307 and a novel hairpin, likely universal among canonical phys, that extends from the PHY domain to contact the GAF domain near the bilin. This hairpin may help transmit chromophore movements within the photosensory module to the C-terminal output region during photoconversion [3, 7, 10] (FIG. 1B,C). The last two mutations (R352A and R322A) eliminate salt bridges between the propionate side chains in P Φ B and the bilin-binding pocket, which presumably help restrain the bilin within the photoreceptor (FIG. 1B,C). Prior studies with bacterial phys (R222 and R254 in *Synechococcus* OS-B' (SyB)-Cph1) showed that these arginines stabilize and destabilize the Pfr conformer, respectively, with their guanidinium side chains undergoing dramatic conformational changes during Pr \rightarrow Pfr photoconversion.

Based on the phy scheme presented in FIG. 1A, we tested how well the *Arabidopsis* phyB mutants would: (i) assemble with P Φ B, (ii) photointerconvert between Pr and Pfr, (iii)

revert thermally from Pfr back to Pr, (iv) accumulate and concentrate after R irradiation into nuclear bodies or "speckles" thought to be important for signaling and/or turnover [23], (v) degrade upon R irradiation, and (vi) stimulate several photomorphogenic processes under full or partial control by phyB, including R-stimulated seed germination, hypocotyl growth inhibition under R, effect of end-of-day (EOD) FR on the hypocotyl R response, and flowering time under a short-day photoperiod (SD) [2, 13, 14]. Pfr turnover is likely driven by the ubiquitin/26S proteasome system (UPS) based on mutant analyses and its sensitivity to the proteasome inhibitor MG132 ([24, 25] see FIG. 6A). Methods used to synthesize photoactive photosensory module fragments recombinantly from phyB assembled with P Φ B and to generate the transgenic plants expressing full-length versions are provided in Example 1.

The photosensory module of all the mutants could be expressed and readily assembled with P Φ B in *Escherichia coli*, and generated reasonably normal Pr absorption spectra with maxima at \sim 663 nm, indicating that none of the substitutions compromised protein folding or bilin conjugation (FIG. 2A; FIG. 7). Given that *Arabidopsis* and other plants are highly sensitive to phy levels [26-29], we chose two isogenic phyB-9 lines in the T3 generation that expressed either unaltered phyB or the mutants to levels which matched most closely that in the wild-type Col-0 plants as judged by immunoblot analysis (FIG. 3D). Importantly, all of the complemented phyB-9 mutant lines had normal etiolated seedling development, indicating that none of the phyB variants signaled in the absence of photoactivation (FIG. 3B).

To examine the ability of the mutants to concentrate in nuclear bodies/speckles as Pfr, we also created a parallel set of transgenic lines expressing the phyB mutants as N-terminal fusions to yellow fluorescent protein (YFP). These bodies were easily seen by confocal fluorescence microscopy as numerous intense puncta that accumulate in the nucleus upon prolonged R irradiation (see FIG. 6B). The phyB-YFP mutant proteins also assembled well with P Φ B in planta, and phenotypically resembled their non-tagged counterparts based on their ability (or inability) to suppress hypocotyl elongation in R when introduced into the phyB-9 background (FIG. 8).

Example 3

phyB^{Y361F} is Hypersensitive to R

Given its predicted proximity to D307 within the bilin-binding pocket, Y361 likely helps enclose the GAF domain around the bilin and fix the position of D307 (FIG. 1B). Surprisingly, the Y361F substitution in *Arabidopsis* phyB permitted proper photochemistry but made the photoreceptor hyperactive with respect to signaling. Recombinant phyB^{Y361F} had relatively normal Pr and Pfr absorption spectra, but displayed slightly reduced Pr \rightarrow Pfr and Pfr \rightarrow Pr photoconversion rates and a slower rate of thermal Pfr \rightarrow Pr reversion (FIG. 2A,B).

Despite the expectations that some of these photochemical alterations might compromise signaling, phyB^{Y361F} more effectively directed phyB-mediated responses compared to phyB^{WT}. Soon after germination, the PHYB^{Y361F} phyB-9 seedlings were more sensitive to continuous R with respect to hypocotyl elongation, and as the seedling developed under a long-day (LD) photoperiod, homozygous PHYB^{Y361F} phyB-9 plants had more compact rosettes with shorter petioles than wild type and PHYB^{WT} phyB-9 plants,

indicative of light hypersensitivity (FIGS. 3A,C and 4A). Analyses of hypocotyl elongation at very low R fluence rates (<1 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$) estimated that the phyB^{I361F} biliprotein was at least 50 times more active at signaling at least with respect to this response (FIG. 4B). A modest but statistically significant hypersensitivity to R was also observed for seed germination and the effect of EOD-FR on R-suppressed hypocotyl growth (FIG. 5A,B). Light hypersensitivity continued for the rosettes of mature plants and dampened the SAR (shade avoidance response) as judged by increased hyponasty (i.e., more prostrate petiole angles) and the smaller leaves and shorter petioles seen for PHYB^{I361F} plants as compared to PHYB^{WT} plants when grown in SD (FIGS. 14 A,B and 9). Flowering time in SD was not significantly altered (FIG. 5C), consistent with the minor role of phyB in detecting photoperiod as compared to phyA [30]. Because the hyperactivity of phyB^{I361F} in R could be canceled by EOD-FR, we concluded that phyB^{I361F} photoreceptor still functions in a R-FR reversible manner in planta. Light hypersensitivity was also evident in the rosettes of mature plants as judged by the smaller leaves and shorter petioles seen for PHYB^{I361F} phyB-9 plants as compared to PHYB^{WT} phyB-9 plants when grown in SD (FIG. 9). Despite changes in photochemistry and signaling, the turnover of phyB^{I361F} as Pfr and its sequestering into nuclear bodies appeared normal (FIG. 6A,B).

Example 4

D307 in phyB is Required for Photoconversion, Robust Signaling and Nuclear Body Formation but not Turnover

In line with the predicted importance of D307 in phyB photochemistry [18-20], we found that the assembled phyB^{D307A} biliprotein failed to photoconvert from Pr to Pfr in R and instead generated a bleached R-absorbing intermediate that would regenerate Pr upon irradiation with FR or more slowly upon prolonged dark incubation (FIG. 2A). Unlike phyB^{WT} which restored R-suppressed hypocotyl elongation and enhanced seed germination in R when introduced into the phyB-9 background, the phyB^{D307A} mutant appeared phenotypically inactive or was greatly reduced in phenotypic activity (FIGS. 3 and 4 and 14C). Such compromised activity was also apparent in mature plants as judged by the elongated leaf blades and petioles and strong leaf epinasty (as measured by the large upward angles of petioles) of PHYB^{D307A} phyB-9 plants grown in SD, which better resembled phyB-9 plants as compared to their PHYB^{WT} phyB-9 counterparts (FIGS. 14 A,B and 9). However, detailed fluence response analysis of hypocotyl growth under very high R fluences and the flowering time in SD revealed that phyB^{D307A} retained some signaling activity despite its inability to photoconvert normally to Pfr (FIGS. 4 and 5). Consistent with diminished photochemistry, the accumulation of phyB^{D307A}-YFP in nuclear bodies upon R irradiation was undetectable even after prolonged irradiation with a high fluence rate of R, a condition where the bodies were clearly evident for the wild-type version. But surprisingly its MG132-sensitive turnover in R was only a little slower (FIG. 6A,B), thus providing the first indication that nuclear aggregation of phyB and its degradation after R irradiation are not coupled.

Example 5

The R322A, R352A, and R582A Mutations Poorly Compromise phyB Signaling

R472 in the PHY domain hairpin of Syn-Cph1 forms an inter-domain salt bridge with D207. We examined the effects

of the comparably positioned arginine in *Arabidopsis* phyB (R582) using an alanine substitution. The phyB^{R582A} PSM had normal Pr and Pfr absorption spectra and Pr→Pfr and Pfr→Pr photoconversion rates but was strikingly slower in Pfr→Pr thermal reversion (initial velocity 9.6 times slower than that of phyB^{WT}), indicating that R582 is not required for photochemistry but helps destabilize the Pfr conformer once formed (FIG. 2A,B). The more stable Pfr for phyB^{R582A} in turn likely generates a slightly higher Pfr/Pr ratio in saturating R as evidenced by the reduced peak height at 655 nm versus that at 724 nm (FIG. 2A). However, phyB^{R582A} appeared to signal normally based on the fluence response of hypocotyl growth to continuous R and its ability to delay flowering in SD (FIGS. 3C, 4A, and 5C). In fact, the phyB^{R582A} chromoprotein appeared to be marginally hyperactive as judged by the slightly stronger repression on hypocotyl growth in R for the PHYB^{R582A} phyB-9 #34 line at intermediate R fluence rates as compared to wild type and PHYB^{WT} phyB-9 seedlings despite accumulating similar levels of photoreceptor (FIG. 4A). Moreover, petiole angles were more prostrate and the rosettes appeared more compact than wild-type and PHYB^{WT} plants in SD (FIGS. 14A,B and 9). We speculate that at least some of this increased activity of the phyB^{R582A} chromoprotein may be related to its higher Pfr/Pr photoequilibrium in continuous R. Regardless of the effects on photochemistry, the nuclear aggregation of phyB^{R582A}-YFP, and the turnover of phyB^{R582A} in R appeared normal (FIG. 6A,B).

R352 is predicted to form an essential salt bridge with the propionate group of pyrrole ring B in PΦB, and, based on the mutational analyses of several prokaryotic phyBs, it appears to be important for bilin binding and proper photochemistry [9, 18, 20]. In fact, replacement of this residue with a glutamine in Dr-BphP (R254Q) is one of the few mutations that block covalent attachment of the bilin, whereas the more subtle arginine to alanine mutations in Dr-BphP and SyB-Cph1 effectively inhibit thermal reversion of Pfr back to Pr.

We found that the R352A substitution in *Arabidopsis* phyB has little impact on Pr and Pfr absorption and photochemistry, but like its bacterial relatives [4, 18], the mutation stabilizes Pfr against thermal reversion (FIG. 2A,B). When introduced into phyB-9 seedlings, phyB^{R352A} behaved similarly to wild-type phyB with respect to its ability to suppress hypocotyl growth in R, restore normal rosette morphology to mature plants, delay flowering in SD, and rapidly degrade after photoconversion to Pfr (FIGS. 3A,C, 4A, 5C, and 6, FIG. 9). At most, phyB^{R352A} was marginally hypoactive phenotypically as judged by the reduced response of the PHYB^{R352A} phyB-9 hypocotyls to almost all fluences of R despite accumulating levels of photoreceptor comparable to that in wild type (FIG. 4A). Such subtle phenotypic effects for both phyB^{R352A} and phyB^{R582A} strongly suggest that the thermal reversion of phyB, which would be expected to diminish the active Pfr conformer over time, does not play a major role in phyB signaling under strong light conditions.

Analogous to R352, R322 in the GAF domain is predicted to contact PΦB, with the solution NMR structure of SyB-Cph1 showing that flexibility of its guanidinium side chain allows for transient interactions with the ring C propionate. phyB^{R322A} PSM assembled with PΦB retained normal absorption spectra and Pr→Pfr and Pfr→Pr photoconversion rates, but unlike the R352A substitution, phyB^{R322A} had a substantially faster rate of Pfr→Pr thermal reversion than phyB^{WT} (1.7 times faster; FIG. 2A, B). Thus, whereas R322 is not required for photochemistry, it helps stabilize the Pfr conformer of phyB once formed. Phenotypically, phyB^{R322A}

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behaved similar to phyB^{WT} as judged by its ability to suppress hypocotyl elongation under R in phyB-9 seedlings, rescue the rosette morphology of mature plants, and delay flowering in SD (FIGS. 3A,C, 4A, and 5C, FIG. 8). Similar to the PhyB^{R582A}, phyB^{R322A} relocalized into nuclear bodies and rapidly degraded like endogenous phyB in response to R (FIG. 6).

Example 6

Analysis of Comparable Mutations in *Arabidopsis* phyA

Comparable mutations (D273A, Y327F, R551A, R318A and R288A (FIG. 1C)) were examined to determine if similar effects were observed on phyA signaling. Phenotypically, phyA is the dominant isoform in etiolated seedlings and in plants exposed to FR-rich environments. As shown in FIGS. 10 and 11, phyA signaling can be easily measured by its ability to restore FR suppression of hypocotyl elongation in phyA null mutants such as phyA-211. Using this assay, we found that the D273A mutation also strongly compromises signaling by phyA with a marginal activity seen only at high FR fluence rates (100 $\mu\text{mole}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$). For the Y327F mutation in phyA, the response of the PHYA^{Y327F} phyA-211 seedlings matched that of PHYA^{WT} phyA-211 seedlings at all fluence rates tested. Whereas the R551A and R318A substitutions had no apparent effect on phyA signaling, a slight hypoactivity for phyA harboring the R288A substitution was observed, suggesting that like its phyB counterpart, phyA missing this C ring propionate contact has slightly compromised Pfr activity. When more mature plants grown under white light in LD were examined, all of the phyA mutants developed similar to WT, phyA-211, and PHYA^{WT} phyA-211 plants, suggesting that none of the mutants interfered with phyB signaling or stimulated atypical photomorphogenesis.

Prophetic Example 7

Transgenic Maize

The promoter and coding regions of *Zea mays* (Zm) PHYB1 are cloned from maize genomic DNA and total mRNA, respectively, according to the publicly available *Zea mays* genome sequence data (see Nucleic Acids Res. 40 (Database issue):D1178-86), and are built into a construction containing a Bar gene for Basta resistance and the nopaline synthase transcription terminator directly after the PHYB1 coding region. The corresponding Y361F mutation (Y359F in ZmPHYB1, ZmPHYB1^{Y359F}) is further introduced into the coding region of ZmPHYB1 in the construction via Quikchange method (Stratagene). Transgenic maize is made by *Agrobacterium tumefaciens*-mediated transformation (*Nat. Protoc.* 2: 1614-1621), and selected for Basta resistance. A total of eight transgenic lines at T1 generation are chosen for further screening based on transgene number, phyB protein level and genetic stability from a large pool of transgenic plants (>100 plants), and are grown, self-pollinated to T4 generation to produce isogenic lines for phenotypic assays.

The selected homogeneous transgenic maize containing ZmPHYB1^{Y359F} are grown in green house for phenotypic characterization. After 30 days, the plant height, size of both the transgenic and wild-type maize will be measured, and the flowering time and seed yield will also be recorded in mature plants. These phenotypic data will also be statisti-

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cally analyzed, and compared to wild-type plant. The transgenic lines are expected to have much reduced height and size with unaltered flowering time and seed yield. These dwarf maize are expected to require much less growth space and therefore increase the maize yield per acre.

Prophetic Example 8

Transgenic Rice

The promoter and coding regions of *Oryza sativa* L. (Os) PHYB are cloned from rice genomic DNA and total mRNA, respectively, according to the OsPHYB coding sequence data from National Center for Biotechnology Information, and are built into a construction containing a Neomycin Phosphotransferase II (NPTII) gene for kanamycin resistance and the nopaline synthase transcription terminator directly after the PHYB coding region. The corresponding Y361F mutation (Y368F in OsPHYB) is further introduced into the coding region of OsPHYB in the construction (OsPHYB^{Y368F}) via Quikchange method (Stratagene). Transgenic rice is made by *Agrobacterium tumefaciens*-mediated transformation (*Plant J.* 1994 (2):271-82), and selected for kanamycin resistance. A total of eight transgenic lines at T1 generation are chosen for further screening based on transgene number, phyB protein level and genetic stability from a pool of over 20 transgenic plants, and are grown and self-pollinated to T4 generation to produce isogenic lines for phenotypic assays.

The selected homogeneous transgenic rice containing OsPHYB1^{Y368F} are grown in green house for phenotypic characterization. After 30 days, the plant height and size of both the transgenic and wild-type rice will be measured, and the flowering time and seed yield will also be recorded in mature plants. These phenotypic data will also be statistically analyzed. Compared to the wild-type plant, the transgenic lines are expected to have much reduced height and size with unaltered flowering time and seed yield. These dwarf rice are expected to require much less growth space and therefore increase the rice yield per acre.

Prophetic Example 9

Transgenic Soybean

The promoter and coding regions of *Glycine max* (Gm) PHYB1 are cloned from soybean genomic DNA and total mRNA, respectively, according to the GmPHYB1 coding sequence data from National Center for Biotechnology Information, and are built into a construction containing a Bar gene for Basta resistance and the nopaline synthase transcription terminator directly after the GmPHYB1 coding region. The corresponding Y361F mutation (Y345F in GmPHYB1) is further introduced into the coding region of GmPHYB1 in the construction (GmPHYB1^{Y345F}) via Quikchange method (Stratagene). Transgenic soybean is made by *Agrobacterium tumefaciens*-mediated transformation (*Plant Biotechnol.* 2007, (24): 533-536), and selected for Basta resistance. A total of eight transgenic lines at T1 generation are chosen for further screening based on transgene number, phyB protein level and genetic stability from a large pool of over 100 transgenic plants, and are grown and self-pollinated to T4 generation to produce isogenic lines for phenotypic assays.

The selected homogeneous transgenic soybean containing GmPHYB1^{Y345F} are grown in the green house for phenotypic characterization. After 30 days, the plant height, size of

both the transgenic and wild-type soybean will be measured, and the flowering time and seed yield will also be recorded in mature plants. These phenotypic data will also be statistically analyzed. Compared to the wild-type plant, the transgenic lines are expected to have much reduced height and size with unaltered flowering time and seed yield. These resulting dwarf soybean should require much less growth space and therefore increase the soybean yield per acre.

Prophetic Example 10

Spectroscopy Analyses of Maize phyB Mutants

A library of structure-guided variants has the potential to alter phy signaling in a number of ways, which in turn offers a host of opportunities to manipulate light perception in maize. To test this notion, we will examine how the mutations corresponding to D307A, Y361F, R582A, R352A and R322A of the *Arabidopsis* sequence affect maize phyB photochemistry and/or phyB-directed photomorphogenesis. The residues corresponding to D307A and Y361F alleles are of special interest given their ability to confer hypo- and hypersensitivity to phyB signaling. (For simplicity, the maize alleles are designated in this example using the *Arabidopsis* counterpart residue numbers; D307A is D305A in maize and Y361F is Y359F in maize.)

Using the protocols described herein and illustrated in FIG. 2, we will first examine the photochemical effects of these amino acid substitutions on the recombinant 6His-tagged PSM of maize phyB1 (amino acids 1-623), the dominance of the two maize phyB paralogs with respect to phenotypes. These mutations will be introduced by the Quikchange method (Stratagene) into the full-length ZmPHYB1 cDNA modified to also contain a C-terminal 6His sequence. They will be expressed in *E. coli* by our well defined, two-plasmid pBAD (Invitrogen) system; one LacZ-controlled plasmid encodes the HO (heme oxygenase) from *Synechocystis* PCC6803 and the P Φ B synthase from *Arabidopsis* (HY2 locus) needed to synthesize the P Φ B chromophore from heme, and the second arabinose-controlled plasmid encodes the ZmphyB1 polypeptide. By sequential induction with IPTG and arabinose, high level accumulation of fully assembled and photochemically active ZmphyB1 PSMs will be possible. The recombinant biliproteins will then be purified by nickel-nitrilotriacetic acid (NiNTA) affinity (Qiagen) chromatography based on the 6His tag, followed by Phenyl Sepharose chromatography. Bilin occupancy of the purified photoreceptors will be assessed by zinc-induced fluorescence of the bound chromophore following SDS-PAGE of the preparation. These samples will be examined for atypical absorption spectra, photoconversion rates, and Pfr stability by spectrometric techniques using techniques disclosed herein for FIG. 2.

The maize mutants are expected to show phenotypes similar to those described herein for *Arabidopsis*.

Prophetic Example 11

Assessment of Signaling Strength for the ZmphyB1 Mutants in Maize

The ZmphyB1 mutations generated in prophetic example 10 will be introduced into maize plants and tested for their ability to direct various processes under ZmphyB control. The amino acid substitutions will be introduced into the full-length ZmPHYB1 cDNA, also appended to a DNA sequence encoding a short C-terminal FLAG epitope tag

(GGDYKDDDDK) (SEQ ID NO: 41), and expressed under the control of the native ZmPHYB1 promoter (2-kbp region upstream of the initiation codon). Use of the native promoter will help avoid artifactual responses generated by ectopic expression of the mutant chromoproteins. These transgenes along with a transgene encoding wild-type ZmphyB-FLAG will be stably introduced into maize using a Maize Transformation protocol which exploits the Hi Type-II background for most transformations, generated from a cross between the B73 and A188 hybrids followed by selection for efficient regeneration of plantlets from cultured embryos. The transgenic plants expressing a range of ZmphyB1 polypeptide levels will be identified by immunoblot analysis with available FLAG and phyB-specific monoclonal antibodies. Independent transformants that express the mutant phyB proteins near to that in wild-type plants will be identified since artificially increased or decreased levels of ZmphyB might significantly influence photomorphogenesis by themselves. Those lines deemed useful will then be backcrossed at least three times to the B73 inbred to generate lines suitable to phenotypic testing. A library of suitable independent lines for each mutation will be generated to avoid potential artifacts generated by insertion position of the transgene and/or differing accumulation of the ZmphyB1 biliprotein.

Some mutants (e.g., phyB^{Y361F}) are expected to work dominantly even in the presence of wild-type ZmphyB1/2. However, others will likely confer more subtle phenotypes that will require eliminating the wild-type photoreceptor for observation. This situation will be accomplished through crosses with the ZmphyB1 and ZmphyB2 mutants developed by Sheenan et al. (2007) using Mu insertional mutagenesis, followed by selfing to identify triple homozygous progeny. Single and double mutant combinations will be generated for the strongest ZmphyB1-Mu563 and ZmphyB2-Mu12053 alleles, which have been backcrossed 4 times into both the B73 and W22 backgrounds.

Plants containing unmodified ZmphyB1-FLAG or the mutant (phyB^{D307A}, phyB^{Y361F}, phyB^{R582A}, phyB^{R352A}, and phyB^{R322A}) in either the wild-type B73 or the ZmphyB1-Mu563 and ZmphyB2-Mu12053 B73-introgressed backgrounds will be examined by various phenotypic assays that specifically measure phyB activity. The germplasm will be tested along side several controls including, near isogenic wild-type B73, B73 expressing unmodified ZmphyB1, and the ZmphyB1-Mu563 and ZmphyB2-Mu12053 B73-introgressed lines either singly or as double mutants. To reduce environmental variability, the plants will be grown in controlled environment cabinets equipped with monochromatic R and FR LED light sources and growth chambers illuminated with white light within the lab and greenhouses supplemented with artificial lighting if needed. Randomized block design will be used to avoid biases based on positions of the plants within the group. Testing of plants in outdoor agricultural plots under natural lighting conditions will be carried out to assess their impact on maize seed yield and plant stature in more representative field settings.

The phenotypes to be tested have been well established in maize and include:

- (1) Architecture of seedling grown in the dark (etiolated), which is expected to be unaffected by the mutations.
- (2) Effect of R, FR, R-FR, and white light pulses on coleoptile, mesocotyl, and leaf sheath and blade elongation for young seedlings.
- (3) Effect of EOD-FR on mesocotyl, and leaf blade elongation for young seedlings grown in light/dark cycles.

(4) Chlorophyll and anthocyanin accumulation in seedlings grown in light/dark cycles.

(5) Effect on internode length, stem diameter, and overall plant height on plants grown in long-day photoperiods.

(6) Effect on flowering time for plants grown in long- and short-day photoperiods.

(7) Number of tillers, cobs, and kernels produced in long-days.

Examining a range of R and FR fluence rates on the photomorphogenic responses of young seedlings will facilitate the quantification of the degree of hypo- or hyperactivity for each mutant, particularly the D307A and Y361F mutations that are expected to greatly impact phyB signaling. It is expected that at least some of the *ZmphyB* mutants will confer useful new traits such as altered flowering time or reduced SAR (shade avoidance response) to maize grown in field situations.

Sequences listed in this application include:

SEQ ID NO: 1 is the *Arabidopsis thaliana* phytochrome B (phyB) polypeptide (translation of SEQ ID NO: 23)

SEQ ID NO: 2 is the *Zea mays* phytochrome B polypeptide (translation of SEQ ID NO: 24)

SEQ ID NO: 3 is the *Oryza sativa* Japonica Group isolate SJ-CDI2 phytochrome B (phyB) polypeptide (translation of SEQ ID NO: 25)

SEQ ID NO: 4 is the *Sorghum bicolor* isolate PHYB-Rtx430 phytochrome B (phyB) polypeptide (translation of SEQ ID NO: 26)

SEQ ID NO: 5 is the *Glycine max* phytochrome B-1 (phyB) polypeptide (translation of SEQ ID NO: 27)

SEQ ID NO: 6 is the *Glycine max* phytochrome B-2 (phyB) polypeptide (translation of SEQ ID NO: 28)

SEQ ID NO: 7 is the *Glycine max* phytochrome B-3 (phyB) polypeptide (translation of SEQ ID NO: 29)

SEQ ID NO: 8 is the *Glycine max* phytochrome B-4 (phyB) polypeptide (translation of SEQ ID NO: 30)

SEQ ID NO: 9 is the *Solanum tuberosum* phytochrome B polypeptide (translation of SEQ ID NO: 31)

SEQ ID NO: 10 is the *Pisum sativum* phytochrome B (phyB) polypeptide (translation of SEQ ID NO: 32)

SEQ ID NO: 11 is the *Vitis vinifera* genotype PN40024 phytochrome B (phyB) polypeptide (translation of SEQ ID NO: 33)

SEQ ID NO: 12 is the *Arabidopsis* phyB GAF domain

SEQ ID NO: 13 is the maize phyB GAF domain

SEQ ID NO: 14 is the rice phyB GAF domain

SEQ ID NO: 15 is the *sorghum* phyB GAF domain

SEQ ID NO: 16 is the soybean phyB1 GAF domain

SEQ ID NO: 17 is the soybean phyB2 GAF domain

SEQ ID NO: 18 is the soybean phyB3 GAF domain

SEQ ID NO: 19 is the soybean phyB4 GAF domain

SEQ ID NO: 20 is the potato phyB GAF domain

SEQ ID NO: 21 is the pea phyB GAF domain

SEQ ID NO: 22 is the grape phyB GAF domain

SEQ ID NO: 23 is the *Arabidopsis thaliana* phytochrome B (PHYB) nucleotide (Gen Bank Accession No NM_127435)

SEQ ID NO: 24 is the *Zea mays* phytochrome B nucleotide (Phytozome Accession No. GRMZM2G124532)

SEQ ID NO: 25 is the *Oryza sativa* Japonica Group isolate SJ-CDI2 phytochrome B (PHYB) nucleotide (GenBank Accession No: JN594210)

SEQ ID NO: 26 is the *Sorghum bicolor* isolate PHYB-Rtx430 phytochrome B (PHYB) nucleotide (GenBank Accession No: AY466089)

SEQ ID NO: 27 is the *Glycine max* phytochrome B-1 (PHYB) nucleotide (GenBank: Accession No: EU428749)

SEQ ID NO: 28 is the *Glycine max* phytochrome B-2 (PHYB) nucleotide (GenBank Accession No: EU428750.2)

SEQ ID NO: 29 is the *Glycine max* phytochrome B-3 (PHYB) nucleotide (GenBank Accession No: EU428751.1)

SEQ ID NO: 30 is the *Glycine max* phytochrome B-4 (PHYB) nucleotide (GenBank Accession No: EU428752.1)

SEQ ID NO: 31 is the *Solanum tuberosum* phytochrome B nucleotide (GenBank Accession No: DQ342235.1)

SEQ ID NO: 32 is the *Pisum sativum* phytochrome B (PHYB) nucleotide (GenBank Accession No: AF069305.1)

SEQ ID NO: 33 is the *Vitis vinifera* genotype PN40024 phytochrome B (PHYB) nucleotide (GenBank Accession No: EU436650.1)

SEQ ID NO: 34 is the cyanobacteriophytochrome GAF domain from *Synechocystis* PCC6803 (Syn Cph_GAF)

SEQ ID NO: 35 is the bacteriophytochrome GAF domain from *Deinococcus radiodurans* (Dr Bph_GAF)

SEQ ID NO: 36 is the bacteriophytochrome GAF domain from *Pseudomonas aeruginosa* (Pa BphP_GAF)

SEQ ID NO: 37 is the bacteriophytochrome GAF domain from *Rhodospseudomonas palustris* (Rp BphP3_GAF)

SEQ ID NO: 38 is the cyanobacteriophytochrome GAF domain from *Synechococcus* OS-B (SyB Cph_GAF)

(SEQ ID NO: 1)

MVSGVGGSGGGRRGGRRGEEEPSSSHTPNRRGGEQQSSGTKS

LRPRSNTESMSKAIQQYTV DARLH AVFEQSGESGKSF DYSQSLKTTTYGSSVPEQQIT

AYLSRIQRGGYIQPF GCMIAVDESSFR IIGYSENAREMLGIMPQSVPTLEKPEILAMG

TDVRS LFTSSSSIL LERAFVAREITLLN PVVHIHSKNTGKPFYAILHRIDVGVV IDLEP

ARTEDPALSIAGAVQS QKLAVRAISQLQALPGGDIKLLCDTVVESVRDLTG YDRVMVY

KFHEDEHGEVVAESKRDDLEPYI GLHYPATDIPQASRFLFKQNRVRMIVDCNATPVLV

VQDDRLTQSMCLV GSTLRAPHGCHSQYMANMGSIASLAMA VIINGNEDDGSNVASGRS

SMRLWGLVVCHHTSSRCI PFPLRYACEFLMQAPGLQLNMELQLALQMSEKRVLR TQTL

LCDMLLRDS PAGIVTQSPS IMDLVKCDGAAFLYHGKYYPLGVAPSEVQIKDVVEWLLA

NHADSTGLSTDS LGDAGYPGAALGD AVCGMAVAYI TKRDFLFWFRSHTAKEI KWGGA

KHHPEDKDDGQRMHPRSS FQAFLEVVKRSR SQPWETAEMDAIHS LQLILRDSFKESEAA

MNSKVVDGVVQPCRDMAGEQIDELGAVAREMVR LIETATVPIFAVDAGGCINGWNAK

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 EKQISRIVGDMDESIEDGSFVLKREFFPLGSVINAIIVSQAMFLLRDRGLQLIRDIPE
 EIKSIEVFGDQIRIQQLLAEFLLSIIRYAPSQEWVEIHLSQLSKQADGFAAIRTEFR
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 LELPVPRKRPLSTASGSGDMMMLMPY

(SEQ ID NO: 2)

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 VEHVRELTGYDRVMVYRFHEDEHGEVVAESRRDNLEPYLGLH
 YPATDIPQASRFLFRQNRVMIADCHATPVRV IQDPGLSQPLCLVGLSTRAPHGCHAO
 YMANMGSIASLVMAVI I SSGDDEQ TGRGGISSAMKWLGLVV
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 IIEWLTVPFHGDS TGLSTDSLADAGYLGAAALGEAVCGMAVAYITPSDYLFWFRSHTAK
 EIKWGGAKHHPEDKDDGQRMHPRSSFKAFLVVKRSRSLPWEN
 AEMDAIHSLQLILRDSFRDAEAGTNNKAI VNGQVQLRELELRGINELSSVAREMVRL
 IETATVPIFAVDTDGCINGWNAKIAELTGLSVVEAMGKSLVN
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 IVGVCFVQDVGTGQKVVMDKFNVIQGDYKAIVHNPNLIPPI
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 EIV

(SEQ ID NO: 3)

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 PPPVSLGADARLLFAPSSAVLLERAFAREISLLNPLWIHSRVSSKPFYAILHRIDVG
 VVIDLEPARTEDPALS IAGAVQSQKLAVRAISRQLALPGGDVKLLCDTVVEHVRELTG
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(SEQ ID NO: 4)

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 LLLRERDLQLIRDIPDEIKDASAYGDQFRIQQVLADLFLSMVRSAPSENGWVEIQVRP
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(SEQ ID NO: 5)

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 D ELSSVAREMVRLIETATAPIFAVDVDGHVNGWNAKVELTGLPVEEAMGKSLVHDLV
 F KESEETMKNLLSRALKGEEDKNVEIKMRTFGPEHQNKAVFLVFNACSSKDFTNVVG
 V CFVGDVDTGQKIVMDKFINIQGDYKAI VHSNPLIPPIFASDDNTCCLEWNTAMEKLT
 GWGRVDVIGKMLVGEVFGSCCQLKGSDSITKFMIVLHNLGGQDTDKFPFSLDRHGK
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(SEQ ID NO: 6)

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 SSKC

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(SEQ ID NO: 7)

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 SAVRVVQDEALVQPLCLV GSTLRAPHGCHAQYMANMGS IASLVMAVI INGND EEGVGG
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(SEQ ID NO: 8)

MIAVDEPSFRILAYS DNARDMLGITPQSVPSLDDKNDAAFALGT
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(SEQ ID NO: 9)

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 MVRLIETATAPI FAVDVEGRINGWNAKVAELTGLSV EAMGKSLVHEL VYKESQETAE
 KLLYNALRGEEDKNVEIKLRTFGAEQLEKAVFVVNACASKDYTN NIVGVC FVGQDVT
 GEKVVMDKFINIQGDYKAI VHSNPLIPPI FASDENTCCSEWNTAMEKLTGWSRGEIV
 GKMLVGEIPGSCCRLKGPDMTKFMI VLNHNAI GGQD TDKFPFS FDRNGKYVQALLTA
 NKRVNMEGNTIGAFCFIQIASPELQQALRVQRQOEKKCYSQMKELAYICQEI KSPLNG
 IRFTNSLLEATNLTENQKQYLETSAACERQMSKI IRDVL ENIEDGSLTLEKEDPFLG
 SVIDAVVSQVMLLLREKGVQLIRDIPEEIKTLTVHGDQVRIQQV LADFLNMVRYAPS
 PDGWVEIQLRPSMMPISDGVTVGHIELRIICPGEGLPPELVQDMFHSSRWVTQEGLGL
 STCRKMLKLMNGEIQYIRESERCYFLIVLDLPMTRKGP KSVG

(SEQ ID NO: 10)

SNNNNNRNIKRESLSMRKAI AQYTEDAXLHAVFEKSGDSFDYAQ
 SIRVTAATESVPEQQITAYLAKIQRGGFIQPF GSMIAVDETSFRVLAYSENARDMLGI
 APQSVSMEDDSSSSSFFSLGV DVS LFSASSV LLEKAFSAREI SLMNPIWIHSRST
 GKPFYILHRIDIGVVIDLEPARSED PALS IAGAVQSQKLAVRAISQLQALPGGDV KL
 LCDAVVESVRELTGYDRVMVYKFHEDEHGEVVAESKRVDLEPYIGLHYPATDIPQASR
 FLFKQNRVRMIVDCNASPVRVPQDEALVQPVCLVGS TLRAPHGCHAQYMANMGS IASL
 AMAVI INGNDEGGGIGGAARGSMRLWGLV VCHHTSARCI PPFLRYACEFLMQAFGLQ
 LNMELQLAVQSLEKRVLKTQTLLCDMLLRDSHTGIVTQSPS IMDLVKCDGAALYYQGN
 YHPLGVTPTESQIRDIIDWLLAFHSDSTGLSTDSLADAGYPGAASLGDAVCGMAVAYI
 TEKDFLFWFRSHTAKEIKWGGAKHHPEDKDDGQKM HPRS SFKAFLEVVKIRSMQWDNA
 EMDAIHSLQLILRDSFKEAENNSKAVVHTHMAELELQGVDELSSVAREMVRLIETAT
 APIFAVDVDGRINGWNAKVSELTGLLVEEAMGKSLVHDLVYKESRETVDKLLSHALKG
 EEDKNVEIKMKTFGPGNQNKAVFIVVNACSSKDYTN NIVGVC FVGQDITGQKVVMDKF
 INIQGDYKAI VHSNPLIPPI FASDDNTCCLEWNNAMEKLSGWSRADVI GKLLVGEVF
 GSFCQLKSGDAMTKFMI VLNHNAI GGHD TDKFPFLDRHGKYVHTFLTANKRVNMDGQ
 IIGAFCFLQIVNPELQQALTVQRQDSSSLARMKELAYICQEVKNPLSGIRFTNSLLE
 STCLTDEQKQLETSVACEKQMLKIVRDIALESIEDGSL ELEKQEFLENVINAVVSQ
 VMLLLRDRKLQLIRDIPEEIKALAVYDQLRIQQV LADFLNMVRYAPSPDGWVEIHV
 FPRIKQISEGLTLLHAEFRMVCPGEGLPPELIQDMFHNSRWVTQEGLGLSMSRKIIKL
 MNGEVQYVREAEERCYFLV LLELPVTRRSKAIN

(SEQ ID NO: 11)

MSSGNRGTQSHHQASSGT SNLRVYHTDSMSKAI AQYTM DARLH
 AVYEQSGESGKSFYYSQSVRTTTSVPEQQITAYLSKIQRGGHIQPF GCM LAVDEATF

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RVIAPSENAREMLGLTPQSVPSLEKPEILLVGTDVRTLFTPSSAVLLEKAFRAREITL
 LNPVWIHSKNSGKPFYAILHRIDVGVIVDLEPARTEDPALSIAGAVQSQKLAVRAISH
 LQSLPGGDINLLCETVVENVRELTGYDRVMVYKFHEDEHGEVVAESKRSDELEPYIGLH
 YPATDIPQASRFLFRQNRVMIVDCHATPVLVIQDEGLMQPLCLVGVSTLRAPHGCHAQ
 YMANMGSTASLAMAVIINGSDEEAI GGRNLMRLWGLVVCCHTSARCI PFPLRYACEFL
 MQAFGLQLNMELQLASQLSEKHVLRQTLLCDMLLRDSPTGIVTQSPSMDLVKCDGA
 ALYYQKGYPTGVTPTEAQIKDIAEWLLANHADSTGLSTDSLADAGYPGAASLGDAVC
 GMNAVYITSRDFLWFRSHTAKEIKWGGAKHHPEDKDDQORMHPRSSFKAFLEVVKSR
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 VRLIETATAPIFAVDVDCINGWNAKVAELTGLSVEEAMGKSLVHDLVYKESEETVVK
 LLHHALRGEEDKNVEIKLRTFDSQQHKKAVFVVVNACSSRDYTNIVGVCFVQDVTG
 QKVVMDFIHIQGDYKAIVHSPNPLIPPIFASDENTVCSEWNTAMEKLTGWSRGDIIG
 KILVGEIFGSSCRLKGPDALTKFMIVLHNAIGGQDTDKPFPFFDQNGKYVQALLTAN
 KRVNIEGQIIGAFCLQIASPELQQALKVQRQEKCFARMKELAYICQEIKNPLSGI
 RFTNSLLEATDLTEDQKQFLETSAAACEKQMSKIIIRDVDLDSIEDGSLELERAEFLLS
 VINAVVSQVMILLRERDLQLIRDIPPEVKT LAVYGDQVRIQQVLADFLLMVRYAPSP
 DGWIEIQVCPRLKQISEEVKLMHIEFRMVCPGEGLPNLIQDMFHSRWMTQEGGLGS
 MCRKILKLINGEVQYIRESERCYFLISIELPIPHRGSKSVD"

(See FIG. 13)

SEQ ID NO: 12

(See FIG. 13)

SEQ ID NO: 13

(See FIG. 13)

SEQ ID NO: 14

(See FIG. 13)

SEQ ID NO: 15

(See FIG. 13)

SEQ ID NO: 16

(See FIG. 13)

SEQ ID NO: 17

(See FIG. 13)

SEQ ID NO: 18

(See FIG. 13)

SEQ ID NO: 19

(See FIG. 13)

SEQ ID NO: 20

(See FIG. 13)

SEQ ID NO: 21

(See FIG. 13)

SEQ ID NO: 22

Arabidopsis thaliana phytochrome B (PHYB) nucleotide (GenBank Accession No
 NM_127435)

(SEQ ID NO: 23)

at ggtttccgga gtcgggggta gtggcgggtg cegtggcggt
 ggccgtggcg gagaagaaga accgtcgtca agtcacactc ctaataaccg aagaggagga
 gaacaagctc aatcgctggg aacgaaatct ctacagaccaa gaagcaaac tgaatcaatg
 agcaaagcaa ttcaacagta caccgtcgac gcaagactcc acgccgtttt cgaacaatcc
 ggcgaaatcag ggaaatcatt cgactactca caatcactca aaacgacgac gtacgggtcc

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 ggtttaggtc taagcgtatg tcgaaagatt ttaaagctaa tgaacgggtga ggttcaatac
 atccgagaat cagaacggtc ctatttctc atcattctgg aactccctgt acctcgaag
 cgaccattgt caactgctag tggaaagtgt gacatgatgc tgatgatgac atat

Zea mays phytochrome B nucleotide (Phytozome Accession No. GRMZM2G124532)

(SEQ ID NO: 24)

ATGGCGTCGGGCGCCGCGCCACGCCACGCGCTCCCCCTCCTCCGCGGGCCGAGGCGCCGCGTCACGCGCACCA
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 AGGCCGTCGCCCAGTACACCTTAGACGCGCCTACACGCGGTGTCGAGCAATCGGGCGCGTCGGGCCGAGCTTC
 GACTACTCCCAATCGTCGCGCGCCGCCACGCCGCTCCTCCGAGCAGCAGATCGCGCCTACCTCTCCCGCATCCA
 GCGCGGGCGCCACATCCAGCCCTTCGGCTGCAGCTCGCGTCCGCGACGACTCCTCCTCCGCTCCTCGCTTCT
 CCGAGAACTCCCCGACTGCTCGACCTGTGCGCTCACCCTCCGTTCCCTCGCTGGACTCCTCTGCGCGCCCCAC
 GTTCCCTGGGTGCCAGCGCGCCTCCTCTCCCTCGTCCGCGTCTCCTAGAGCGCGCCTTCGCGCGCGG
 CGAGATCTCGTCTCAACCCGATATGGATCCACTCCAGGTCCTCCAAGCCGTTCTACGCCATCCTCCACCGCA
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 TCCCAGAACTGGCGTCCGCGCATCTCCGCTCCAGGCGTACCCGGCGGGGACGTCGAAGCTTCTCTGCGACAC
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 GTTACGCAAGGATGAAAGAATTGGCCATATATTGCCAGGAGATAAAGAATCCTCTTAGTGGCATCCGATTTACCAAC
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 GTCCAGGATATGTTACGAATTCCTCAATGGTCAACACAAGAAGGCGTAGGACTAAGCACATGCAGGAAGATCCTCAA
 ATGTATGGGTGGCGAGGTTCAATACATCAGAGAGTCAGAGCGGAGTTTCTTCTCATCGTCTCGAGCAGCCCCAAC
 CTCGTCCAGCAGCTGGTAGAGAAATCGTC

Oryza sativa Japonica Group isolate SJ-CDI2 phytochrome b (phyB) nucleotide
 (GenBank Accession No: JN594210)

(SEQ ID NO: 25)

atgggctcgg gtagccgcgc cagcccacg cgtccccct cctccgcgcg gcccgcgcg
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 acttgagctg ccccagcctc agcaagcagc aagtaggggg acaagc

Sorghum bicolor isolate PHYB-Rtx430 phytochrome B (PHYB) nucleotide (GenBank
 Accession No: AY466089)

(SEQ ID NO: 26)

atggcgctcg gcagccgcgc cacgcccacg cgtccccct cctccgcgcg acccgaggcg
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Glycine max phytochrome B-1 (phyB) nucleotide (GenBank: EU428749)

(SEQ ID NO: 27)

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Glycine max phytochrome B-2 (phyB) mRNA nucleotide (GenBank Accession NO: EU428750.2)

(SEQ ID NO: 28)

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 3421 gtgacacgga gaagctctaa aaagtgt

Glycine max phytochrome B-3 (phyB) nucleotide (GenBank Accession
 No: EU428751.1)

(SEQ ID NO: 29)

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Glycine max phytochrome B-4 (phyB) nucleotide
 GenBank: EU428752.1

(SEQ ID NO: 30)

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Solanum tuberosum phytochrome B nucleotide
GenBank: DQ342235.1

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Pisum sativum phytochrome B (PHYB) nucleotide
 GenBank: AF069305.1

(SEQ ID NO: 32)

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Vitis vinifera genotype PN40024 phytochrome B (PHYB) nucleotide
 GenBank: EU436650.1

(SEQ ID NO: 33)

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 35 40 45
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 50 55 60
 Ala Arg Leu His Ala Val Phe Glu Gln Ser Gly Glu Ser Gly Lys Ser
 65 70 75 80
 Phe Asp Tyr Ser Gln Ser Leu Lys Thr Thr Thr Tyr Gly Ser Ser Val
 85 90 95
 Pro Glu Gln Gln Ile Thr Ala Tyr Leu Ser Arg Ile Gln Arg Gly Gly
 100 105 110
 Tyr Ile Gln Pro Phe Gly Cys Met Ile Ala Val Asp Glu Ser Ser Phe
 115 120 125
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 Glu Ser Lys Arg Asp Asp Leu Glu Pro Tyr Ile Gly Leu His Tyr Pro
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 Ala Thr Asp Ile Pro Gln Ala Ser Arg Phe Leu Phe Lys Gln Asn Arg
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 Ala Ser Leu Ala Met Ala Val Ile Ile Asn Gly Asn Glu Asp Asp Gly
 370 375 380
 Ser Asn Val Ala Ser Gly Arg Ser Ser Met Arg Leu Trp Gly Leu Val
 385 390 395 400
 Val Cys His His Thr Ser Ser Arg Cys Ile Pro Phe Pro Leu Arg Tyr
 405 410 415
 Ala Cys Glu Phe Leu Met Gln Ala Phe Gly Leu Gln Leu Asn Met Glu
 420 425 430
 Leu Gln Leu Ala Leu Gln Met Ser Glu Lys Arg Val Leu Arg Thr Gln
 435 440 445
 Thr Leu Leu Cys Asp Met Leu Leu Arg Asp Ser Pro Ala Gly Ile Val
 450 455 460
 Thr Gln Ser Pro Ser Ile Met Asp Leu Val Lys Cys Asp Gly Ala Ala
 465 470 475 480
 Phe Leu Tyr His Gly Lys Tyr Tyr Pro Leu Gly Val Ala Pro Ser Glu
 485 490 495
 Val Gln Ile Lys Asp Val Val Glu Trp Leu Leu Ala Asn His Ala Asp
 500 505 510
 Ser Thr Gly Leu Ser Thr Asp Ser Leu Gly Asp Ala Gly Tyr Pro Gly
 515 520 525
 Ala Ala Ala Leu Gly Asp Ala Val Cys Gly Met Ala Val Ala Tyr Ile
 530 535 540
 Thr Lys Arg Asp Phe Leu Phe Trp Phe Arg Ser His Thr Ala Lys Glu
 545 550 555 560
 Ile Lys Trp Gly Gly Ala Lys His His Pro Glu Asp Lys Asp Asp Gly
 565 570 575
 Gln Arg Met His Pro Arg Ser Ser Phe Gln Ala Phe Leu Glu Val Val
 580 585 590
 Lys Ser Arg Ser Gln Pro Trp Glu Thr Ala Glu Met Asp Ala Ile His

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595					600					605					
Ser	Leu	Gln	Leu	Ile	Leu	Arg	Asp	Ser	Phe	Lys	Glu	Ser	Glu	Ala	Ala
610						615					620				
Met	Asn	Ser	Lys	Val	Val	Asp	Gly	Val	Val	Gln	Pro	Cys	Arg	Asp	Met
625				630						635					640
Ala	Gly	Glu	Gln	Gly	Ile	Asp	Glu	Leu	Gly	Ala	Val	Ala	Arg	Glu	Met
				645					650					655	
Val	Arg	Leu	Ile	Glu	Thr	Ala	Thr	Val	Pro	Ile	Phe	Ala	Val	Asp	Ala
			660					665					670		
Gly	Gly	Cys	Ile	Asn	Gly	Trp	Asn	Ala	Lys	Ile	Ala	Glu	Leu	Thr	Gly
		675					680					685			
Leu	Ser	Val	Glu	Glu	Ala	Met	Gly	Lys	Ser	Leu	Val	Ser	Asp	Leu	Ile
	690					695					700				
Tyr	Lys	Glu	Asn	Glu	Ala	Thr	Val	Asn	Lys	Leu	Leu	Ser	Arg	Ala	Leu
705				710						715					720
Arg	Gly	Asp	Glu	Glu	Lys	Asn	Val	Glu	Val	Lys	Leu	Lys	Thr	Phe	Ser
			725					730						735	
Pro	Glu	Leu	Gln	Gly	Lys	Ala	Val	Phe	Val	Val	Val	Asn	Ala	Cys	Ser
			740					745					750		
Ser	Lys	Asp	Tyr	Leu	Asn	Asn	Ile	Val	Gly	Val	Cys	Phe	Val	Gly	Gln
		755					760					765			
Asp	Val	Thr	Ser	Gln	Lys	Ile	Val	Met	Asp	Lys	Phe	Ile	Asn	Ile	Gln
	770					775					780				
Gly	Asp	Tyr	Lys	Ala	Ile	Val	His	Ser	Pro	Asn	Pro	Leu	Ile	Pro	Pro
785					790					795					800
Ile	Phe	Ala	Ala	Asp	Glu	Asn	Thr	Cys	Cys	Leu	Glu	Trp	Asn	Met	Ala
				805						810				815	
Met	Glu	Lys	Leu	Thr	Gly	Trp	Ser	Arg	Ser	Glu	Val	Ile	Gly	Lys	Met
			820					825					830		
Ile	Val	Gly	Glu	Val	Phe	Gly	Ser	Cys	Cys	Met	Leu	Lys	Gly	Pro	Asp
		835					840						845		
Ala	Leu	Thr	Lys	Phe	Met	Ile	Val	Leu	His	Asn	Ala	Ile	Gly	Gly	Gln
						855					860				
Asp	Thr	Asp	Lys	Phe	Pro	Phe	Pro	Phe	Phe	Asp	Arg	Asn	Gly	Lys	Phe
865					870					875					880
Val	Gln	Ala	Leu	Leu	Thr	Ala	Asn	Lys	Arg	Val	Ser	Leu	Glu	Gly	Lys
				885					890					895	
Val	Ile	Gly	Ala	Phe	Cys	Phe	Leu	Gln	Ile	Pro	Ser	Pro	Glu	Leu	Gln
			900					905					910		
Gln	Ala	Leu	Ala	Val	Gln	Arg	Arg	Gln	Asp	Thr	Glu	Cys	Phe	Thr	Lys
		915					920						925		
Ala	Lys	Glu	Leu	Ala	Tyr	Ile	Cys	Gln	Val	Ile	Lys	Asn	Pro	Leu	Ser
		930				935						940			
Gly	Met	Arg	Phe	Ala	Asn	Ser	Leu	Leu	Glu	Ala	Thr	Asp	Leu	Asn	Glu
945					950					955					960
Asp	Gln	Lys	Gln	Leu	Leu	Glu	Thr	Ser	Val	Ser	Cys	Glu	Lys	Gln	Ile
				965					970					975	
Ser	Arg	Ile	Val	Gly	Asp	Met	Asp	Leu	Glu	Ser	Ile	Glu	Asp	Gly	Ser
			980					985					990		
Phe	Val	Leu	Lys	Arg	Glu	Glu	Phe	Phe	Leu	Gly	Ser	Val	Ile	Asn	Ala
			995				1000						1005		
Ile	Val	Ser	Gln	Ala	Met	Phe	Leu	Leu	Arg	Asp	Arg	Gly	Leu	Gln	
	1010						1015						1020		

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Leu Ile Arg Asp Ile Pro Glu Glu Ile Lys Ser Ile Glu Val Phe
 1025 1030 1035
 Gly Asp Gln Ile Arg Ile Gln Gln Leu Leu Ala Glu Phe Leu Leu
 1040 1045 1050
 Ser Ile Ile Arg Tyr Ala Pro Ser Gln Glu Trp Val Glu Ile His
 1055 1060 1065
 Leu Ser Gln Leu Ser Lys Gln Met Ala Asp Gly Phe Ala Ala Ile
 1070 1075 1080
 Arg Thr Glu Phe Arg Met Ala Cys Pro Gly Glu Gly Leu Pro Pro
 1085 1090 1095
 Glu Leu Val Arg Asp Met Phe His Ser Ser Arg Trp Thr Ser Pro
 1100 1105 1110
 Glu Gly Leu Gly Leu Ser Val Cys Arg Lys Ile Leu Lys Leu Met
 1115 1120 1125
 Asn Gly Glu Val Gln Tyr Ile Arg Glu Ser Glu Arg Ser Tyr Phe
 1130 1135 1140
 Leu Ile Ile Leu Glu Leu Pro Val Pro Arg Lys Arg Pro Leu Ser
 1145 1150 1155
 Thr Ala Ser Gly Ser Gly Asp Met Met Leu Met Met Pro Tyr
 1160 1165 1170

<210> SEQ ID NO 2
 <211> LENGTH: 1161
 <212> TYPE: PRT
 <213> ORGANISM: Zea mays

<400> SEQUENCE: 2

Met Ala Ser Gly Ser Arg Ala Thr Pro Thr Arg Ser Pro Ser Ser Ala
 1 5 10 15
 Arg Pro Glu Ala Pro Arg His Ala His His His His His Ser Gln Ser
 20 25 30
 Ser Gly Gly Ser Thr Ser Arg Ala Gly Gly Gly Ala Ala Ala Thr Glu
 35 40 45
 Ser Val Ser Lys Ala Val Ala Gln Tyr Thr Leu Asp Ala Arg Leu His
 50 55 60
 Ala Val Phe Glu Gln Ser Gly Ala Ser Gly Arg Ser Phe Asp Tyr Ser
 65 70 75 80
 Gln Ser Leu Arg Ala Pro Pro Thr Pro Ser Ser Glu Gln Gln Ile Ala
 85 90 95
 Ala Tyr Leu Ser Arg Ile Gln Arg Gly Gly His Ile Gln Pro Phe Gly
 100 105 110
 Cys Thr Leu Ala Val Ala Asp Asp Ser Ser Phe Arg Leu Leu Ala Phe
 115 120 125
 Ser Glu Asn Ser Pro Asp Leu Leu Asp Leu Ser Pro His His Ser Val
 130 135 140
 Pro Ser Leu Asp Ser Ser Ala Pro Pro His Val Ser Leu Gly Ala Asp
 145 150 155 160
 Ala Arg Leu Leu Phe Ser Pro Ser Ser Ala Val Leu Leu Glu Arg Ala
 165 170 175
 Phe Ala Ala Arg Glu Ile Ser Leu Leu Asn Pro Ile Trp Ile His Ser
 180 185 190
 Arg Val Ser Ser Lys Pro Phe Tyr Ala Ile Leu His Arg Ile Asp Val
 195 200 205
 Gly Val Val Ile Asp Leu Glu Pro Ala Arg Thr Glu Asp Pro Ala Leu

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Glu Leu Arg Gly Ile Asn Glu Leu Ser Ser Val Ala Arg Glu Met Val
 645 650 655
 Arg Leu Ile Glu Thr Ala Thr Val Pro Ile Phe Ala Val Asp Thr Asp
 660 665 670
 Gly Cys Ile Asn Gly Trp Asn Ala Lys Ile Ala Glu Leu Thr Gly Leu
 675 680 685
 Ser Val Glu Glu Ala Met Gly Lys Ser Leu Val Asn Asp Leu Ile Phe
 690 695 700
 Lys Glu Ser Glu Ala Thr Val Glu Lys Leu Leu Ser Arg Ala Leu Arg
 705 710 715 720
 Gly Glu Glu Asp Lys Asn Val Glu Ile Lys Leu Lys Thr Phe Gly Ser
 725 730 735
 Glu Gln Tyr Lys Gly Pro Ile Phe Val Val Val Asn Ala Cys Ser Ser
 740 745 750
 Arg Asp Tyr Thr Gln Asn Ile Val Gly Val Cys Phe Val Gly Gln Asp
 755 760 765
 Val Thr Gly Gln Lys Val Val Met Asp Lys Phe Val Asn Ile Gln Gly
 770 775 780
 Asp Tyr Lys Ala Ile Val His Asn Pro Asn Pro Leu Ile Pro Pro Ile
 785 790 795 800
 Phe Ala Ser Asp Glu Asn Thr Ser Cys Ser Glu Trp Asn Thr Ala Met
 805 810 815
 Glu Lys Leu Thr Gly Trp Ser Arg Gly Glu Val Val Gly Lys Phe Leu
 820 825 830
 Ile Gly Glu Val Phe Gly Asn Cys Cys Arg Leu Lys Gly Pro Asp Ala
 835 840 845
 Leu Thr Lys Phe Met Val Ile Ile His Asn Ala Ile Gly Gly Gln Asp
 850 855 860
 Tyr Glu Lys Phe Pro Phe Ser Phe Phe Asp Lys Asn Gly Lys Tyr Val
 865 870 875 880
 Gln Ala Leu Leu Thr Ala Asn Thr Arg Ser Lys Met Asp Gly Lys Ser
 885 890 895
 Ile Gly Ala Phe Cys Phe Leu Gln Ile Ala Ser Thr Glu Ile Gln Gln
 900 905 910
 Ala Phe Glu Ile Gln Arg Gln Gln Glu Lys Lys Cys Tyr Ala Arg Met
 915 920 925
 Lys Glu Leu Ala Tyr Ile Cys Gln Glu Ile Lys Asn Pro Leu Ser Gly
 930 935 940
 Ile Arg Phe Thr Asn Ser Leu Leu Gln Met Thr Asp Leu Asn Asp Asp
 945 950 955 960
 Gln Arg Gln Phe Leu Glu Thr Ser Ser Ala Cys Glu Lys Gln Met Ser
 965 970 975
 Lys Ile Val Lys Asp Ala Ser Leu Gln Ser Ile Glu Asp Gly Ser Leu
 980 985 990
 Val Leu Glu Gln Ser Glu Phe Ser Leu Gly Asp Val Met Asn Ala Val
 995 1000 1005
 Val Ser Gln Ala Met Leu Leu Leu Arg Glu Arg Asp Leu Gln Leu
 1010 1015 1020
 Ile Arg Asp Ile Pro Asp Glu Ile Lys Asp Ala Ser Ala Tyr Gly
 1025 1030 1035
 Asp Gln Cys Arg Ile Gln Gln Val Leu Ala Asp Phe Leu Leu Ser
 1040 1045 1050

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Met Val Arg Ser Ala Pro Ser Glu Asn Gly Trp Val Glu Ile Gln
1055 1060 1065

Val Arg Pro Asn Val Lys Gln Asn Ser Asp Gly Thr Asn Thr Glu
1070 1075 1080

Leu Phe Ile Phe Arg Phe Ala Cys Pro Gly Glu Gly Leu Pro Ala
1085 1090 1095

Asp Val Val Gln Asp Met Phe Ser Asn Ser Gln Trp Ser Thr Gln
1100 1105 1110

Glu Gly Val Gly Leu Ser Thr Cys Arg Lys Ile Leu Lys Leu Met
1115 1120 1125

Gly Gly Glu Val Gln Tyr Ile Arg Glu Ser Glu Arg Ser Phe Phe
1130 1135 1140

Leu Ile Val Leu Glu Gln Pro Gln Pro Arg Pro Ala Ala Gly Arg
1145 1150 1155

Glu Ile Val
1160

<210> SEQ ID NO 3
 <211> LENGTH: 1171
 <212> TYPE: PRT
 <213> ORGANISM: *Oryza sativa*

<400> SEQUENCE: 3

Met Gly Ser Gly Ser Arg Ala Thr Pro Thr Arg Ser Pro Ser Ser Ala
1 5 10 15

Arg Pro Ala Ala Pro Arg His Gln His His His Ser Gln Ser Ser Gly
20 25 30

Gly Ser Thr Ser Arg Ala Gly Gly Gly Gly Gly Gly Gly Gly Gly
35 40 45

Gly Gly Gly Ala Ala Ala Ala Glu Ser Val Ser Lys Ala Val Ala Gln
50 55 60

Tyr Thr Leu Asp Ala Arg Leu His Ala Val Phe Glu Gln Ser Gly Ala
65 70 75 80

Ser Gly Arg Ser Phe Asp Tyr Thr Gln Ser Leu Arg Ala Ser Pro Thr
85 90 95

Pro Ser Ser Glu Gln Gln Ile Ala Ala Tyr Leu Ser Arg Ile Gln Arg
100 105 110

Gly Gly His Ile Gln Pro Phe Gly Cys Thr Leu Ala Val Ala Asp Asp
115 120 125

Ser Ser Phe Arg Leu Leu Ala Tyr Ser Glu Asn Thr Ala Asp Leu Leu
130 135 140

Asp Leu Ser Pro His His Ser Val Pro Ser Leu Asp Ser Ser Ala Val
145 150 155 160

Pro Pro Pro Val Ser Leu Gly Ala Asp Ala Arg Leu Leu Phe Ala Pro
165 170 175

Ser Ser Ala Val Leu Leu Glu Arg Ala Phe Ala Ala Arg Glu Ile Ser
180 185 190

Leu Leu Asn Pro Leu Trp Ile His Ser Arg Val Ser Ser Lys Pro Phe
195 200 205

Tyr Ala Ile Leu His Arg Ile Asp Val Gly Val Val Ile Asp Leu Glu
210 215 220

Pro Ala Arg Thr Glu Asp Pro Ala Leu Ser Ile Ala Gly Ala Val Gln
225 230 235 240

Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Arg Leu Gln Ala Leu Pro
245 250 255

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Gly Gly Asp Val Lys Leu Leu Cys Asp Thr Val Val Glu His Val Arg
260 265 270

Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Arg Phe His Glu Asp
275 280 285

Glu His Gly Glu Val Val Ala Glu Ser Arg Arg Ser Asn Leu Glu Pro
290 295 300

Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
305 310 315 320

Phe Leu Phe Arg Gln Asn Arg Val Arg Met Ile Ala Asp Cys His Ala
325 330 335

Ala Pro Val Arg Val Ile Gln Asp Pro Ala Leu Thr Gln Pro Leu Cys
340 345 350

Leu Val Gly Ser Thr Leu Arg Ser Pro His Gly Cys His Ala Gln Tyr
355 360 365

Met Ala Asn Met Gly Ser Ile Ala Ser Leu Val Met Ala Val Ile Ile
370 375 380

Ser Ser Gly Gly Asp Asp Asp His Asn Ile Ala Arg Gly Ser Ile Pro
385 390 395 400

Ser Ala Met Lys Leu Trp Gly Leu Val Val Cys His His Thr Ser Pro
405 410 415

Arg Cys Ile Pro Phe Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln
420 425 430

Ala Phe Gly Leu Gln Leu Asn Met Glu Leu Gln Leu Ala His Gln Leu
435 440 445

Ser Glu Lys His Ile Leu Arg Thr Gln Thr Leu Leu Cys Asp Met Leu
450 455 460

Leu Arg Asp Ser Pro Thr Gly Ile Val Thr Gln Ser Pro Ser Ile Met
465 470 475 480

Asp Leu Val Lys Cys Asp Gly Ala Ala Leu Tyr Tyr His Gly Lys Tyr
485 490 495

Tyr Pro Leu Gly Val Thr Pro Thr Glu Val Gln Ile Lys Asp Ile Ile
500 505 510

Glu Trp Leu Thr Met Cys His Gly Asp Ser Thr Gly Leu Ser Thr Asp
515 520 525

Ser Leu Ala Asp Ala Gly Tyr Ser Gly Ala Ala Ala Leu Gly Asp Ala
530 535 540

Val Ser Gly Met Ala Val Ala Tyr Ile Thr Pro Ser Asp Tyr Leu Phe
545 550 555 560

Trp Phe Arg Ser His Thr Ala Lys Glu Ile Lys Trp Gly Gly Ala Lys
565 570 575

His His Pro Glu Asp Lys Asp Asp Gly Gln Arg Met His Pro Arg Ser
580 585 590

Ser Phe Lys Ala Phe Leu Glu Val Val Lys Ser Arg Ser Leu Pro Trp
595 600 605

Glu Asn Ala Glu Met Asp Ala Ile His Ser Leu Gln Leu Ile Leu Arg
610 615 620

Asp Ser Phe Arg Asp Ser Ala Glu Gly Thr Ser Asn Ser Lys Ala Ile
625 630 635 640

Val Asn Gly Gln Val Gln Leu Gly Glu Leu Glu Leu Arg Gly Ile Asp
645 650 655

Glu Leu Ser Ser Val Ala Arg Glu Met Val Arg Leu Ile Glu Thr Ala
660 665 670

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Thr Val Pro Ile Phe Ala Val Asp Thr Asp Gly Cys Ile Asn Gly Trp
 675 680 685
 Asn Ala Lys Val Ala Glu Leu Thr Gly Leu Ser Val Glu Glu Ala Met
 690 695 700
 Gly Lys Ser Leu Val Asn Asp Leu Ile Phe Lys Glu Ser Glu Glu Thr
 705 710 715 720
 Val Asn Lys Leu Leu Ser Arg Ala Leu Arg Gly Asp Glu Asp Lys Asn
 725 730 735
 Val Glu Ile Lys Leu Lys Thr Phe Gly Pro Glu Gln Ser Lys Gly Pro
 740 745 750
 Ile Phe Val Ile Val Asn Ala Cys Ser Ser Arg Asp Tyr Thr Lys Asn
 755 760 765
 Ile Val Gly Val Cys Phe Val Gly Gln Asp Val Thr Gly Gln Lys Val
 770 775 780
 Val Met Asp Lys Phe Ile Asn Ile Gln Gly Asp Tyr Lys Ala Ile Val
 785 790 795 800
 His Asn Pro Asn Pro Leu Ile Pro Pro Ile Phe Ala Ser Asp Glu Asn
 805 810 815
 Thr Cys Cys Leu Glu Trp Asn Thr Ala Met Glu Lys Leu Thr Gly Trp
 820 825 830
 Ser Arg Gly Glu Val Val Gly Lys Leu Leu Val Gly Glu Val Phe Gly
 835 840 845
 Asn Cys Cys Arg Leu Lys Gly Pro Asp Ala Leu Thr Lys Phe Met Ile
 850 855 860
 Val Leu His Asn Ala Ile Gly Gly Gln Asp Cys Glu Lys Phe Pro Phe
 865 870 875 880
 Ser Phe Phe Asp Lys Asn Gly Lys Tyr Val Gln Ala Leu Leu Thr Ala
 885 890 895
 Asn Thr Arg Ser Arg Met Asp Gly Glu Ala Ile Gly Ala Phe Cys Phe
 900 905 910
 Leu Gln Ile Ala Ser Pro Glu Leu Gln Gln Ala Phe Glu Ile Gln Arg
 915 920 925
 His His Glu Lys Lys Cys Tyr Ala Arg Met Lys Glu Leu Ala Tyr Ile
 930 935 940
 Tyr Gln Glu Ile Lys Asn Pro Leu Asn Gly Ile Arg Phe Thr Asn Ser
 945 950 955 960
 Leu Leu Glu Met Thr Asp Leu Lys Asp Asp Gln Arg Gln Phe Leu Glu
 965 970 975
 Thr Ser Thr Ala Cys Glu Lys Gln Met Ser Lys Ile Val Lys Asp Ala
 980 985 990
 Ser Leu Gln Ser Ile Glu Asp Gly Ser Leu Val Leu Glu Lys Gly Glu
 995 1000 1005
 Phe Ser Leu Gly Ser Val Met Asn Ala Val Val Ser Glu Val Met
 1010 1015 1020
 Ile Gln Leu Arg Glu Arg Asp Leu Gln Leu Ile Arg Asp Ile Pro
 1025 1030 1035
 Asp Glu Ile Lys Glu Ala Ser Ala Tyr Gly Asp Gln Tyr Arg Ile
 1040 1045 1050
 Gln Gln Val Leu Cys Asp Phe Leu Leu Ser Met Val Arg Phe Ala
 1055 1060 1065
 Pro Ala Glu Asn Gly Trp Val Glu Ile Gln Val Arg Pro Asn Ile
 1070 1075 1080
 Lys Gln Asn Ser Asp Gly Thr Asp Thr Met Leu Phe Leu Phe Arg

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1085	1090	1095
Phe Ala Cys Pro Gly Glu Gly Leu Pro Pro Glu Ile Val Gln Asp 1100 1105 1110		
Met Phe Ser Asn Ser Arg Trp Thr Thr Gln Glu Gly Ile Gly Leu 1115 1120 1125		
Ser Ile Cys Arg Lys Ile Leu Lys Leu Met Gly Gly Glu Val Gln 1130 1135 1140		
Tyr Ile Arg Glu Ser Glu Arg Ser Phe Phe His Ile Val Leu Glu 1145 1150 1155		
Leu Pro Gln Pro Gln Gln Ala Ala Ser Arg Gly Thr Ser 1160 1165 1170		

<210> SEQ ID NO 4

<211> LENGTH: 1178

<212> TYPE: PRT

<213> ORGANISM: Sorghum bicolor

<400> SEQUENCE: 4

Met Ala Ser Gly Ser Arg Ala Thr Pro Thr Arg Ser Pro Ser Ser Ala 1 5 10 15
Arg Pro Glu Ala Pro Arg His Ala His His His His His His His Ser 20 25 30
Gln Ser Ser Gly Gly Ser Thr Ser Arg Ala Gly Gly Gly Gly Gly Gly 35 40 45
Gly Gly Gly Gly Gly Gly Thr Ala Ala Thr Ala Thr Ala Thr Ala Thr 50 55 60
Glu Ser Val Ser Lys Ala Val Ala Gln Tyr Thr Leu Asp Ala Arg Leu 65 70 75 80
His Ala Val Phe Glu Gln Ser Gly Ala Ser Gly Arg Ser Phe Asp Tyr 85 90 95
Ser Gln Ser Leu Arg Ala Pro Pro Thr Pro Ser Ser Glu Gln Gln Ile 100 105 110
Ala Ala Tyr Leu Ser Arg Ile Gln Arg Gly Gly His Ile Gln Pro Phe 115 120 125
Gly Cys Thr Leu Ala Val Ala Asp Asp Ser Ser Phe Arg Leu Leu Ala 130 135 140
Phe Ser Glu Asn Ala Ala Asp Leu Leu Asp Leu Ser Pro His His Ser 145 150 155 160
Val Pro Ser Leu Asp Ser Ala Ala Pro Pro Pro Val Ser Leu Gly Ala 165 170 175
Asp Ala Arg Leu Leu Phe Ser Pro Ser Ser Ala Val Leu Leu Glu Arg 180 185 190
Ala Phe Ala Ala Arg Glu Ile Ser Leu Leu Asn Pro Leu Trp Ile His 195 200 205
Ser Arg Val Ser Ser Lys Pro Phe Tyr Ala Ile Leu His Arg Ile Asp 210 215 220
Val Gly Val Val Ile Asp Leu Glu Pro Ala Arg Thr Glu Asp Pro Ala 225 230 235 240
Leu Ser Ile Ala Gly Ala Val Gln Ser Gln Lys Leu Ala Val Arg Ala 245 250 255
Ile Ser Arg Leu Gln Ala Leu Pro Gly Gly Asp Ile Lys Leu Leu Cys 260 265 270
Asp Thr Val Val Glu His Val Arg Glu Leu Thr Gly Tyr Asp Arg Val 275 280 285

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Met Val Tyr Arg Phe His Glu Asp Glu His Gly Glu Val Val Ala Glu
 290 295 300
 Ser Arg Arg Asp Asn Leu Glu Pro Tyr Leu Gly Leu His Tyr Pro Ala
 305 310 315 320
 Thr Asp Ile Pro Gln Ala Ser Arg Phe Leu Phe Arg Gln Asn Arg Val
 325 330 335
 Arg Met Ile Ala Asp Cys His Ala Thr Pro Val Arg Val Ile Gln Asp
 340 345 350
 Pro Gly Met Ser Gln Pro Leu Cys Leu Val Gly Ser Thr Leu Arg Ala
 355 360 365
 Pro His Gly Cys His Ala Gln Tyr Met Ala Asn Met Gly Ser Ile Ala
 370 375 380
 Ser Leu Val Met Ala Val Ile Ile Ser Ser Gly Gly Asp Asp Glu Gln
 385 390 395 400
 Thr Gly Arg Gly Gly Ile Ser Ser Ala Met Lys Leu Trp Gly Leu Val
 405 410 415
 Val Cys His His Thr Ser Pro Arg Cys Ile Pro Phe Pro Leu Arg Tyr
 420 425 430
 Ala Cys Glu Phe Leu Met Gln Ala Phe Gly Leu Gln Leu Asn Met Glu
 435 440 445
 Leu Gln Leu Ala His Gln Leu Ser Glu Lys His Ile Leu Arg Thr Gln
 450 455 460
 Thr Leu Leu Cys Asp Met Leu Leu Arg Asp Ser Pro Thr Gly Ile Val
 465 470 475 480
 Thr Gln Ser Pro Ser Ile Met Asp Leu Val Lys Cys Asp Gly Ala Ala
 485 490 495
 Leu Tyr Tyr His Gly Lys Tyr Tyr Pro Leu Gly Val Thr Pro Thr Glu
 500 505 510
 Ser Gln Ile Lys Asp Ile Ile Glu Trp Leu Thr Val Cys His Gly Asp
 515 520 525
 Ser Thr Gly Leu Ser Thr Asp Ser Leu Ala Asp Ala Gly Tyr Leu Gly
 530 535 540
 Ala Ala Ala Leu Gly Asp Ala Val Cys Gly Met Ala Val Ala Tyr Ile
 545 550 555 560
 Thr Pro Ser Asp Tyr Leu Phe Trp Phe Arg Ser His Thr Ala Lys Glu
 565 570 575
 Ile Lys Trp Gly Gly Ala Lys His His Pro Glu Asp Lys Asp Asp Gly
 580 585 590
 Gln Arg Met His Pro Arg Ser Ser Phe Lys Ala Phe Leu Glu Val Val
 595 600 605
 Lys Ser Arg Ser Leu Pro Trp Glu Asn Ala Glu Met Asp Ala Ile His
 610 615 620
 Ser Leu Gln Leu Ile Leu Arg Asp Ser Phe Arg Asp Ala Ala Glu Gly
 625 630 635 640
 Thr Ser Asn Ser Lys Ala Ile Val Asn Gly Gln Ala Gln Leu Gly Glu
 645 650 655
 Leu Glu Leu Arg Gly Ile Asn Glu Leu Ser Ser Val Pro Arg Glu Met
 660 665 670
 Val Arg Leu Ile Glu Thr Ala Thr Val Pro Ile Phe Ala Val Asp Thr
 675 680 685
 Asp Gly Cys Ile Asn Gly Trp Asn Ala Lys Ile Ala Glu Leu Thr Gly
 690 695 700
 Leu Ser Val Glu Glu Ala Met Gly Lys Ser Leu Val Asn Asp Leu Ile

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705	710	715	720
Phe Lys Glu Ser Glu Glu Ile Val Glu Lys Leu Leu Ser Arg Ala Leu	725	730	735
Arg Gly Glu Glu Asp Lys Asn Val Glu Ile Lys Leu Lys Thr Phe Gly	740	745	750
Ser Glu Gln Ser Asn Gly Ala Ile Phe Val Ile Val Asn Ala Cys Ser	755	760	765
Ser Arg Asp Tyr Thr Gln Asn Ile Val Gly Val Cys Phe Val Gly Gln	770	775	780
Asp Val Thr Gly Gln Lys Val Val Met Asp Lys Phe Ile Asn Ile Gln	785	790	800
Gly Asp Tyr Lys Ala Ile Val His Asn Pro Asn Pro Leu Ile Pro Pro	805	810	815
Ile Phe Ala Ser Asp Glu Asn Thr Ser Cys Ser Glu Trp Asn Thr Ala	820	825	830
Met Glu Lys Leu Thr Gly Trp Ser Arg Gly Glu Val Val Gly Lys Phe	835	840	845
Leu Ile Gly Glu Val Phe Gly Ser Phe Cys Arg Leu Lys Gly Pro Asp	850	855	860
Ala Leu Thr Lys Phe Met Val Val Ile His Asn Ala Ile Gly Gly Gln	865	870	875
Asp Tyr Glu Lys Phe Pro Phe Ser Phe Phe Asp Lys Asn Gly Lys Tyr	885	890	895
Val Gln Ala Leu Leu Thr Ala Asn Thr Arg Ser Lys Met Asp Gly Lys	900	905	910
Ser Ile Gly Ala Phe Cys Phe Leu Gln Ile Ala Ser Ala Glu Ile Gln	915	920	925
Gln Ala Phe Glu Ile Gln Arg Gln Gln Glu Lys Lys Cys Tyr Ala Arg	930	935	940
Met Lys Glu Leu Ala Tyr Ile Cys Gln Glu Ile Lys Asn Pro Leu Ser	945	950	955
Gly Ile Arg Phe Thr Asn Ser Leu Leu Gln Met Thr Asp Leu Asn Asp	965	970	975
Asp Gln Arg Gln Phe Leu Glu Thr Cys Ser Ala Cys Glu Lys Gln Met	980	985	990
Ser Lys Ile Val Lys Asp Ala Thr Leu Gln Ser Ile Glu Asp Gly Ser	995	1000	1005
Leu Val Leu Glu Lys Ser Glu Phe Ser Phe Gly Asp Val Met Asn	1010	1015	1020
Ala Val Val Ser Gln Ala Met Leu Leu Leu Arg Glu Arg Asp Leu	1025	1030	1035
Gln Leu Ile Arg Asp Ile Pro Asp Glu Ile Lys Asp Ala Ser Ala	1040	1045	1050
Tyr Gly Asp Gln Phe Arg Ile Gln Gln Val Leu Ala Asp Phe Leu	1055	1060	1065
Leu Ser Met Val Arg Ser Ala Pro Ser Glu Asn Gly Trp Val Glu	1070	1075	1080
Ile Gln Val Arg Pro Asn Val Lys Gln Asn Ser Asp Gly Thr Asp	1085	1090	1095
Thr Glu Leu Phe Ile Phe Arg Phe Ala Cys Pro Gly Glu Gly Leu	1100	1105	1110
Pro Ala Asp Ile Val Gln Asp Met Phe Ser Asn Ser Gln Trp Ser	1115	1120	1125

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Thr Gln Glu Gly Val Gly Leu Ser Thr Cys Arg Lys Ile Leu Lys
 1130 1135 1140
 Leu Met Gly Gly Glu Val Gln Tyr Ile Arg Glu Ser Glu Arg Ser
 1145 1150 1155
 Phe Phe Leu Ile Val Leu Glu Leu Pro Gln Pro Arg Pro Ala Ala
 1160 1165 1170
 Asp Arg Glu Ile Ser
 1175

 <210> SEQ ID NO 5
 <211> LENGTH: 1137
 <212> TYPE: PRT
 <213> ORGANISM: Glycine max

 <400> SEQUENCE: 5

 Met Ala Ser Ala Ser Gly Ala Ala Asn Ser Ser Val Pro Pro Pro Gln
 1 5 10 15
 Ile His Thr Ser Arg Thr Lys Leu Ser His His Ser Ser Asn Asn Asn
 20 25 30
 Asn Asn Ile Asp Ser Met Ser Lys Ala Ile Ala Gln Tyr Thr Glu Asp
 35 40 45
 Ala Arg Leu His Ala Val Phe Glu Gln Ser Gly Glu Ser Gly Arg Ser
 50 55 60
 Phe Asn Tyr Ser Glu Ser Ile Arg Ile Ala Ser Glu Ser Val Pro Glu
 65 70 75 80
 Gln Gln Ile Thr Ala Tyr Leu Val Lys Ile Gln Arg Gly Gly Phe Ile
 85 90 95
 Gln Pro Phe Gly Ser Met Ile Ala Val Asp Glu Pro Ser Phe Arg Ile
 100 105 110
 Leu Gly Tyr Ser Asp Asn Ala Arg Asp Met Leu Gly Ile Thr Pro Gln
 115 120 125
 Ser Val Pro Ser Leu Asp Asp Lys Asn Asp Ala Ala Phe Ala Leu Gly
 130 135 140
 Thr Asp Val Arg Ala Leu Phe Thr His Ser Ser Ala Leu Leu Leu Glu
 145 150 155 160
 Lys Ala Phe Ser Ala Arg Glu Ile Ser Leu Met Asn Pro Ile Trp Ile
 165 170 175
 His Ser Arg Thr Ser Gly Lys Pro Phe Tyr Gly Ile Leu His Arg Ile
 180 185 190
 Asp Val Gly Ile Val Ile Asp Leu Glu Pro Ala Arg Thr Glu Asp Pro
 195 200 205
 Ala Leu Ser Ile Ala Gly Ala Val Gln Ser Gln Lys Leu Ala Val Arg
 210 215 220
 Ala Ile Ser Gln Leu Gln Ser Leu Pro Gly Gly Asp Val Lys Leu Leu
 225 230 235 240
 Cys Asp Thr Val Val Glu Ser Val Arg Glu Leu Thr Gly Tyr Asp Arg
 245 250 255
 Val Met Val Tyr Lys Phe His Glu Asp Glu His Gly Glu Val Val Ser
 260 265 270
 Glu Ser Lys Arg Pro Asp Leu Glu Pro Tyr Ile Gly Leu His Tyr Pro
 275 280 285
 Ala Thr Asp Ile Pro Gln Ala Ser Arg Phe Leu Phe Lys Gln Asn Arg
 290 295 300
 Val Arg Met Ile Val Asp Cys His Ala Ser Ala Val Arg Val Val Gln

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305	310	315	320
Asp Glu Ala Leu Val Gln Pro Leu Cys Leu Val Gly Ser Thr Leu Arg	325	330	335
Ala Pro His Gly Cys His Ala Gln Tyr Met Ala Asn Met Gly Ser Ile	340	345	350
Ala Ser Leu Val Met Ala Val Ile Ile Asn Gly Asn Asp Glu Glu Gly	355	360	365
Val Gly Gly Arg Ser Ser Met Arg Leu Trp Gly Leu Val Val Cys His	370	375	380
His Thr Ser Ala Arg Cys Ile Pro Phe Pro Leu Arg Tyr Ala Cys Glu	385	390	395
Phe Leu Met Gln Ala Phe Gly Leu Gln Leu Asn Met Glu Leu Gln Leu	405	410	415
Ala Ala Gln Ser Leu Glu Lys Arg Val Leu Arg Thr Gln Thr Leu Leu	420	425	430
Cys Asp Met Leu Leu Arg Asp Ser Pro Thr Gly Ile Val Thr Gln Ser	435	440	445
Pro Ser Ile Met Asp Leu Val Lys Cys Asp Gly Ala Ala Leu Tyr Phe	450	455	460
Gln Gly Asn Tyr Tyr Pro Leu Gly Val Thr Pro Thr Glu Ala Gln Ile	465	470	475
Arg Asp Ile Ile Glu Trp Leu Leu Ala Phe His Gly Asp Ser Thr Gly	485	490	495
Leu Ser Thr Asp Ser Leu Gly Asp Ala Gly Tyr Pro Gly Ala Ala Ser	500	505	510
Leu Gly Asp Ala Val Cys Gly Met Ala Val Ala Tyr Ile Thr Glu Lys	515	520	525
Asp Phe Leu Phe Trp Phe Arg Ser His Thr Ala Lys Glu Ile Lys Trp	530	535	540
Gly Gly Ala Lys His His Pro Glu Asp Lys Asp Asp Gly Gln Arg Met	545	550	555
His Pro Arg Ser Ser Phe Lys Ala Phe Leu Glu Val Val Lys Ser Arg	565	570	575
Ser Leu Pro Trp Glu Asn Ala Glu Met Asp Ala Ile His Ser Leu Gln	580	585	590
Leu Ile Leu Arg Asp Ser Phe Lys Asp Ala Glu His Arg Asn Ser Lys	595	600	605
Ala Val Val Asp Pro His Val Ser Glu Gln Glu Leu Gln Gly Val Asp	610	615	620
Glu Leu Ser Ser Val Ala Arg Glu Met Val Arg Leu Ile Glu Thr Ala	625	630	635
Thr Ala Pro Ile Phe Ala Val Asp Val Asp Gly His Val Asn Gly Trp	645	650	655
Asn Ala Lys Val Ser Glu Leu Thr Gly Leu Pro Val Glu Glu Ala Met	660	665	670
Gly Lys Ser Leu Val His Asp Leu Val Phe Lys Glu Ser Glu Glu Thr	675	680	685
Met Asn Lys Leu Leu Ser Arg Ala Leu Lys Gly Glu Glu Asp Lys Asn	690	695	700
Val Glu Ile Lys Met Arg Thr Phe Gly Pro Glu His Gln Asn Lys Ala	705	710	715
Val Phe Leu Val Val Asn Ala Cys Ser Ser Lys Asp Phe Thr Asn Asn	725	730	735

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Val Val Gly Val Cys Phe Val Gly Gln Asp Val Thr Gly Gln Lys Ile
 740 745 750

Val Met Asp Lys Phe Ile Asn Ile Gln Gly Asp Tyr Lys Ala Ile Val
 755 760 765

His Ser Pro Asn Pro Leu Ile Pro Pro Ile Phe Ala Ser Asp Asp Asn
 770 775 780

Thr Cys Cys Leu Glu Trp Asn Thr Ala Met Glu Lys Leu Thr Gly Trp
 785 790 795 800

Gly Arg Val Asp Val Ile Gly Lys Met Leu Val Gly Glu Val Phe Gly
 805 810 815

Ser Cys Cys Gln Leu Lys Gly Ser Asp Ser Ile Thr Lys Phe Met Ile
 820 825 830

Val Leu His Asn Ala Leu Gly Gly Gln Asp Thr Asp Lys Phe Pro Phe
 835 840 845

Ser Phe Leu Asp Arg His Gly Lys Tyr Val Gln Thr Phe Leu Thr Ala
 850 855 860

Asn Lys Arg Val Asn Met Glu Gly Gln Ile Ile Gly Ala Phe Cys Phe
 865 870 875 880

Leu Gln Ile Met Ser Pro Glu Leu Gln Gln Ala Leu Lys Ala Gln Arg
 885 890 895

Gln Gln Glu Lys Asn Ser Phe Gly Arg Met Lys Glu Leu Ala Tyr Ile
 900 905 910

Cys Gln Gly Val Lys Asn Pro Leu Ser Gly Ile Arg Phe Thr Asn Ser
 915 920 925

Leu Leu Glu Ala Thr Ser Leu Thr Asn Glu Gln Lys Gln Phe Leu Glu
 930 935 940

Thr Ser Val Ala Cys Glu Lys Gln Met Leu Lys Ile Ile Arg Asp Val
 945 950 955 960

Asp Leu Glu Ser Ile Glu Asp Gly Ser Leu Glu Leu Glu Lys Gly Glu
 965 970 975

Phe Leu Leu Gly Asn Val Ile Asn Ala Val Val Ser Gln Val Met Leu
 980 985 990

Leu Leu Arg Glu Arg Asn Leu Gln Leu Ile Arg Asp Ile Pro Glu Glu
 995 1000 1005

Ile Lys Thr Leu Ala Val Tyr Gly Asp Gln Leu Arg Ile Gln Gln
 1010 1015 1020

Val Leu Ser Asp Phe Leu Leu Asn Ile Val Arg Tyr Ala Pro Ser
 1025 1030 1035

Pro Asp Gly Trp Val Glu Ile His Val Arg Pro Arg Ile Lys Gln
 1040 1045 1050

Ile Ser Asp Gly Leu Thr Leu Leu His Ala Glu Phe Arg Met Val
 1055 1060 1065

Cys Pro Gly Glu Gly Leu Pro Pro Glu Leu Ile Gln Asp Met Phe
 1070 1075 1080

Asn Asn Ser Arg Trp Gly Thr Gln Glu Gly Leu Gly Leu Ser Met
 1085 1090 1095

Ser Arg Lys Ile Leu Lys Leu Met Asn Gly Glu Val Gln Tyr Ile
 1100 1105 1110

Arg Glu Ala Glu Arg Cys Tyr Phe Tyr Val Leu Leu Glu Leu Pro
 1115 1120 1125

Val Thr Arg Arg Ser Ser Lys Lys Cys
 1130 1135

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<210> SEQ ID NO 6
<211> LENGTH: 1149
<212> TYPE: PRT
<213> ORGANISM: Glycine max

<400> SEQUENCE: 6

Met Ala Ser Ala Ser Gly Ala Glu Asn Ser Ser Val Pro Pro Ser Pro
1          5          10          15

Leu Pro Pro Pro Pro Pro Pro Gln Ile His Thr Ser Arg Thr Lys Leu
20          25          30

Ser His His His His Asn Asn Asn Asn Asn Asn Asn Asn Ile Asp
35          40          45

Ser Thr Ser Lys Ala Ile Ala Gln Tyr Thr Glu Asp Ala Arg Leu His
50          55          60

Ala Val Phe Glu Gln Ser Gly Glu Ser Gly Arg Ser Phe Asp Tyr Ser
65          70          75          80

Gln Ser Ile Arg Val Thr Ser Glu Ser Val Pro Glu Gln Gln Ile Thr
85          90          95

Ala Tyr Leu Leu Lys Ile Gln Arg Gly Gly Phe Ile Gln Pro Phe Gly
100         105         110

Ser Met Ile Ala Val Asp Glu Pro Ser Phe Arg Ile Leu Ala Tyr Ser
115        120        125

Asp Asn Ala Arg Asp Met Leu Gly Ile Thr Pro Gln Ser Val Pro Ser
130        135        140

Leu Asp Asp Lys Asn Asp Ala Ala Phe Ala Leu Gly Thr Asp Ile Arg
145        150        155        160

Thr Leu Phe Thr His Ser Ser Ala Val Leu Leu Glu Lys Ala Phe Ser
165        170        175

Ala Arg Glu Ile Ser Leu Met Asn Pro Ile Trp Ile His Ser Arg Thr
180        185        190

Ser Gly Lys Pro Phe Tyr Gly Ile Leu His Arg Ile Asp Val Gly Ile
195        200        205

Val Ile Asp Leu Glu Pro Ala Arg Thr Glu Asp Pro Ala Leu Ser Ile
210        215        220

Ala Gly Ala Val Gln Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Gln
225        230        235        240

Leu Gln Ser Leu Pro Gly Gly Asp Val Lys Leu Leu Cys Asp Thr Val
245        250        255

Val Glu Ser Val Arg Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr
260        265        270

Arg Phe His Glu Asp Glu His Gly Glu Val Val Ala Glu Thr Lys Arg
275        280        285

Pro Asp Leu Glu Pro Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile
290        295        300

Pro Gln Ala Ser Arg Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile
305        310        315        320

Val Asp Cys His Ala Ser Ala Val Arg Val Val Gln Asp Glu Ala Leu
325        330        335

Val Gln Pro Leu Cys Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly
340        345        350

Cys His Ala Gln Tyr Met Ala Asn Met Gly Ser Thr Ala Ser Leu Val
355        360        365

Met Ala Val Ile Ile Asn Gly Asn Asp Glu Glu Gly Val Gly Gly Arg
370        375        380

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Thr Ser Met Arg Leu Trp Gly Leu Val Ile Cys His His Thr Ser Ala
 385 390 395 400
 Arg Cys Ile Pro Phe Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln
 405 410 415
 Ala Phe Gly Leu Gln Leu Asn Met Glu Leu Gln Leu Ala Ala Gln Ser
 420 425 430
 Leu Glu Lys Arg Val Leu Arg Thr Gln Thr Leu Leu Cys Asp Met Leu
 435 440 445
 Leu Arg Asp Ser Pro Thr Gly Ile Val Thr Gln Ser Pro Ser Ile Met
 450 455 460
 Asp Leu Val Lys Cys Asp Gly Ala Ala Leu Tyr Tyr Gln Gly Asn Tyr
 465 470 475 480
 Tyr Pro Leu Gly Val Thr Pro Thr Glu Ala Gln Ile Arg Asp Ile Ile
 485 490 495
 Glu Trp Leu Leu Ala Phe His Arg Asp Ser Thr Gly Leu Ser Thr Asp
 500 505 510
 Ser Leu Ala Asp Ala Gly Tyr Pro Gly Ala Ala Ser Leu Gly Asp Ala
 515 520 525
 Val Cys Gly Met Ala Val Ala Tyr Ile Thr Glu Lys Asp Phe Leu Phe
 530 535 540
 Trp Phe Arg Ser His Thr Ala Lys Glu Ile Lys Trp Gly Gly Ala Lys
 545 550 555 560
 His His Pro Glu Asp Lys Asp Asp Gly Gln Arg Met His Pro Arg Ser
 565 570 575
 Ser Phe Lys Ala Phe Leu Glu Val Val Lys Ser Arg Ser Leu Pro Trp
 580 585 590
 Glu Asn Ala Glu Met Asp Ala Ile His Ser Leu Gln Leu Ile Leu Arg
 595 600 605
 Asp Ser Phe Lys Asp Ala Glu His Ser Asn Ser Lys Ala Val Leu Asp
 610 615 620
 Pro Arg Met Ser Glu Leu Glu Leu Gln Gly Val Asp Glu Leu Ser Ser
 625 630 635 640
 Val Ala Arg Glu Met Val Arg Leu Ile Glu Thr Ala Thr Ala Pro Ile
 645 650 655
 Phe Ala Val Asp Val Asp Gly Arg Ile Asn Gly Trp Asn Ala Lys Val
 660 665 670
 Ser Glu Leu Thr Gly Leu Pro Val Glu Glu Ala Met Gly Lys Ser Leu
 675 680 685
 Val Arg Asp Leu Val Phe Lys Glu Ser Glu Glu Thr Val Asp Lys Leu
 690 695 700
 Leu Ser Arg Ala Leu Lys Gly Glu Glu Asp Lys Asn Val Glu Ile Lys
 705 710 715 720
 Met Arg Thr Phe Gly Pro Glu His Gln Asn Lys Ala Val Phe Val Val
 725 730 735
 Val Asn Ala Cys Ser Ser Lys Asp Tyr Thr Asn Asn Val Val Gly Val
 740 745 750
 Cys Phe Val Gly Gln Asp Val Thr Gly Gln Lys Ile Val Met Asp Lys
 755 760 765
 Phe Ile Asn Ile Gln Gly Asp Tyr Lys Ala Ile Val His Asn Pro Asn
 770 775 780
 Pro Leu Ile Pro Pro Ile Phe Ala Ser Asp Asp Asn Thr Cys Cys Leu
 785 790 795 800

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Glu Trp Asn Thr Ala Met Glu Lys Leu Thr Gly Trp Ser Arg Ala Asp
805 810 815

Val Ile Gly Lys Met Leu Val Gly Glu Val Phe Gly Ser Cys Cys Gln
820 825 830

Leu Lys Gly Ser Asp Ser Ile Thr Lys Phe Met Ile Val Leu His Asn
835 840 845

Ala Leu Gly Gly His Asp Thr Asp Arg Phe Pro Phe Ser Phe Leu Asp
850 855 860

Arg Tyr Gly Lys His Val Gln Ala Phe Leu Thr Ala Asn Lys Arg Val
865 870 875 880

Asn Met Asp Gly Gln Ile Ile Gly Ala Phe Cys Phe Leu Gln Ile Val
885 890 895

Ser Pro Glu Leu Gln Gln Ala Leu Lys Ala Gln Arg Gln Gln Glu Lys
900 905 910

Asn Ser Phe Ala Arg Met Lys Glu Leu Ala Tyr Ile Cys Gln Gly Val
915 920 925

Lys Asn Pro Leu Ser Gly Ile Arg Phe Thr Asn Ser Leu Leu Glu Ala
930 935 940

Thr Cys Leu Ser Asn Glu Gln Lys Gln Phe Leu Glu Thr Ser Ala Ala
945 950 955 960

Cys Glu Lys Gln Met Leu Lys Ile Ile His Asp Val Asp Ile Glu Ser
965 970 975

Ile Glu Asp Gly Ser Leu Glu Leu Glu Lys Gly Glu Phe Leu Leu Gly
980 985 990

Asn Val Ile Asn Ala Val Val Ser Gln Val Met Leu Leu Leu Arg Glu
995 1000 1005

Arg Asn Leu Gln Leu Ile Arg Asp Ile Pro Glu Glu Ile Lys Thr
1010 1015 1020

Leu Ala Val Tyr Gly Asp Gln Leu Arg Ile Gln Gln Val Leu Ser
1025 1030 1035

Asp Phe Leu Leu Asn Ile Val Arg Tyr Ala Pro Ser Pro Asp Gly
1040 1045 1050

Trp Val Glu Ile His Val His Pro Arg Ile Lys Gln Ile Ser Asp
1055 1060 1065

Gly Leu Thr Leu Leu His Ala Glu Phe Arg Met Val Cys Pro Gly
1070 1075 1080

Glu Gly Leu Pro Pro Glu Leu Ile Gln Asn Met Phe Asn Asn Ser
1085 1090 1095

Gly Trp Gly Thr Gln Glu Gly Leu Gly Leu Ser Met Ser Arg Lys
1100 1105 1110

Ile Leu Lys Leu Met Asn Gly Glu Val Gln Tyr Ile Arg Glu Ala
1115 1120 1125

Gln Arg Cys Tyr Phe Tyr Val Leu Leu Glu Leu Pro Val Thr Arg
1130 1135 1140

Arg Ser Ser Lys Lys Cys
1145

<210> SEQ ID NO 7

<211> LENGTH: 1100

<212> TYPE: PRT

<213> ORGANISM: Glycine max

<400> SEQUENCE: 7

Met Ser Lys Ala Ile Ala Gln Tyr Thr Glu Asp Ala Arg Leu His Ala
1 5 10 15

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Val Phe Glu Gln Ser Gly Glu Ser Gly Arg Ser Phe Asn Tyr Ser Glu
 20 25 30
 Ser Ile Arg Ile Ala Ser Glu Ser Val Pro Glu Gln Gln Ile Thr Ala
 35 40 45
 Tyr Leu Val Lys Ile Gln Arg Gly Gly Phe Ile Gln Pro Phe Gly Ser
 50 55 60
 Met Ile Ala Val Asp Glu Pro Ser Phe Arg Ile Leu Gly Tyr Ser Asp
 65 70 75 80
 Asn Ala Arg Asp Met Leu Gly Ile Thr Pro Gln Ser Val Pro Ser Leu
 85 90 95
 Asp Asp Lys Asn Asp Ala Ala Phe Ala Leu Gly Thr Asp Val Arg Ala
 100 105 110
 Leu Phe Thr His Ser Ser Ala Leu Leu Leu Glu Lys Ala Phe Ser Ala
 115 120 125
 Arg Glu Ile Ser Leu Met Asn Pro Ile Trp Ile His Ser Arg Thr Ser
 130 135 140
 Gly Lys Pro Phe Tyr Gly Ile Leu His Arg Ile Asp Val Gly Ile Val
 145 150 155 160
 Ile Asp Leu Glu Pro Ala Arg Thr Glu Asp Pro Ala Leu Ser Ile Ala
 165 170 175
 Gly Ala Val Gln Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Gln Leu
 180 185 190
 Gln Ser Leu Pro Gly Gly Asp Val Lys Leu Leu Cys Asp Thr Val Val
 195 200 205
 Glu Ser Val Arg Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Lys
 210 215 220
 Phe His Glu Asp Glu His Gly Glu Val Val Ser Glu Ser Lys Arg Pro
 225 230 235 240
 Asp Leu Glu Pro Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro
 245 250 255
 Gln Ala Ser Arg Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val
 260 265 270
 Asp Cys His Ala Ser Ala Val Arg Val Val Gln Asp Glu Ala Leu Val
 275 280 285
 Gln Pro Leu Cys Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys
 290 295 300
 His Ala Gln Tyr Met Ala Asn Met Gly Ser Ile Ala Ser Leu Val Met
 305 310 315 320
 Ala Val Ile Ile Asn Gly Asn Asp Glu Glu Gly Val Gly Gly Arg Ser
 325 330 335
 Ser Met Arg Leu Trp Gly Leu Val Val Cys His His Thr Ser Ala Arg
 340 345 350
 Cys Ile Pro Phe Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala
 355 360 365
 Phe Gly Leu Gln Leu Asn Met Glu Leu Gln Leu Ala Ala Gln Ser Leu
 370 375 380
 Glu Lys Arg Val Leu Arg Thr Gln Thr Leu Leu Cys Asp Met Leu Leu
 385 390 395 400
 Arg Asp Ser Pro Thr Gly Ile Val Thr Gln Ser Pro Ser Ile Met Asp
 405 410 415
 Leu Val Lys Cys Asp Gly Ala Ala Leu Tyr Phe Gln Gly Asn Tyr Tyr
 420 425 430

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Pro Leu Gly Val Thr Pro Thr Glu Ala Gln Ile Arg Asp Ile Ile Glu
 435 440 445

Trp Leu Leu Ala Phe His Gly Asp Ser Thr Gly Leu Ser Thr Asp Ser
 450 455 460

Leu Gly Asp Ala Gly Tyr Pro Gly Ala Ala Ser Leu Gly Asp Ala Val
 465 470 475 480

Cys Gly Met Ala Val Ala Tyr Ile Thr Glu Lys Asp Phe Leu Phe Trp
 485 490 495

Phe Arg Ser His Thr Ala Lys Glu Ile Lys Trp Gly Gly Ala Lys His
 500 505 510

His Pro Glu Asp Lys Asp Asp Gly Gln Arg Met His Pro Arg Ser Ser
 515 520 525

Phe Lys Ala Phe Leu Glu Val Val Lys Ser Arg Ser Leu Pro Trp Glu
 530 535 540

Asn Ala Glu Met Asp Ala Ile His Ser Leu Gln Leu Ile Leu Arg Asp
 545 550 555 560

Ser Phe Lys Asp Ala Glu His Arg Asn Ser Lys Ala Val Ala Asp Pro
 565 570 575

Arg Val Ser Glu Gln Glu Leu Gln Gly Val Asp Glu Leu Ser Ser Val
 580 585 590

Ala Arg Glu Met Val Arg Leu Ile Glu Thr Ala Thr Ala Pro Ile Phe
 595 600 605

Ala Val Asp Val Asp Gly His Val Asn Gly Trp Asn Ala Lys Val Ser
 610 615 620

Glu Leu Thr Gly Leu Pro Val Glu Glu Ala Met Gly Lys Ser Leu Val
 625 630 635 640

His Asp Leu Val Phe Lys Glu Ser Glu Glu Thr Met Asn Lys Leu Leu
 645 650 655

Ser Arg Ala Leu Lys Gly Glu Glu Asp Lys Asn Val Glu Ile Lys Met
 660 665 670

Arg Thr Phe Gly Pro Glu Arg Gln Asn Lys Ala Val Phe Leu Val Val
 675 680 685

Asn Ala Cys Ser Ser Lys Asp Phe Thr Asn Asn Val Val Gly Val Cys
 690 695 700

Phe Val Gly Gln Asp Val Thr Gly Gln Lys Ile Val Met Asp Lys Phe
 705 710 715 720

Ile Asn Ile Gln Gly Asp Tyr Lys Ala Ile Val His Ser Pro Asn Pro
 725 730 735

Leu Ile Pro Pro Ile Phe Ala Ser Asp Asp Asn Thr Cys Cys Leu Glu
 740 745 750

Trp Asn Thr Ala Met Glu Lys Leu Thr Gly Trp Gly Arg Val Asp Val
 755 760 765

Ile Gly Lys Met Leu Val Gly Glu Val Phe Gly Ser Cys Cys Gln Leu
 770 775 780

Lys Gly Ser Asp Ser Ile Thr Lys Phe Met Ile Val Leu His Asn Ala
 785 790 795 800

Leu Gly Gly Gln Asp Thr Asp Lys Phe Pro Phe Ser Phe Leu Asp Arg
 805 810 815

His Gly Lys Tyr Val Gln Thr Phe Leu Thr Ala Asn Lys Arg Val Asn
 820 825 830

Met Glu Gly Gln Ile Ile Gly Ala Phe Cys Phe Leu Gln Ile Met Ser
 835 840 845

Pro Glu Leu Gln Gln Ala Leu Lys Ala Gln Arg Gln Gln Glu Lys Asn

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850						855						860					
Ser	Phe	Gly	Arg	Met	Lys	Glu	Leu	Ala	Tyr	Ile	Cys	Gln	Gly	Val	Lys		
865					870					875					880		
Asn	Pro	Leu	Ser	Gly	Ile	Arg	Phe	Thr	Asn	Ser	Leu	Leu	Glu	Ala	Thr		
			885						890					895			
Ser	Leu	Thr	Asn	Glu	Gln	Lys	Gln	Phe	Leu	Glu	Thr	Ser	Val	Ala	Cys		
			900					905					910				
Glu	Lys	Gln	Met	Leu	Lys	Ile	Ile	Arg	Asp	Val	Asp	Leu	Glu	Ser	Ile		
		915				920					925						
Glu	Asp	Gly	Ser	Leu	Glu	Leu	Glu	Lys	Gly	Glu	Phe	Leu	Leu	Gly	Asn		
	930				935						940						
Val	Ile	Asn	Ala	Val	Val	Ser	Gln	Val	Met	Leu	Leu	Leu	Arg	Glu	Arg		
945				950					955					960			
Asn	Leu	Gln	Leu	Ile	Arg	Asp	Ile	Pro	Glu	Glu	Ile	Lys	Thr	Leu	Ala		
			965					970						975			
Val	Tyr	Gly	Asp	Gln	Leu	Arg	Ile	Gln	Gln	Val	Leu	Ser	Asp	Phe	Leu		
			980					985					990				
Leu	Asn	Ile	Val	Arg	Tyr	Ala	Pro	Ser	Pro	Asp	Gly	Trp	Val	Glu	Ile		
		995				1000						1005					
His	Val	Arg	Pro	Arg	Ile	Lys	Gln	Ile	Ser	Asp	Gly	Leu	Thr	Leu			
1010						1015					1020						
Leu	His	Ala	Glu	Phe	Arg	Met	Val	Cys	Pro	Gly	Glu	Gly	Leu	Pro			
1025						1030					1035						
Pro	Glu	Leu	Ile	Gln	Asp	Met	Phe	Asn	Asn	Ser	Arg	Trp	Gly	Thr			
1040						1045					1050						
Gln	Glu	Gly	Leu	Gly	Leu	Ser	Met	Ser	Arg	Lys	Ile	Leu	Lys	Leu			
1055						1060					1065						
Met	Asn	Gly	Glu	Val	Gln	Tyr	Ile	Arg	Glu	Ala	Glu	Arg	Cys	Tyr			
1070						1075					1080						
Phe	Tyr	Val	Leu	Leu	Glu	Leu	Pro	Val	Thr	Arg	Arg	Ser	Ser	Lys			
1085						1090					1095						
Lys	Cys																
1100																	

<210> SEQ ID NO 8
 <211> LENGTH: 867
 <212> TYPE: PRT
 <213> ORGANISM: Glycine max

<400> SEQUENCE: 8

Met	Ile	Ala	Val	Asp	Glu	Pro	Ser	Phe	Arg	Ile	Leu	Ala	Tyr	Ser	Asp
1				5					10					15	
Asn	Ala	Arg	Asp	Met	Leu	Gly	Ile	Thr	Pro	Gln	Ser	Val	Pro	Ser	Leu
		20						25					30		
Asp	Asp	Lys	Asn	Asp	Ala	Ala	Phe	Ala	Leu	Gly	Thr	Asp	Ile	Arg	Thr
		35					40					45			
Leu	Phe	Thr	His	Ser	Ser	Ala	Val	Leu	Leu	Glu	Lys	Ala	Phe	Ser	Ala
		50				55					60				
Arg	Glu	Ile	Ser	Leu	Met	Asn	Pro	Ile	Trp	Ile	His	Ser	Arg	Thr	Ser
65					70					75					80
Gly	Lys	Pro	Phe	Tyr	Gly	Ile	Leu	His	Arg	Ile	Asp	Val	Gly	Ile	Val
				85					90					95	
Ile	Asp	Leu	Glu	Pro	Ala	Arg	Thr	Glu	Asp	Pro	Ala	Leu	Ser	Ile	Ala
				100					105						110

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Gly Ala Val Gln Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Gln Leu
 115 120 125
 Gln Ser Leu Pro Gly Gly Asp Val Lys Leu Leu Cys Asp Thr Val Val
 130 135 140
 Glu Ser Val Arg Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Arg
 145 150 155 160
 Phe His Glu Asp Glu His Gly Glu Val Val Ala Glu Thr Lys Arg Pro
 165 170 175
 Asp Leu Glu Pro Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro
 180 185 190
 Gln Ala Ser Arg Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val
 195 200 205
 Asp Cys His Ala Ser Ala Val Arg Val Val Gln Asp Glu Ala Leu Val
 210 215 220
 Gln Pro Leu Cys Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys
 225 230 235 240
 His Ala Gln Tyr Met Ala Asn Met Gly Ser Thr Ala Ser Leu Val Met
 245 250 255
 Ala Val Ile Ile Asn Gly Asn Asp Glu Glu Gly Val Gly Gly Arg Thr
 260 265 270
 Ser Met Arg Leu Trp Gly Leu Val Ile Cys His His Thr Ser Ala Arg
 275 280 285
 Cys Ile Pro Phe Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala
 290 295 300
 Phe Gly Leu Gln Leu Asn Met Glu Leu Gln Leu Ala Ala Gln Ser Leu
 305 310 315 320
 Glu Lys Arg Val Leu Arg Thr Gln Thr Leu Leu Cys Asp Met Leu Leu
 325 330 335
 Arg Asp Ser Pro Thr Gly Ile Val Thr Gln Ser Pro Ser Ile Met Asp
 340 345 350
 Leu Val Lys Cys Asp Gly Ala Ala Leu Tyr Tyr Gln Gly Asn Tyr Tyr
 355 360 365
 Pro Leu Gly Val Thr Pro Thr Glu Ala Gln Ile Arg Asp Ile Ile Glu
 370 375 380
 Trp Leu Leu Ala Phe His Arg Asp Ser Thr Gly Leu Ser Thr Asp Ser
 385 390 395 400
 Leu Ala Asp Ala Gly Tyr Pro Gly Ala Ala Ser Leu Gly Asp Ala Val
 405 410 415
 Cys Gly Met Ala Val Ala Tyr Ile Thr Glu Lys Asp Phe Leu Phe Trp
 420 425 430
 Phe Arg Ser His Thr Ala Lys Glu Ile Lys Trp Gly Gly Ala Lys His
 435 440 445
 His Pro Glu Asp Lys Asp Asp Gly Gln Arg Met His Pro Arg Ser Ser
 450 455 460
 Phe Lys Ala Phe Leu Glu Val Val Lys Ser Arg Ser Leu Pro Trp Glu
 465 470 475 480
 Ser Ala Glu Met Asp Ala Ile His Ser Leu Gln Leu Ile Leu Arg Asp
 485 490 495
 Ser Phe Lys Asp Ala Glu His Ser Asn Ser Lys Ala Val Leu Asp Pro
 500 505 510
 Arg Met Ser Glu Leu Glu Leu Gln Gly Val Asp Glu Leu Ser Ser Val
 515 520 525
 Ala Arg Glu Met Val Arg Leu Ile Glu Thr Ala Thr Ala Pro Ile Phe

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530			535			540		
Ala Val Asp	Val Asp Gly	Arg Ile Asn Gly	Trp Asn Ala Lys	Val Ser				
545	550	555	555	560				
Glu Leu Thr	Gly Leu Pro	Val Glu Glu Ala	Met Gly Lys Ser	Leu Val				
	565		570	575				
Arg Asp Leu	Val Phe Lys	Glu Ser Glu Glu	Thr Val Asp Lys	Leu Leu				
	580		585	590				
Ser Arg Ala	Leu Lys Gly	Glu Glu Asp Lys	Asn Val Glu Ile	Lys Met				
	595		600	605				
Arg Thr Phe	Gly Pro Glu	His Gln Asn Lys	Ala Val Phe	Val Val Val				
	610		615	620				
Asn Ala Cys	Ser Ser Lys	Asp Tyr Thr Asn	Asn Val Val Gly	Val Cys				
625	630		635	640				
Phe Val Gly	Gln Asp Val	Thr Gly Gln Lys	Ile Val Met	Asp Lys Phe				
	645		650	655				
Ile Asn Ile	Gln Gly Asp	Tyr Lys Ala Ile	Val His Asn Pro	Asn Pro				
	660		665	670				
Leu Ile Pro	Pro Ile Phe	Ala Ser Asp Asp	Asn Thr Cys Cys	Leu Glu				
	675		680	685				
Trp Asn Thr	Ala Met Glu	Lys Leu Thr Gly	Trp Ser Arg	Ala Asp Val				
	690		695	700				
Ile Gly Lys	Met Leu Val	Gly Glu Val Phe	Gly Ser Cys Cys	Gln Leu				
705	710		715	720				
Lys Gly Ser	Asp Ser Ile	Thr Lys Phe Met	Ile Val Leu His	Asn Ala				
	725		730	735				
Leu Gly Gly	His Asp Thr	Asp Arg Phe Pro	Phe Ser Phe Leu	Asp Arg				
	740		745	750				
Tyr Gly Lys	His Val Gln	Ala Phe Leu Thr	Ala Asn Lys Arg	Val Asn				
	755		760	765				
Met Asp Gly	Gln Ile Ile	Gly Ala Phe Cys	Phe Leu Gln Ile	Val Ser				
	770		775	780				
Pro Glu Leu	Gln Gln Ala	Leu Lys Ala Gln	Arg Gln Gln	Glu Lys Asn				
785	790		795	800				
Ser Phe Ala	Arg Met Lys	Glu Leu Ala Tyr	Ile Cys Gln Gly	Val Lys				
	805		810	815				
Asn Pro Leu	Ser Gly Ile	Arg Phe Thr Asn	Ser Leu Leu Glu	Ala Thr				
	820		825	830				
Cys Leu Ser	Asn Glu Gln	Lys Gln Phe Leu	Glu Thr Ser Ala	Ala Cys				
	835		840	845				
Glu Lys Gln	Met Leu Lys	Ile Ile His Asp	Val Asp Ile Glu	Ser Ile				
	850		855	860				
Glu Asp Gly								
865								

<210> SEQ ID NO 9

<211> LENGTH: 1130

<212> TYPE: PRT

<213> ORGANISM: Solanum tuberosum

<400> SEQUENCE: 9

Met Ala Ser	Gly Ser Arg	Thr Lys His	Ser His His	Asn Ser Ser	Gln
1	5	10	15		

Ala Gln Ser	Ser Gly Thr	Ser Asn Val	Asn Tyr Lys	Asp Ser Ile	Ser
20	25	30			

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Lys Ala Ile Ala Gln Tyr Thr Ala Asp Ala Arg Leu His Ala Val Phe
 35 40 45

 Glu Gln Ser Gly Glu Ser Gly Lys Phe Phe Asp Tyr Ser Glu Ser Val
 50 55 60

 Lys Thr Thr Thr Gln Ser Val Pro Glu Arg Gln Ile Thr Ala Tyr Leu
 65 70 75 80

 Thr Lys Ile Gln Arg Gly Gly His Ile Gln Pro Phe Gly Cys Met Ile
 85 90 95

 Ala Val Asp Glu Ala Ser Phe Arg Val Ile Ala Tyr Ser Glu Asn Ala
 100 105 110

 Phe Glu Met Leu Ser Leu Thr Pro Gln Ser Val Pro Ser Leu Glu Lys
 115 120 125

 Cys Glu Ile Leu Thr Ile Gly Thr Asp Val Arg Thr Leu Phe Thr Pro
 130 135 140

 Ser Ser Ser Val Leu Leu Glu Arg Ala Phe Gly Ala Arg Glu Ile Thr
 145 150 155

 Leu Leu Asn Pro Ile Trp Ile His Ser Lys Asn Ser Gly Lys Pro Phe
 165 170 175

 Tyr Ala Ile Leu His Arg Val Asp Val Gly Ile Ala Ile Asp Leu Glu
 180 185 190

 Pro Ala Arg Thr Glu Asp Pro Ala Leu Ser Ile Ala Gly Ala Val Gln
 195 200 205

 Ser Gln Lys Leu Ala Val Arg Ala Ile Ser His Leu Gln Ser Leu Pro
 210 215 220

 Gly Gly Asp Ile Lys Leu Leu Cys Asp Thr Val Val Glu Ser Val Arg
 225 230 235 240

 Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Lys Phe His Glu Asp
 245 250 255

 Glu His Gly Glu Val Val Ala Glu Ser Lys Arg Ser Asp Leu Glu Pro
 260 265 270

 Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
 275 280 285

 Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val Asp Cys His Ala
 290 295 300

 Thr Pro Val Arg Val Thr Gln Asp Glu Ser Leu Met Gln Pro Leu Cys
 305 310 315 320

 Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys His Ala Gln Tyr
 325 330 335

 Met Ala Asn Met Gly Ser Ile Ala Ser Leu Thr Leu Ala Val Ile Ile
 340 345 350

 Asn Gly Asn Asp Glu Glu Ala Val Gly Gly Gly Arg Asn Ser Met Arg
 355 360 365

 Leu Trp Gly Leu Val Val Gly His His Thr Ser Val Arg Ser Ile Pro
 370 375 380

 Phe Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala Phe Gly Leu
 385 390 395 400

 Gln Leu Asn Met Glu Leu Gln Leu Ala Ser Gln Leu Ser Glu Lys His
 405 410 415

 Val Leu Arg Thr Gln Thr Leu Leu Cys Asp Met Leu Leu Arg Asp Ser
 420 425 430

 Pro Pro Gly Ile Val Thr Gln Ser Pro Ser Ile Met Asp Leu Val Lys
 435 440 445

 Cys Asp Gly Ala Ala Leu Tyr Tyr Gln Gly Lys Tyr Tyr Pro Leu Gly

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Gln Gln Ala Leu Arg Val Gln Arg Gln Gln Glu Lys Lys Cys Tyr Ser
      885                               890                               895

Gln Met Lys Glu Leu Ala Tyr Ile Cys Gln Glu Ile Lys Ser Pro Leu
      900                               905                               910

Asn Gly Ile Arg Phe Thr Asn Ser Leu Leu Glu Ala Thr Asn Leu Thr
      915                               920                               925

Glu Asn Gln Lys Gln Tyr Leu Glu Thr Ser Ala Ala Cys Glu Arg Gln
      930                               935                               940

Met Ser Lys Ile Ile Arg Asp Val Asp Leu Glu Asn Ile Glu Asp Gly
945                               950                               955                               960

Ser Leu Thr Leu Glu Lys Glu Asp Phe Phe Leu Gly Ser Val Ile Asp
      965                               970                               975

Ala Val Val Ser Gln Val Met Leu Leu Leu Arg Glu Lys Gly Val Gln
      980                               985                               990

Leu Ile Arg Asp Ile Pro Glu Glu Ile Lys Thr Leu Thr Val His Gly
      995                               1000                              1005

Asp Gln Val Arg Ile Gln Gln Val Leu Ala Asp Phe Leu Leu Asn
1010                               1015                              1020

Met Val Arg Tyr Ala Pro Ser Pro Asp Gly Trp Val Glu Ile Gln
1025                               1030                              1035

Leu Arg Pro Ser Met Met Pro Ile Ser Asp Gly Val Thr Gly Val
1040                               1045                              1050

His Ile Glu Leu Arg Ile Ile Cys Pro Gly Glu Gly Leu Pro Pro
1055                               1060                              1065

Glu Leu Val Gln Asp Met Phe His Ser Ser Arg Trp Val Thr Gln
1070                               1075                              1080

Glu Gly Leu Gly Leu Ser Thr Cys Arg Lys Met Leu Lys Leu Met
1085                               1090                              1095

Asn Gly Glu Ile Gln Tyr Ile Arg Glu Ser Glu Arg Cys Tyr Phe
1100                               1105                              1110

Leu Ile Val Leu Asp Leu Pro Met Thr Arg Lys Gly Pro Lys Ser
1115                               1120                              1125

Val Gly
1130

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<210> SEQ ID NO 10

<211> LENGTH: 1121

<212> TYPE: PRT

<213> ORGANISM: Pisum sativum

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<222> LOCATION: (28)..(28)

<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid
(Ser or Arg)

<400> SEQUENCE: 10

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Ser Asn Asn Asn Asn Asn Arg Asn Ile Lys Arg Glu Ser Leu Ser Met
1      5              10              15

Arg Lys Ala Ile Ala Gln Tyr Thr Glu Asp Ala Xaa Leu His Ala Val
      20              25              30

Phe Glu Lys Ser Gly Asp Ser Phe Asp Tyr Ala Gln Ser Ile Arg Val
      35              40              45

Thr Ala Ala Thr Glu Ser Val Pro Glu Gln Gln Ile Thr Ala Tyr Leu
      50              55              60

Ala Lys Ile Gln Arg Gly Gly Phe Ile Gln Pro Phe Gly Ser Met Ile
      65              70              75              80

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-continued

Ala Val Asp Glu Thr Ser Phe Arg Val Leu Ala Tyr Ser Glu Asn Ala
85 90 95

Arg Asp Met Leu Gly Ile Ala Pro Gln Ser Val Pro Ser Met Glu Asp
100 105 110

Asp Ser Ser Ser Ser Ser Phe Phe Ser Leu Gly Val Asp Val Arg Ser
115 120 125

Leu Phe Ser Ala Ser Ser Ser Val Leu Leu Glu Lys Ala Phe Ser Ala
130 135 140

Arg Glu Ile Ser Leu Met Asn Pro Ile Trp Ile His Ser Arg Ser Thr
145 150 155 160

Gly Lys Pro Phe Tyr Gly Ile Leu His Arg Ile Asp Ile Gly Val Val
165 170 175

Ile Asp Leu Glu Pro Ala Arg Ser Glu Asp Pro Ala Leu Ser Ile Ala
180 185 190

Gly Ala Val Gln Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Gln Leu
195 200 205

Gln Ala Leu Pro Gly Gly Asp Val Lys Leu Leu Cys Asp Ala Val Val
210 215 220

Glu Ser Val Arg Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Lys
225 230 235 240

Phe His Glu Asp Glu His Gly Glu Val Val Ala Glu Ser Lys Arg Val
245 250 255

Asp Leu Glu Pro Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro
260 265 270

Gln Ala Ser Arg Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val
275 280 285

Asp Cys Asn Ala Ser Pro Val Arg Val Phe Gln Asp Glu Ala Leu Val
290 295 300

Gln Pro Val Cys Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys
305 310 315 320

His Ala Gln Tyr Met Ala Asn Met Gly Ser Ile Ala Ser Leu Ala Met
325 330 335

Ala Val Ile Ile Asn Gly Asn Asp Glu Asp Gly Gly Gly Ile Gly Gly
340 345 350

Ala Ala Arg Gly Ser Met Arg Leu Trp Gly Leu Val Val Cys His His
355 360 365

Thr Ser Ala Arg Cys Ile Pro Phe Pro Leu Arg Tyr Ala Cys Glu Phe
370 375 380

Leu Met Gln Ala Phe Gly Leu Gln Leu Asn Met Glu Leu Gln Leu Ala
385 390 395 400

Val Gln Ser Leu Glu Lys Arg Val Leu Lys Thr Gln Thr Leu Leu Cys
405 410 415

Asp Met Leu Leu Arg Asp Ser His Thr Gly Ile Val Thr Gln Ser Pro
420 425 430

Ser Ile Met Asp Leu Val Lys Cys Asp Gly Ala Ala Leu Tyr Tyr Gln
435 440 445

Gly Asn Tyr His Pro Leu Gly Val Thr Pro Thr Glu Ser Gln Ile Arg
450 455 460

Asp Ile Ile Asp Trp Leu Leu Ala Phe His Ser Asp Ser Thr Gly Leu
465 470 475 480

Ser Thr Asp Ser Leu Ala Asp Ala Gly Tyr Pro Gly Ala Ala Ser Leu
485 490 495

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Gly Asp Ala Val Cys Gly Met Ala Val Ala Tyr Ile Thr Glu Lys Asp
 500 505 510
 Phe Leu Phe Trp Phe Arg Ser His Thr Ala Lys Glu Ile Lys Trp Gly
 515 520 525
 Gly Ala Lys His His Pro Glu Asp Lys Asp Asp Gly Gln Lys Met His
 530 535 540
 Pro Arg Ser Ser Phe Lys Ala Phe Leu Glu Val Val Lys Ile Arg Ser
 545 550 555 560
 Met Gln Trp Asp Asn Ala Glu Met Asp Ala Ile His Ser Leu Gln Leu
 565 570 575
 Ile Leu Arg Asp Ser Phe Lys Glu Ala Glu Asn Asn Asp Ser Lys Ala
 580 585 590
 Val Val His Thr His Met Ala Glu Leu Glu Leu Gln Gly Val Asp Glu
 595 600 605
 Leu Ser Ser Val Ala Arg Glu Met Val Arg Leu Ile Glu Thr Ala Thr
 610 615 620
 Ala Pro Ile Phe Ala Val Asp Val Asp Gly Arg Ile Asn Gly Trp Asn
 625 630 635 640
 Ala Lys Val Ser Glu Leu Thr Gly Leu Leu Val Glu Glu Ala Met Gly
 645 650 655
 Lys Ser Leu Val His Asp Leu Val Tyr Lys Glu Ser Arg Glu Thr Val
 660 665 670
 Asp Lys Leu Leu Ser His Ala Leu Lys Gly Glu Glu Asp Lys Asn Val
 675 680 685
 Glu Ile Lys Met Lys Thr Phe Gly Pro Gly Asn Gln Asn Lys Ala Val
 690 695 700
 Phe Ile Val Val Asn Ala Cys Ser Ser Lys Asp Tyr Thr Asn Asn Ile
 705 710 715 720
 Val Gly Val Cys Phe Val Gly Gln Asp Ile Thr Gly Gln Lys Val Val
 725 730 735
 Met Asp Lys Phe Ile Asn Ile Gln Gly Asp Tyr Lys Ala Ile Val His
 740 745 750
 Ser Pro Asn Pro Leu Ile Pro Pro Ile Phe Ala Ser Asp Asp Asn Thr
 755 760 765
 Cys Cys Leu Glu Trp Asn Asn Ala Met Glu Lys Leu Ser Gly Trp Ser
 770 775 780
 Arg Ala Asp Val Ile Gly Lys Leu Leu Val Gly Glu Val Phe Gly Ser
 785 790 795 800
 Phe Cys Gln Leu Lys Gly Ser Asp Ala Met Thr Lys Phe Met Ile Val
 805 810 815
 Leu His Asn Ala Leu Gly Gly His Asp Thr Asp Lys Phe Pro Leu Ser
 820 825 830
 Phe Leu Asp Arg His Gly Lys Tyr Val His Thr Phe Leu Thr Ala Asn
 835 840 845
 Lys Arg Val Asn Met Asp Gly Gln Ile Ile Gly Ala Phe Cys Phe Leu
 850 855 860
 Gln Ile Val Asn Pro Glu Leu Gln Gln Ala Leu Thr Val Gln Arg Gln
 865 870 875 880
 Gln Asp Ser Ser Ser Leu Ala Arg Met Lys Glu Leu Ala Tyr Ile Cys
 885 890 895
 Gln Glu Val Lys Asn Pro Leu Ser Gly Ile Arg Phe Thr Asn Ser Leu
 900 905 910
 Leu Glu Ser Thr Cys Leu Thr Asp Glu Gln Lys Gln Leu Leu Glu Thr

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915	920	925
Ser Val Ala Cys Glu Lys Gln Met Leu Lys Ile Val Arg Asp Ile Ala		
930	935	940
Leu Glu Ser Ile Glu Asp Gly Ser Leu Glu Leu Glu Lys Gln Glu Phe		
945	950	955
Leu Leu Glu Asn Val Ile Asn Ala Val Val Ser Gln Val Met Leu Leu		
	965	970
Leu Arg Asp Arg Lys Leu Gln Leu Ile Arg Asp Ile Pro Glu Glu Ile		
	980	985
Lys Ala Leu Ala Val Tyr Gly Asp Gln Leu Arg Ile Gln Gln Val Leu		
	995	1000
Ala Asp Phe Leu Met Asn Val Val Arg Tyr Ala Pro Ser Pro Asp		
1010	1015	1020
Gly Trp Val Glu Ile His Val Phe Pro Arg Ile Lys Gln Ile Ser		
1025	1030	1035
Glu Gly Leu Thr Leu Leu His Ala Glu Phe Arg Met Val Cys Pro		
1040	1045	1050
Gly Glu Gly Leu Pro Pro Glu Leu Ile Gln Asp Met Phe His Asn		
1055	1060	1065
Ser Arg Trp Val Thr Gln Glu Gly Leu Gly Leu Ser Met Ser Arg		
1070	1075	1080
Lys Ile Ile Lys Leu Met Asn Gly Glu Val Gln Tyr Val Arg Glu		
1085	1090	1095
Ala Glu Arg Cys Tyr Phe Leu Val Leu Leu Glu Leu Pro Val Thr		
1100	1105	1110
Arg Arg Ser Ser Lys Ala Ile Asn		
1115	1120	

<210> SEQ ID NO 11

<211> LENGTH: 1129

<212> TYPE: PRT

<213> ORGANISM: Vitis vinifera

<400> SEQUENCE: 11

Met Ser Ser Gly Asn Arg Gly Thr Gln Ser His His Gln Ala Gln Ser			
1	5	10	15
Ser Gly Thr Ser Asn Leu Arg Val Tyr His Thr Asp Ser Met Ser Lys			
	20	25	30
Ala Ile Ala Gln Tyr Thr Met Asp Ala Arg Leu His Ala Val Tyr Glu			
	35	40	45
Gln Ser Gly Glu Ser Gly Lys Ser Phe Asp Tyr Ser Gln Ser Val Arg			
	50	55	60
Thr Thr Thr Gln Ser Val Pro Glu Gln Gln Ile Thr Ala Tyr Leu Ser			
65	70	75	80
Lys Ile Gln Arg Gly Gly His Ile Gln Pro Phe Gly Cys Met Leu Ala			
	85	90	95
Val Asp Glu Ala Thr Phe Arg Val Ile Ala Phe Ser Glu Asn Ala Arg			
	100	105	110
Glu Met Leu Gly Leu Thr Pro Gln Ser Val Pro Ser Leu Glu Lys Pro			
	115	120	125
Glu Ile Leu Leu Val Gly Thr Asp Val Arg Thr Leu Phe Thr Pro Ser			
130	135	140	
Ser Ala Val Leu Leu Glu Lys Ala Phe Arg Ala Arg Glu Ile Thr Leu			
145	150	155	160

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Leu Asn Pro Val Trp Ile His Ser Lys Asn Ser Gly Lys Pro Phe Tyr
 165 170 175
 Ala Ile Leu His Arg Ile Asp Val Gly Ile Val Ile Asp Leu Glu Pro
 180 185 190
 Ala Arg Thr Glu Asp Pro Ala Leu Ser Ile Ala Gly Ala Val Gln Ser
 195 200 205
 Gln Lys Leu Ala Val Arg Ala Ile Ser His Leu Gln Ser Leu Pro Gly
 210 215
 Gly Asp Ile Asn Leu Leu Cys Glu Thr Val Val Glu Asn Val Arg Glu
 225 230 235 240
 Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Lys Phe His Glu Asp Glu
 245 250 255
 His Gly Glu Val Val Ala Glu Ser Lys Arg Ser Asp Leu Glu Pro Tyr
 260 265 270
 Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg Phe
 275 280 285
 Leu Phe Arg Gln Asn Arg Val Arg Met Ile Val Asp Cys His Ala Thr
 290 295 300
 Pro Val Leu Val Ile Gln Asp Glu Gly Leu Met Gln Pro Leu Cys Leu
 305 310 315 320
 Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys His Ala Gln Tyr Met
 325 330 335
 Ala Asn Met Gly Ser Thr Ala Ser Leu Ala Met Ala Val Ile Ile Asn
 340 345 350
 Gly Ser Asp Glu Glu Ala Ile Gly Gly Arg Asn Leu Met Arg Leu Trp
 355 360 365
 Gly Leu Val Val Cys His His Thr Ser Ala Arg Cys Ile Pro Phe Pro
 370 375 380
 Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala Phe Gly Leu Gln Leu
 385 390 395 400
 Asn Met Glu Leu Gln Leu Ala Ser Gln Leu Ser Glu Lys His Val Leu
 405 410 415
 Arg Thr Gln Thr Leu Leu Cys Asp Met Leu Leu Arg Asp Ser Pro Thr
 420 425 430
 Gly Ile Val Thr Gln Ser Pro Ser Ile Met Asp Leu Val Lys Cys Asp
 435 440 445
 Gly Ala Ala Leu Tyr Tyr Gln Gly Lys Tyr Tyr Pro Thr Gly Val Thr
 450 455 460
 Pro Thr Glu Ala Gln Ile Lys Asp Ile Ala Glu Trp Leu Leu Ala Asn
 465 470 475 480
 His Ala Asp Ser Thr Gly Leu Ser Thr Asp Ser Leu Ala Asp Ala Gly
 485 490 495
 Tyr Pro Gly Ala Ala Ser Leu Gly Asp Ala Val Cys Gly Met Ala Val
 500 505 510
 Ala Tyr Ile Thr Ser Arg Asp Phe Leu Phe Trp Phe Arg Ser His Thr
 515 520 525
 Ala Lys Glu Ile Lys Trp Gly Gly Ala Lys His His Pro Glu Asp Lys
 530 535 540
 Asp Asp Gly Gln Arg Met His Pro Arg Ser Ser Phe Lys Ala Phe Leu
 545 550 555 560
 Glu Val Val Lys Ser Arg Ser Leu Pro Trp Glu Asn Ala Glu Met Asp
 565 570 575
 Ala Ile His Ser Leu Gln Leu Ile Leu Arg Asp Ser Phe Lys Asp Ala

-continued

580				585				590							
Thr	Asp	Gly	Ser	Asn	Ser	Lys	Ala	Val	Met	His	Ala	Gln	Leu	Gly	Glu
	595						600					605			
Leu	Glu	Leu	Gln	Gly	Met	Asp	Glu	Leu	Ser	Ser	Val	Ala	Arg	Glu	Met
	610					615					620				
Val	Arg	Leu	Ile	Glu	Thr	Ala	Thr	Ala	Pro	Ile	Phe	Ala	Val	Asp	Val
	625				630					635					640
Asp	Gly	Cys	Ile	Asn	Gly	Trp	Asn	Ala	Lys	Val	Ala	Glu	Leu	Thr	Gly
			645						650					655	
Leu	Ser	Val	Glu	Glu	Ala	Met	Gly	Lys	Ser	Leu	Val	His	Asp	Leu	Val
		660						665					670		
Tyr	Lys	Glu	Ser	Glu	Glu	Thr	Val	Asp	Lys	Leu	Leu	His	His	Ala	Leu
		675					680						685		
Arg	Gly	Glu	Glu	Asp	Lys	Asn	Val	Glu	Ile	Lys	Leu	Arg	Thr	Phe	Asp
		690				695					700				
Ser	Gln	Gln	His	Lys	Lys	Ala	Val	Phe	Val	Val	Val	Asn	Ala	Cys	Ser
					710						715				720
Ser	Arg	Asp	Tyr	Thr	Asn	Asn	Ile	Val	Gly	Val	Cys	Phe	Val	Gly	Gln
			725						730					735	
Asp	Val	Thr	Gly	Gln	Lys	Val	Val	Met	Asp	Lys	Phe	Ile	His	Ile	Gln
		740						745					750		
Gly	Asp	Tyr	Lys	Ala	Ile	Val	His	Ser	Pro	Asn	Pro	Leu	Ile	Pro	Pro
		755					760					765			
Ile	Phe	Ala	Ser	Asp	Glu	Asn	Thr	Val	Cys	Ser	Glu	Trp	Asn	Thr	Ala
		770				775					780				
Met	Glu	Lys	Leu	Thr	Gly	Trp	Ser	Arg	Gly	Asp	Ile	Ile	Gly	Lys	Ile
					790					795					800
Leu	Val	Gly	Glu	Ile	Phe	Gly	Ser	Ser	Cys	Arg	Leu	Lys	Gly	Pro	Asp
			805						810					815	
Ala	Leu	Thr	Lys	Phe	Met	Ile	Val	Leu	His	Asn	Ala	Ile	Gly	Gly	Gln
			820					825					830		
Asp	Thr	Asp	Lys	Phe	Pro	Phe	Ser	Phe	Phe	Asp	Gln	Asn	Gly	Lys	Tyr
		835					840						845		
Val	Gln	Ala	Leu	Leu	Thr	Ala	Asn	Lys	Arg	Val	Asn	Ile	Glu	Gly	Gln
		850				855					860				
Ile	Ile	Gly	Ala	Phe	Cys	Phe	Leu	Gln	Ile	Ala	Ser	Pro	Glu	Leu	Gln
		865			870					875					880
Gln	Ala	Leu	Lys	Val	Gln	Arg	Gln	Gln	Glu	Lys	Lys	Cys	Phe	Ala	Arg
			885						890					895	
Met	Lys	Glu	Leu	Ala	Tyr	Ile	Cys	Gln	Glu	Ile	Lys	Asn	Pro	Leu	Ser
		900						905					910		
Gly	Ile	Arg	Phe	Thr	Asn	Ser	Leu	Leu	Glu	Ala	Thr	Asp	Leu	Thr	Glu
		915					920						925		
Asp	Gln	Lys	Gln	Phe	Leu	Glu	Thr	Ser	Ala	Ala	Cys	Glu	Lys	Gln	Met
		930				935					940				
Ser	Lys	Ile	Ile	Arg	Asp	Val	Asp	Leu	Asp	Ser	Ile	Glu	Asp	Gly	Ser
					950					955					960
Leu	Glu	Leu	Glu	Arg	Ala	Glu	Phe	Leu	Leu	Gly	Ser	Val	Ile	Asn	Ala
			965						970					975	
Val	Val	Ser	Gln	Val	Met	Ile	Leu	Leu	Arg	Glu	Arg	Asp	Leu	Gln	Leu
			980						985					990	
Ile	Arg	Asp	Ile	Pro	Glu	Glu	Val	Lys	Thr	Leu	Ala	Val	Tyr	Gly	Asp
		995					1000							1005	

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Gln Val Arg Ile Gln Gln Val Leu Ala Asp Phe Leu Leu Asn Met
 1010 1015 1020
 Val Arg Tyr Ala Pro Ser Pro Asp Gly Trp Ile Glu Ile Gln Val
 1025 1030 1035
 Cys Pro Arg Leu Lys Gln Ile Ser Glu Glu Val Lys Leu Met His
 1040 1045 1050
 Ile Glu Phe Arg Met Val Cys Pro Gly Glu Gly Leu Pro Pro Asn
 1055 1060 1065
 Leu Ile Gln Asp Met Phe His Ser Ser Arg Trp Met Thr Gln Glu
 1070 1075 1080
 Gly Leu Gly Leu Ser Met Cys Arg Lys Ile Leu Lys Leu Ile Asn
 1085 1090 1095
 Gly Glu Val Gln Tyr Ile Arg Glu Ser Glu Arg Cys Tyr Phe Leu
 1100 1105 1110
 Ile Ser Ile Glu Leu Pro Ile Pro His Arg Gly Ser Lys Ser Val
 1115 1120 1125

Asp

<210> SEQ ID NO 12
 <211> LENGTH: 200
 <212> TYPE: PRT
 <213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 12

Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Gln Leu Gln Ala Leu Pro
 1 5 10 15
 Gly Gly Asp Ile Lys Leu Leu Cys Asp Thr Val Val Glu Ser Val Arg
 20 25 30
 Asp Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Lys Phe His Glu Asp
 35 40 45
 Glu His Gly Glu Val Val Ala Glu Ser Lys Arg Asp Asp Leu Glu Pro
 50 55 60
 Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
 65 70 75 80
 Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val Asp Cys Asn Ala
 85 90 95
 Thr Pro Val Leu Val Val Gln Asp Asp Arg Leu Thr Gln Ser Met Cys
 100 105 110
 Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys His Ser Gln Tyr
 115 120 125
 Met Ala Asn Met Gly Ser Ile Ala Ser Leu Ala Met Ala Val Ile Ile
 130 135 140
 Asn Gly Asn Glu Asp Asp Gly Ser Asn Val Ala Ser Gly Arg Ser Ser
 145 150 155 160
 Met Arg Leu Trp Gly Leu Val Val Cys His His Thr Ser Ser Arg Cys
 165 170 175
 Ile Pro Phe Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala Phe
 180 185 190
 Gly Leu Gln Leu Asn Met Glu Leu
 195 200

<210> SEQ ID NO 13
 <211> LENGTH: 201
 <212> TYPE: PRT
 <213> ORGANISM: Zea mays

-continued

<400> SEQUENCE: 13

Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Arg Leu Gln Ala Leu Pro
 1 5 10 15
 Gly Gly Asp Val Lys Leu Leu Cys Asp Thr Val Val Glu His Val Arg
 20 25 30
 Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Arg Phe His Glu Asp
 35 40 45
 Glu His Gly Glu Val Val Ala Glu Ser Arg Arg Asp Asn Leu Glu Pro
 50 55 60
 Tyr Leu Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
 65 70 75 80
 Phe Leu Phe Arg Gln Asn Arg Val Arg Met Ile Ala Asp Cys His Ala
 85 90 95
 Thr Pro Val Arg Val Ile Gln Asp Pro Gly Leu Ser Gln Pro Leu Cys
 100 105 110
 Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys His Ala Gln Tyr
 115 120 125
 Met Ala Asn Met Gly Ser Ile Ala Ser Leu Val Met Ala Val Ile Ile
 130 135 140
 Ser Ser Gly Gly Asp Asp Glu Gln Thr Gly Arg Gly Gly Ile Ser Ser
 145 150 155 160
 Ala Met Lys Leu Trp Gly Leu Val Val Cys His His Thr Ser Pro Arg
 165 170 175
 Cys Ile Pro Phe Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala
 180 185 190
 Phe Gly Leu Gln Leu Asn Met Glu Leu
 195 200

<210> SEQ ID NO 14

<211> LENGTH: 202

<212> TYPE: PRT

<213> ORGANISM: *Oryza sativa*

<400> SEQUENCE: 14

Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Arg Leu Gln Ala Leu Pro
 1 5 10 15
 Gly Gly Asp Val Lys Leu Leu Cys Asp Thr Val Val Glu His Val Arg
 20 25 30
 Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Arg Phe His Glu Asp
 35 40 45
 Glu His Gly Glu Val Val Ala Glu Ser Arg Arg Ser Asn Leu Glu Pro
 50 55 60
 Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
 65 70 75 80
 Phe Leu Phe Arg Gln Asn Arg Val Arg Met Ile Ala Asp Cys His Ala
 85 90 95
 Ala Pro Val Arg Val Ile Gln Asp Pro Ala Leu Thr Gln Pro Leu Cys
 100 105 110
 Leu Val Gly Ser Thr Leu Arg Ser Pro His Gly Cys His Ala Gln Tyr
 115 120 125
 Met Ala Asn Met Gly Ser Ile Ala Ser Leu Val Met Ala Val Ile Ile
 130 135 140
 Ser Ser Gly Gly Asp Asp Asp His Asn Ile Ala Arg Gly Ser Ile Pro
 145 150 155 160

-continued

Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val Asp Cys His Ala
85 90 95

Ser Ala Val Arg Val Val Gln Asp Glu Ala Leu Val Gln Pro Leu Cys
100 105 110

Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys His Ala Gln Tyr
115 120 125

Met Ala Asn Met Gly Ser Ile Ala Ser Leu Val Met Ala Val Ile Ile
130 135 140

Asn Gly Asn Asp Glu Glu Gly Val Gly Gly Arg Ser Ser Met Arg Leu
145 150 155 160

Trp Gly Leu Val Val Cys His His Thr Ser Ala Arg Cys Ile Pro Phe
165 170 175

Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala Phe Gly Leu Gln
180 185 190

Leu Asn Met Glu Leu
195

<210> SEQ ID NO 17
<211> LENGTH: 197
<212> TYPE: PRT
<213> ORGANISM: Glycine max

<400> SEQUENCE: 17

Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Gln Leu Gln Ser Leu Pro
1 5 10 15

Gly Gly Asp Val Lys Leu Leu Cys Asp Thr Val Val Glu Ser Val Arg
20 25 30

Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Arg Phe His Glu Asp
35 40 45

Glu His Gly Glu Val Val Ala Glu Thr Lys Arg Pro Asp Leu Glu Pro
50 55 60

Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
65 70 75 80

Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val Asp Cys His Ala
85 90 95

Ser Ala Val Arg Val Val Gln Asp Glu Ala Leu Val Gln Pro Leu Cys
100 105 110

Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys His Ala Gln Tyr
115 120 125

Met Ala Asn Met Gly Ser Thr Ala Ser Leu Val Met Ala Val Ile Ile
130 135 140

Asn Gly Asn Asp Glu Glu Gly Val Gly Gly Arg Thr Ser Met Arg Leu
145 150 155 160

Trp Gly Leu Val Ile Cys His His Thr Ser Ala Arg Cys Ile Pro Phe
165 170 175

Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala Phe Gly Leu Gln
180 185 190

Leu Asn Met Glu Leu
195

<210> SEQ ID NO 18
<211> LENGTH: 197
<212> TYPE: PRT
<213> ORGANISM: Glycine max

<400> SEQUENCE: 18

-continued

Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Gln Leu Gln Ser Leu Pro
 1 5 10 15
 Gly Gly Asp Val Lys Leu Leu Cys Asp Thr Val Val Glu Ser Val Arg
 20 25 30
 Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Lys Phe His Glu Asp
 35 40 45
 Glu His Gly Glu Val Val Ser Glu Ser Lys Arg Pro Asp Leu Glu Pro
 50 55 60
 Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
 65 70 75 80
 Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val Asp Cys His Ala
 85 90 95
 Ser Ala Val Arg Val Val Gln Asp Glu Ala Leu Val Gln Pro Leu Cys
 100 105 110
 Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys His Ala Gln Tyr
 115 120 125
 Met Ala Asn Met Gly Ser Ile Ala Ser Leu Val Met Ala Val Ile Ile
 130 135 140
 Asn Gly Asn Asp Glu Glu Gly Val Gly Gly Arg Ser Ser Met Arg Leu
 145 150 155 160
 Trp Gly Leu Val Val Cys His His Thr Ser Ala Arg Cys Ile Pro Phe
 165 170 175
 Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala Phe Gly Leu Gln
 180 185 190
 Leu Asn Met Glu Leu
 195

<210> SEQ ID NO 19
 <211> LENGTH: 197
 <212> TYPE: PRT
 <213> ORGANISM: Glycine max

<400> SEQUENCE: 19

Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Gln Leu Gln Ser Leu Pro
 1 5 10 15
 Gly Gly Asp Val Lys Leu Leu Cys Asp Thr Val Val Glu Ser Val Arg
 20 25 30
 Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Arg Phe His Glu Asp
 35 40 45
 Glu His Gly Glu Val Val Ala Glu Thr Lys Arg Pro Asp Leu Glu Pro
 50 55 60
 Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
 65 70 75 80
 Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val Asp Cys His Ala
 85 90 95
 Ser Ala Val Arg Val Val Gln Asp Glu Ala Leu Val Gln Pro Leu Cys
 100 105 110
 Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys His Ala Gln Tyr
 115 120 125
 Met Ala Asn Met Gly Ser Thr Ala Ser Leu Val Met Ala Val Ile Ile
 130 135 140
 Asn Gly Asn Asp Glu Glu Gly Val Gly Gly Arg Thr Ser Met Arg Leu
 145 150 155 160
 Trp Gly Leu Val Ile Cys His His Thr Ser Ala Arg Cys Ile Pro Phe

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      165              170              175
Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala Phe Gly Leu Gln
      180              185              190

Leu Asn Met Glu Leu
      195

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<210> SEQ ID NO 20
<211> LENGTH: 198
<212> TYPE: PRT
<213> ORGANISM: Solanum tuberosum

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<400> SEQUENCE: 20

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Ser Gln Lys Leu Ala Val Arg Ala Ile Ser His Leu Gln Ser Leu Pro
 1          5          10          15
Gly Gly Asp Ile Lys Leu Leu Cys Asp Thr Val Val Glu Ser Val Arg
      20          25          30
Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Lys Phe His Glu Asp
      35          40          45
Glu His Gly Glu Val Val Ala Glu Ser Lys Arg Ser Asp Leu Glu Pro
      50          55          60
Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
      65          70          75          80
Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val Asp Cys His Ala
      85          90          95
Thr Pro Val Arg Val Thr Gln Asp Glu Ser Leu Met Gln Pro Leu Cys
      100          105          110
Leu Val Gly Ser Thr Leu Arg Ala Pro His Gly Cys His Ala Gln Tyr
      115          120          125
Met Ala Asn Met Gly Ser Ile Ala Ser Leu Thr Leu Ala Val Ile Ile
      130          135          140
Asn Gly Asn Asp Glu Glu Ala Val Gly Gly Gly Arg Asn Ser Met Arg
      145          150          155          160
Leu Trp Gly Leu Val Val Gly His His Thr Ser Val Arg Ser Ile Pro
      165          170          175
Phe Pro Leu Arg Tyr Ala Cys Glu Phe Leu Met Gln Ala Phe Gly Leu
      180          185          190

Gln Leu Asn Met Glu Leu
      195

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<210> SEQ ID NO 21
<211> LENGTH: 201
<212> TYPE: PRT
<213> ORGANISM: Pisum sativum

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<400> SEQUENCE: 21

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Ser Gln Lys Leu Ala Val Arg Ala Ile Ser Gln Leu Gln Ala Leu Pro
 1          5          10          15
Gly Gly Asp Val Lys Leu Leu Cys Asp Ala Val Val Glu Ser Val Arg
      20          25          30
Glu Leu Thr Gly Tyr Asp Arg Val Met Val Tyr Lys Phe His Glu Asp
      35          40          45
Glu His Gly Glu Val Val Ala Glu Ser Lys Arg Val Asp Leu Glu Pro
      50          55          60
Tyr Ile Gly Leu His Tyr Pro Ala Thr Asp Ile Pro Gln Ala Ser Arg
      65          70          75          80
Phe Leu Phe Lys Gln Asn Arg Val Arg Met Ile Val Asp Cys Asn Ala

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	85		90		95										
Ser	Pro	Val	Arg	Val	Phe	Gln	Asp	Glu	Ala	Leu	Val	Gln	Pro	Val	Cys
			100					105					110		
Leu	Val	Gly	Ser	Thr	Leu	Arg	Ala	Pro	His	Gly	Cys	His	Ala	Gln	Tyr
		115					120					125			
Met	Ala	Asn	Met	Gly	Ser	Ile	Ala	Ser	Leu	Ala	Met	Ala	Val	Ile	Ile
		130				135					140				
Asn	Gly	Asn	Asp	Glu	Asp	Gly	Gly	Gly	Ile	Gly	Gly	Ala	Ala	Arg	Gly
145					150					155					160
Ser	Met	Arg	Leu	Trp	Gly	Leu	Val	Val	Cys	His	His	Thr	Ser	Ala	Arg
			165						170					175	
Cys	Ile	Pro	Phe	Pro	Leu	Arg	Tyr	Ala	Cys	Glu	Phe	Leu	Met	Gln	Ala
			180					185					190		
Phe	Gly	Leu	Gln	Leu	Asn	Met	Glu	Leu							
		195					200								

<210> SEQ ID NO 22

<211> LENGTH: 197

<212> TYPE: PRT

<213> ORGANISM: *Vitis vinifera*

<400> SEQUENCE: 22

Ser	Gln	Lys	Leu	Ala	Val	Arg	Ala	Ile	Ser	His	Leu	Gln	Ser	Leu	Pro
1				5				10						15	
Gly	Gly	Asp	Ile	Asn	Leu	Leu	Cys	Glu	Thr	Val	Val	Glu	Asn	Val	Arg
			20					25					30		
Glu	Leu	Thr	Gly	Tyr	Asp	Arg	Val	Met	Val	Tyr	Lys	Phe	His	Glu	Asp
		35				40						45			
Glu	His	Gly	Glu	Val	Val	Ala	Glu	Ser	Lys	Arg	Ser	Asp	Leu	Glu	Pro
		50				55					60				
Tyr	Ile	Gly	Leu	His	Tyr	Pro	Ala	Thr	Asp	Ile	Pro	Gln	Ala	Ser	Arg
65					70					75					80
Phe	Leu	Phe	Arg	Gln	Asn	Arg	Val	Arg	Met	Ile	Val	Asp	Cys	His	Ala
				85					90					95	
Thr	Pro	Val	Leu	Val	Ile	Gln	Asp	Glu	Gly	Leu	Met	Gln	Pro	Leu	Cys
			100					105					110		
Leu	Val	Gly	Ser	Thr	Leu	Arg	Ala	Pro	His	Gly	Cys	His	Ala	Gln	Tyr
		115					120					125			
Met	Ala	Asn	Met	Gly	Ser	Thr	Ala	Ser	Leu	Ala	Met	Ala	Val	Ile	Ile
		130				135					140				
Asn	Gly	Ser	Asp	Glu	Glu	Ala	Ile	Gly	Gly	Arg	Asn	Leu	Met	Arg	Leu
145					150					155					160
Trp	Gly	Leu	Val	Val	Cys	His	His	Thr	Ser	Ala	Arg	Cys	Ile	Pro	Phe
			165						170					175	
Pro	Leu	Arg	Tyr	Ala	Cys	Glu	Phe	Leu	Met	Gln	Ala	Phe	Gly	Leu	Gln
			180					185					190		
Leu	Asn	Met	Glu	Leu											
		195													

<210> SEQ ID NO 23

<211> LENGTH: 3516

<212> TYPE: DNA

<213> ORGANISM: *Arabidopsis thaliana*

<400> SEQUENCE: 23

atggtttccg gagtcggggg tagtggcggg gcccggtggcg gtggccgtgg cggagaagaa 60

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gaaccgtcgt caagtcacac tcctaataac cgaagaggag gagaacaagc tcaatcgtcg	120
ggaacgaaat ctctcagacc aagaagcaac actgaatcaa tgagcaaagc aattcaacag	180
tacaccgtcg acgcaagact ccacgcggtt ttcgaacaat cgggcgaatc agggaaatca	240
ttcgactact cacaatcact caaaaacgacg acgtacgggtt cctctgtacc tgagcaacag	300
atcacagctt atctctctcg aatccagcga ggtggttaca ttcagccttt cggatgtatg	360
atcgccgtcg atgaatccag tttccggatc atcggttaca gtgaaaacgc cagagaaatg	420
ttagggatta tgcctcaatc tgttcctact cttgagaaac ctgagattct agctatggga	480
actgatgtga gatctttgtt cacttctctg agctcgattc tactcgagcg tgctttcgtt	540
gctcgagaga ttaccttggtt aaatccgggtt tggatccatt ccaagaatac tggtaaaccg	600
ttttacgcca ttcttcatag gattgatggt ggtggtggtta ttgatttaga gccagctaga	660
actgaagatc ctgctcttcc tattgctggt gctgttcaat cgcagaaact cgcggttcgt	720
gcgatttctc agttacaggc tcttcctggt ggagatatta agcttttctg tgacactgtc	780
gtggaagtgt tgagggactt gactggttat gatcgtggtta tggttataaa gtttcatgaa	840
gatgagcatg gagaagtgtt agctgagagt aaacgagatg atttagagcc ttatattgga	900
ctgcattatc ctgctactga tattcctcaa gcgtcaaggt tcttgtttaa gcagaacgt	960
gtccgaatga tagtagattg caatgccaca cctgttcttg tgggccagga cgataggcta	1020
actcagtcta tgtgcttggg tggttctact cttagggtctc ctcatggttg tcaactctcag	1080
tatatggcta acatgggcatc tattgctctt ttagcaatgg cggttataat caatggaat	1140
gaagatgatg ggagcaatgt agctagtgga agaagctcga tgaggctttg gggtttgggt	1200
gtttgccatc acacttcttc tcgctgcata ccgtttccgc taaggatgac ttgtgagttt	1260
ttgatgcagg ctcttcggtt acagttaaac atggaattgc agttagcttt gcaaatgtca	1320
gagaaaacgcg ttttgagaac gcagacactg ttatgtgata tgcctctcgc tgactcgctt	1380
gctggaattg ttacacagag tcccagatc atggacttag tgaaatgtga cgggtgcagca	1440
tttctttacc acgggaagta ttacccttg ggtggtgctc ctagtgaagt tcagataaaa	1500
gatggtgtgg agtggttgct tgcgaatcat gcggattcaa cgggattaag cactgatagt	1560
ttaggcgatg cggggatcc cgggtcagct gcgtagggg atgctgtgtg cggtatggca	1620
gttgcatata tcacaaaaag agactttctt ttttggttc gatctcacac tgcgaaagaa	1680
atcaaatggg gaggcgctaa gcatcatccg gaggataaag atgatgggca acgaaatgcat	1740
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actgcgaaaa tggatgcgat tcaactcctc cagcttatto tgagagactc ttttaaagaa	1860
tctgagcggg ctatgaactc taaagttgtg gatggtgtgg ttcagccatg tagggatatg	1920
gcgggggaac aggggattga tgagttaggt gcagttgcaa gagagatggt taggctcatt	1980
gagactgcaa ctgttcctat attcgtctgtg gatgccggag gctgcatcaa tggatggaac	2040
gctaagattg cagagttgac aggtctctca gttgaagaag ctatggggaa gtctctgggt	2100
tctgatttaa tatacaaaga gaatgaagca actgtcaata agcttcttcc tcgtgctttg	2160
agaggggacg aggaaaagaa tgtggaggtt aagctgaaaa ctttcagccc cgaactacaa	2220
gggaaagcag tttttgtggt tgtgaatgct tgttccagca aggactactt gaacaacatt	2280
gtcgcgcttt gttttgttgg acaagacggt actagtccaga aaatcgtaat ggataagttc	2340
atcaacatac aaggagatta caaggctatt gtacatagcc caaacctctt aatccgcca	2400

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atTTTTgctg ctgacgagaa cacgtgctgc ctggaatgga acatggcgat ggaaaagctt 2460
acgggttggt ctcgcagtga agtgattggg aaaatgattg tcggggaagt gtttgggagc 2520
tgttgcatgc taaaggggcc tgatgcttta accaagttca tgattgtatt gcataatgcg 2580
attggtggcc aagatacggg taagttccct ttcccattct ttgaccgcaa tgggaagttt 2640
gttcaggctc tattgactgc aaacaagcgg gtagcctcg agggaaaggt tattggggct 2700
ttctgtttct tgcaaatccc gagccctgag ctgcagcaag ctttagcagt ccaacggag 2760
caggacacag agtgtttcac gaaggcaaaa gagttggctt atatttgtca ggtgataaag 2820
aatcctttga gcggtatgcg tttcgcaaac tcattgttgg agggccacaga cttgaacgag 2880
gaccagaagc agttacttga aacaagtgtt tcttgcgaga aacagatctc aaggatcgtc 2940
ggggacatgg atcttgaagc cattgaagac ggttcatttg tgctaaagag ggaagagttt 3000
ttccttgtaa gtgcataaaa cgcgattgta agtcaagcga tgttcttatt aaggacaga 3060
ggtcttcagc tgatccgtga cattcccga gagatcaaat caatagaggt ttttgagac 3120
cagataagga ttcaacagct cctggctgag tttctgctga gtataatccg gtatgcacca 3180
tctcaagagt ggggtggagat ccatttaagc caacttcaa agcaaatggc tgatggattc 3240
gcccgcctcc gcacagaatt cagaatggcg tgtccaggtg aaggtctgcc tccagagcta 3300
gtccgagaca tgttccatag cagcaggtgg acaagccctg aaggtttagg tctaagcgt 3360
tgtcgaaaga ttttaaagct aatgaacggt gaggttcaat acatccgaga atcagaacgg 3420
tcctatttcc tcactattct ggaactccct gtacctgaa agcgaccatt gtcaactgct 3480
agtgaagtg gtgacatgat gctgatgatg ccatat 3516

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<210> SEQ ID NO 24

<211> LENGTH: 3483

<212> TYPE: DNA

<213> ORGANISM: Zea mays

<400> SEQUENCE: 24

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atggcgctcg gcagccgcgc cacgcccacg cgctccccct cctccgcgcg gcccgaggcg 60
ccgctgcacg cgcaccacca ccaccactcc cagtcgctcg gcgggagcac gtcccgcgcg 120
ggcgggggag ccgcgccac ggagtcggtc tccaaggccg tcgcccagta cacccatagc 180
gcgcgcctac acgcggtgtt cgagcaatcg ggcgcgctcg gccgcagctt cgaactctcc 240
caatcgctgc gcgcgcccgc cacgcccgtcc tccgagcagc agatcgccgc ctacctctcc 300
cgcatccagc gcgcgcccca catccagccc ttcggctgca cgctcgccgt cgcgacgac 360
tcctccttcc gctcctcgcg cttctccgag aactcccccg acctgctcga cctgtcgct 420
caccactccg ttccctcgct ggactcctct gcgcccgcc acgtttccct gggtgccgac 480
gcgcgcctcc tcttctcccc ctgctccgcg gtccctcctag agcgcgcctt cgcgcgcgcg 540
gagatctcgc tgctcaacc gatatggatc cactccaggg tctcctccaa gccgttctac 600
gccatcctcc accgcatcga cgtcggcgtc gtcacgacc tcgagcccgc ccgcaccgag 660
gaccccgctc tctccatcgc cggtcgagtc cagtcccaga aactggcggg ccgcgccatc 720
tcccgcctcc aggcgctacc cggcggggac gtcaagcttc tctgcgacac agtcgtggag 780
catgttcgcg agctcaeggg ttatgaccgt gtcattgggt acaggttcca tgaagacgag 840
cacggggaag ttgtgcgca gagccggcgc gacaaccttg agcctacct cggattgcat 900
tatcccgcca cagatatccc ccaggcgtcg cgcttccgtg tccggcagaa ccgcgtgca 960
atgattgccg attgcatgc caccggtg agagttattc aagatcctgg gctgtcgcag 1020

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cctctgtggt	tggtaggtc	cacgctacgc	gctccacacg	ggtgtcatgc	acagtacatg	1080
gcgaacatgg	ggtcaattgc	gtcgtctggt	atggcagtca	tcattagcag	tgccggtgac	1140
gatgagcaaa	caggtcgggg	tgcatctcgc	tcggcaatga	agttgtgggg	gtagtggtg	1200
tgccaccata	catcaccacg	gtgtatecct	ttccattga	ggtatgcttg	cgagtttctc	1260
atgcaggcat	ttgggttgca	gctcaacatg	gagttgcagc	ttgcgcacca	gctgtcagag	1320
aagcacatct	tgcaactca	gacgctattg	tgtgacatgc	tactacgaga	ttaccaact	1380
ggcatcgtca	cgcagagccc	cagcatcatg	gaccttgtga	agtgcgacgg	ggctgcactg	1440
tattatcatg	ggaaatacta	tccattgggt	gtcactccca	ctgagtctca	gattaaggat	1500
atcatcgagt	ggttgacggt	gtttcatggg	gactcaacag	ggctcagcac	agatagcctg	1560
gctgatgcag	gctaccttgg	tgtctgtgca	ctaggggagg	ctgtgtgtgg	aatggcggtg	1620
gcttatatta	caccgagtga	ttacttgttt	tggtttcggg	cacacacagc	taaagagatc	1680
aaatggggtg	gcgcaaaagca	tcaccctgag	gataaggatg	atggtcagag	gatgcacca	1740
cggctcgtcat	tcaaggcatt	tcttgaagtg	gttaaaagca	gaagcctgcc	atgggagaat	1800
gcagaaatgg	acgcaataca	ttccttgacg	ctcatattgc	gtgactcctt	cagggatgct	1860
gcagagggca	ccaacaactc	aaaagccatt	gtcaatggac	aagttcagct	tcgggagcta	1920
gaattcgggg	ggataaatga	gcttagttcc	gtagcaagag	agatggttcg	gttgatagag	1980
acagcaacag	taccatatt	tgcagtagat	actgatgggt	gtataaatgg	ttggaatgca	2040
aagattgctg	agttgacagg	gctttcagtt	gaggaggcaa	tgggcaaatc	tctggtaaat	2100
gatcttatct	tcaaggaatc	tgaggcgaca	gttgaaaaac	tactctcagc	agctttaaga	2160
ggtgaggaag	acaaaaatgt	ggagataaag	ttgaagacat	ttgggtcaga	gcaatataag	2220
ggaccaatat	ttgttgtgt	caatgcttgt	tctagtagag	attacacaca	aaatattgta	2280
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aacatacaag	gggactacaa	agctattgta	cacaatccta	atcctctgat	accaccaatt	2400
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tgtcgactca	agggcccaga	tgcattgaca	aaattcatgg	ttattattca	caacgctata	2580
ggaggacagg	attatgagaa	gttccctttt	tcattttttg	acaagaatgg	aaagtatgtg	2640
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cagaggcagt	tccttgaaac	tagctctgct	tgtgagaaac	agatgtccaa	gattgttaag	2940
gacgccagtc	tccaaagtat	cgaggacggc	tctttggtgc	ttgagcaaaag	tgagttttct	3000
cttgagagcg	ttatgaatgc	tgttgtcagc	caagcaatgt	tattgttgag	agagagggat	3060
ttacaactta	ttcgggacat	ccctgatgaa	atcaaggatg	cgtcagcgta	tggtgatcaa	3120
tgtagaatc	aacaagtgtt	ggctgacttc	ttgctaagca	tggtgcggtc	tgctccatcc	3180
gagaatggtt	gggtagaat	acaagtcaga	ccaaatgtaa	aacagaatc	tgatggaaca	3240
aatacagaac	tttctatatt	caggtttgcc	tgccttggtg	agggcctccc	tgctgacgtc	3300
gtccaggata	tgttcagcaa	ttcccaatgg	tcaacacaag	aaggcgtagg	actaagcaca	3360

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tgcaggaaga tctcaaat gatgggtggc gaggtccaat acatcagaga gtcagagcgg	3420
agtttcttcc tcatcgctct cgagcagccc caacctcgtc cagcagctgg tagagaaatc	3480
gtc	3483

<210> SEQ ID NO 25
 <211> LENGTH: 3513
 <212> TYPE: DNA
 <213> ORGANISM: *Oryza sativa*

<400> SEQUENCE: 25

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cccgccacc agcaccacca ctgcagctcc tgggggggga gcacgtcccg cgcgggaggg	120
ggtggcgggg gcgggggagg gggagggggc ggcgcgcccg ccgcgagctc ggtgtccaag	180
gccgtggcgc agtacacct ggacgcgcgc ctccacgcgg tgttcgagca gtcgggcgcg	240
tggggccgca gcttcgacta cagcagctcg ctgcgtgctt ccccccccc gtctccgag	300
cagcagatcg ccgcctacct ctccgcctc cagcgcggcg ggcacataca gcccttcggc	360
tgacgcctcg ccgtgcgca cgaactctcc ttccgcctcc tgcctactc cgagaacacc	420
gccgacctgc tcgacctgtc gccccaccac tccgtccct cgttcgactc ctccgcggtg	480
ctccccccg tctcgtcggc cgcagacgcg cgcctcctt tgcctccctc gtcgcccgtc	540
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tctcagaagc tcgcggtccg tgccatctcc cgcctccagg cgttccccg cggtgacgtc	780
aagctccttt gcgacaccgt tgttgagcat gttagagagc tcacaggtta tgaccgcgtt	840
atggtgtaca ggttccatga ggatgagcat ggagaagtcg ttgccgagag ccggcgcagt	900
aaccttgagc cctacatcgg gttgcattat cctgctacag atatcccaca ggcacacgc	960
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<210> SEQ ID NO 26

<211> LENGTH: 3534

<212> TYPE: DNA

<213> ORGANISM: Sorghum bicolor

<400> SEQUENCE: 26

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cgcgcggggc ggggaggtgg aggagggagg ggtggcgggg gcaccgcggc cacggctacg 180
gccacggcca cggagtcggt ctccaaggcc gtggcgcagt acaccctaga cgcgcggctc 240
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cgcgcgggcc acatccagcc cttcggtgac acgctcgcg tcgcccagca ctctccttc 420
cgcctcctcg cttctcega gaacgcgcgc gacctgctcg acctgtcgcg gcaccactcc 480
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ctcttctccc cctcgtccgc ggtcctcctg gagegcgcct tcgccgcgcg cgagatctcg	600
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tgttatgcaa ggatgaaaga attggcctat atttgccagg agataaagaa tcctcttagt	2880
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<210> SEQ ID NO 27

<211> LENGTH: 3411

<212> TYPE: DNA

<213> ORGANISM: Glycine max

<400> SEQUENCE: 27

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gccatcgcgc agtacacgga ggacgcgcgg ctccacgccc tcttcgagca gtccggcgag 180
tcgggagggt ccttcaacta ctccgaatca atccgcatcg catcggaatc cgtccccgag 240
cagcagataa cggttaacct tgtcaaaatc cagcgcggcg gcttcattca gcccttcggc 300
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<210> SEQ ID NO 28

<211> LENGTH: 3447

<212> TYPE: DNA

<213> ORGANISM: Glycine max

<400> SEQUENCE: 28

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aacaacaaca acaacaacat cgactccagc agcaaggcca tcgcgcagta cacggaggac 180
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<210> SEQ ID NO 29

<211> LENGTH: 3300

<212> TYPE: DNA

<213> ORGANISM: Glycine max

<400> SEQUENCE: 29

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<210> SEQ ID NO 30

<211> LENGTH: 2601

<212> TYPE: DNA

<213> ORGANISM: Glycine max

<400> SEQUENCE: 30

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<210> SEQ ID NO 31
<211> LENGTH: 3390
<212> TYPE: DNA
<213> ORGANISM: Solanum tuberosum

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<400> SEQUENCE: 31

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<210> SEQ ID NO 32
<211> LENGTH: 3363
<212> TYPE: DNA
<213> ORGANISM: Pisum sativum
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (84)..(84)
<223> OTHER INFORMATION: n is a, c, g, or t

<400> SEQUENCE: 32

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<210> SEQ ID NO 33

<211> LENGTH: 3387

<212> TYPE: DNA

<213> ORGANISM: Vitis vinifera

<400> SEQUENCE: 33

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gctcgctcc acgccgtata ogaacagtcc ggcgagtcg gtaagtcatt cgactactcg	180
cagtcgggta gaaccacaac gcaatcggtc cctgagcaac aatcactgc gtatttatcg	240
aaaattcaac ggggtggcca tatacagccc tttgggtgta tgcttgccgt cgatgagggc	300
acttttcggg tcattgcttt cagcgaat gcccagaaa tgctcggtct cactccgcaa	360
tcggttccga gccttgagaa gcccagatc ctctagtag gtactgatgt tcgcacgctt	420
ttcactcctc cgagcgcagt tctcctcgaa aaggcgttcc gggctcggga aattacgttg	480
ttaatcccg tgtggattca ttccaagaat tctgaaaa ccttttacgc aattttgcat	540
agaattgatg tgggaattgt aattgattg gacccctgcaa ggactgagga ccctgctctg	600
tccattgctg gggcgggca gtcgcagaag ttggccgttc gagcaatttc ccatcttcaa	660
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cttactgggt atgatcgggt catggtttac aaatttcacg aggatgaaca tgggtgaggtc	780
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tgccatgcca cgcctgttct ggtgattcaa gatgaagggc ttatgcagcc tctatgctta	960
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catgcggatt caacaggttt aagcactgac agtttggctg atgctggcta ccctggggca	1500
gcctcacttg gtgatgcagt ttgtggaatg gctgttgcct atatcacttc aagagatttt	1560
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ccagaggaca aggcagatgg gcagaggatg catcctcgtt cttcattcaa ggcattttta 1680
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<210> SEQ ID NO 34

<211> LENGTH: 187

<212> TYPE: PRT

<213> ORGANISM: Synechocystis PCC6803

<400> SEQUENCE: 34

```

Leu Gly Phe Tyr His Met Ala Asn Ala Ala Leu Asn Arg Leu Arg Gln
1           5           10           15
Gln Ala Asn Leu Arg Asp Phe Tyr Asp Val Ile Val Glu Glu Val Arg
20           25           30
Arg Met Thr Gly Phe Asp Arg Val Met Leu Tyr Arg Phe Asp Glu Asn
35           40           45
Asn His Gly Asp Val Ile Ala Glu Asp Lys Arg Asp Asp Met Glu Pro

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-continued

50					55					60					
Tyr	Leu	Gly	Leu	His	Tyr	Pro	Glu	Ser	Asp	Ile	Pro	Gln	Pro	Ala	Arg
65				70					75						80
Arg	Leu	Phe	Ile	His	Asn	Pro	Ile	Arg	Val	Ile	Pro	Asp	Val	Tyr	Gly
			85					90					95		
Val	Ala	Val	Pro	Leu	Thr	Pro	Ala	Val	Asn	Pro	Ser	Thr	Asn	Arg	Ala
			100					105					110		
Val	Asp	Leu	Thr	Glu	Ser	Ile	Leu	Arg	Ser	Ala	Tyr	His	Cys	His	Leu
		115					120					125			
Thr	Tyr	Leu	Lys	Asn	Met	Gly	Val	Gly	Ala	Ser	Leu	Thr	Ile	Ser	Leu
		130				135					140				
Ile	Lys	Asp	Gly	His	Leu	Trp	Gly	Leu	Ile	Ala	Cys	His	His	Gln	Thr
145				150					155						160
Pro	Lys	Val	Ile	Pro	Phe	Glu	Leu	Arg	Lys	Ala	Cys	Glu	Phe	Phe	Gly
				165				170							175
Arg	Val	Val	Phe	Ser	Asn	Ile	Ser	Ala	Gln	Glu					
			180				185								

<210> SEQ ID NO 35

<211> LENGTH: 187

<212> TYPE: PRT

<213> ORGANISM: Deinococcus radiodurans

<400> SEQUENCE: 35

Ser	Thr	Gly	Pro	His	Ala	Leu	Arg	Asn	Ala	Met	Phe	Ala	Leu	Glu	Ser
1				5					10					15	
Ala	Pro	Asn	Leu	Arg	Ala	Leu	Ala	Glu	Val	Ala	Thr	Gln	Thr	Val	Arg
			20					25					30		
Glu	Leu	Thr	Gly	Phe	Asp	Arg	Val	Met	Leu	Tyr	Lys	Phe	Ala	Pro	Asp
		35				40						45			
Ala	Thr	Gly	Glu	Val	Ile	Ala	Glu	Ala	Arg	Arg	Glu	Gly	Leu	His	Ala
		50				55					60				
Phe	Leu	Gly	His	Arg	Phe	Pro	Ala	Ser	Asp	Ile	Pro	Ala	Gln	Ala	Arg
65				70					75						80
Ala	Leu	Tyr	Thr	Arg	His	Leu	Leu	Arg	Leu	Thr	Ala	Asp	Thr	Arg	Ala
			85					90						95	
Ala	Ala	Val	Pro	Leu	Asp	Pro	Val	Leu	Asn	Pro	Gln	Thr	Asn	Ala	Pro
			100					105					110		
Thr	Pro	Leu	Gly	Gly	Ala	Val	Leu	Arg	Ala	Thr	Ser	Pro	Met	His	Met
		115					120					125			
Gln	Tyr	Leu	Arg	Asn	Met	Gly	Val	Gly	Ser	Ser	Leu	Ser	Val	Ser	Val
		130				135					140				
Val	Val	Gly	Gly	Gln	Leu	Trp	Gly	Leu	Ile	Ala	Cys	His	His	Gln	Thr
145				150					155						160
Pro	Tyr	Val	Leu	Pro	Pro	Asp	Leu	Arg	Thr	Thr	Leu	Glu	Tyr	Leu	Gly
				165				170							175
Arg	Leu	Leu	Ser	Leu	Gln	Val	Gln	Val	Lys	Glu					
			180				185								

<210> SEQ ID NO 36

<211> LENGTH: 187

<212> TYPE: PRT

<213> ORGANISM: Pseudomonas aeruginosa

<400> SEQUENCE: 36

Thr Ser Phe Thr Leu Asn Ala Gln Arg Ile Ile Ala Gln Val Gln Leu

-continued

1	5	10	15
His Asn Asp Thr Ala Ser Leu Leu Ser Asn Val Thr Asp Glu Leu Arg	20	25	30
Arg Met Thr Gly Tyr Asp Arg Val Met Ala Tyr Arg Phe Arg His Asp	35	40	45
Asp Ser Gly Glu Val Val Ala Glu Ser Arg Arg Glu Asp Leu Glu Ser	50	55	60
Tyr Leu Gly Gln Arg Tyr Pro Ala Ser Asp Ile Pro Ala Gln Ala Arg	65	70	75
Arg Leu Tyr Ile Gln Asn Pro Ile Arg Leu Ile Ala Asp Val Ala Tyr	85	90	95
Thr Pro Met Arg Val Phe Pro Ala Leu Asn Pro Glu Thr Asn Glu Ser	100	105	110
Phe Asp Leu Ser Tyr Ser Val Leu Arg Ser Val Ser Pro Ile His Cys	115	120	125
Glu Tyr Leu Thr Asn Met Gly Val Arg Ala Ser Met Ser Ile Ser Ile	130	135	140
Val Val Gly Gly Lys Leu Trp Gly Leu Phe Ser Cys His His Met Ser	145	150	155
Pro Lys Leu Ile Pro Tyr Pro Val Arg Met Ser Phe Gln Ile Phe Ser	165	170	175
Gln Val Cys Ser Ala Ile Val Glu Arg Leu Glu	180	185	

<210> SEQ ID NO 37

<211> LENGTH: 187

<212> TYPE: PRT

<213> ORGANISM: Rhodopseudomonas palustris

<400> SEQUENCE: 37

Asn Glu Phe Phe Arg Ser Val Arg Val Ala Ile Arg Arg Leu Gln Thr	1	5	10	15
Ala Ala Asp Leu Pro Thr Ala Cys Trp Ile Ala Ala Ser Glu Val Arg	20	25	30	
Arg Ile Thr Gly Phe Asp Arg Ile Lys Val Tyr Gln Phe Ala Ala Asp	35	40	45	
Trp Ser Gly Gln Val Ile Ala Glu Asp Arg Asp Ser Gly Ile Pro Ser	50	55	60	
Leu Leu Asp Phe His Phe Pro Ser Ser Asp Ile Pro Ala Gln Ser Arg	65	70	75	80
Ala Leu Tyr Thr Ile Asn Pro Val Arg Ile Ile Pro Asp Ile Gly Tyr	85	90	95	
Arg Pro Ser Pro Leu Val Pro Asp Ile Asn Pro Arg Leu Gly Gly Pro	100	105	110	
Ile Asp Leu Ser Phe Ser Val Leu Arg Ser Val Ser Pro Thr His Leu	115	120	125	
Glu Tyr Met Val Asn Met Gly Met His Ala Ala Met Ser Ile Ser Ile	130	135	140	
Val Arg Asp Asn Arg Leu Trp Gly Met Ile Ser Cys His Asn Leu Thr	145	150	155	160
Pro Arg Phe Val Ser Tyr Glu Val Arg Gln Ala Cys Glu Leu Ile Ala	165	170	175	
Gln Val Leu Thr Trp Gln Ile Gly Val Leu Glu	180	185		

-continued

<210> SEQ ID NO 38
 <211> LENGTH: 187
 <212> TYPE: PRT
 <213> ORGANISM: Synechococcus

<400> SEQUENCE: 38

```

Ser Arg Asp Ala Leu Ile Asn Arg Ile Thr His Gln Ile Arg Gln Ser
1           5           10           15
Leu Glu Leu Asp Gln Ile Leu Arg Ala Thr Val Glu Glu Val Arg Ala
20           25           30
Phe Leu Gly Thr Asp Arg Val Lys Val Tyr Arg Phe Asp Pro Glu Gly
35           40           45
His Gly Thr Val Val Ala Glu Ala Arg Gly Gly Glu Arg Leu Pro Ser
50           55           60
Leu Leu Gly Leu Thr Phe Pro Ala Gly Asp Ile Pro Glu Glu Ala Arg
65           70           75           80
Arg Leu Phe Arg Leu Ala Gln Val Arg Val Ile Val Asp Val Glu Ala
85           90           95
Gln Ser Arg Ser Ile Ser Gln Pro Glu Ser Trp Gly Leu Ser Ala Arg
100          105          110
Val Pro Leu Gly Glu Pro Leu Gln Arg Pro Val Asp Pro Cys His Val
115          120          125
His Tyr Leu Lys Ser Met Gly Val Ala Ser Ser Leu Val Val Pro Leu
130          135          140
Met His His Gln Glu Leu Trp Gly Leu Leu Val Ser His His Ala Glu
145          150          155          160
Pro Arg Pro Tyr Ser Gln Glu Glu Leu Gln Val Val Gln Leu Leu Ala
165          170          175
Asp Gln Val Ser Ile Ala Ile Ala Gln Ala Glu
180          185

```

<210> SEQ ID NO 39
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic Peptide

<400> SEQUENCE: 39

```

Lys Leu His His His His His His
1           5

```

<210> SEQ ID NO 40
 <211> LENGTH: 11
 <212> TYPE: PRT
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic Peptide

<400> SEQUENCE: 40

```

Gly Gly Gly Asp Tyr Lys Asp Asp Asp Asp Lys
1           5           10

```

<210> SEQ ID NO 41
 <211> LENGTH: 10
 <212> TYPE: PRT
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic Peptide

<400> SEQUENCE: 41

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Gly Gly Asp Tyr Lys Asp Asp Asp Asp Lys
1 5 10

<210> SEQ ID NO 42
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Synechocystis* PCC6803

<400> SEQUENCE: 42

Glu Ser Asp Ile Pro Gln
1 5

<210> SEQ ID NO 43
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Synechocystis* PCC6803

<400> SEQUENCE: 43

Pro Ile Arg Val Ile
1 5

<210> SEQ ID NO 44
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Synechocystis* PCC6803

<400> SEQUENCE: 44

Ile Leu Arg Ser Ala
1 5

<210> SEQ ID NO 45
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Synechocystis* PCC6803

<400> SEQUENCE: 45

Leu Thr Tyr Leu Lys
1 5

<210> SEQ ID NO 46
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Synechocystis* PCC6803

<400> SEQUENCE: 46

His Pro Arg Gln Ser Phe
1 5

<210> SEQ ID NO 47
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Deinococcus radiodurans*

<400> SEQUENCE: 47

Ala Ser Asp Ile Pro Ala
1 5

<210> SEQ ID NO 48
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Deinococcus radiodurans*

<400> SEQUENCE: 48

Leu Leu Arg Leu Thr

-continued

1 5

<210> SEQ ID NO 49
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Deinococcus radiodurans*

<400> SEQUENCE: 49

Val Leu Arg Ala Thr
1 5

<210> SEQ ID NO 50
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Deinococcus radiodurans*

<400> SEQUENCE: 50

Met Gln Tyr Leu Arg
1 5

<210> SEQ ID NO 51
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Deinococcus radiodurans*

<400> SEQUENCE: 51

Gly Pro Arg His Ser Phe
1 5

<210> SEQ ID NO 52
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 52

Ala Ser Asp Ile Pro Ala
1 5

<210> SEQ ID NO 53
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 53

Pro Ile Arg Leu Ile
1 5

<210> SEQ ID NO 54
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 54

Val Leu Arg Ser Val
1 5

<210> SEQ ID NO 55
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 55

Cys Glu Tyr Leu Thr
1 5

-continued

<210> SEQ ID NO 56
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 56

Thr Pro Arg Gly Ser Phe
1 5

<210> SEQ ID NO 57
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Rhodopseudomonas palustris*

<400> SEQUENCE: 57

Ser Ser Asp Ile Pro Ala
1 5

<210> SEQ ID NO 58
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Rhodopseudomonas palustris*

<400> SEQUENCE: 58

Pro Val Arg Ile Ile
1 5

<210> SEQ ID NO 59
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Rhodopseudomonas palustris*

<400> SEQUENCE: 59

Val Leu Arg Ser Val
1 5

<210> SEQ ID NO 60
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Rhodopseudomonas palustris*

<400> SEQUENCE: 60

Leu Glu Tyr Met Val
1 5

<210> SEQ ID NO 61
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Rhodopseudomonas palustris*

<400> SEQUENCE: 61

Gln Thr Arg Ala Ser Phe
1 5

<210> SEQ ID NO 62
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Synechococcus OS-B*

<400> SEQUENCE: 62

Ala Gly Asp Ile Pro Glu
1 5

<210> SEQ ID NO 63

-continued

<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Synechococcus OS-B*

<400> SEQUENCE: 63

Gln Val Arg Val Ile
1 5

<210> SEQ ID NO 64
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Synechococcus OS-B*

<400> SEQUENCE: 64

Leu Gln Arg Pro Val
1 5

<210> SEQ ID NO 65
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Synechococcus OS-B*

<400> SEQUENCE: 65

Val His Tyr Leu Lys
1 5

<210> SEQ ID NO 66
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Synechococcus OS-B*

<400> SEQUENCE: 66

Leu Pro Leu Ile Ser Phe
1 5

<210> SEQ ID NO 67
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: *Arabidopsis thaliana*

<400> SEQUENCE: 67

Ala Thr Asp Ile Pro Gln
1 5

<210> SEQ ID NO 68
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Arabidopsis thaliana*

<400> SEQUENCE: 68

Lys Val Arg Met Ile
1 5

<210> SEQ ID NO 69
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: *Arabidopsis thaliana*

<400> SEQUENCE: 69

Arg Val Arg Met Ile
1 5

<210> SEQ ID NO 70
<211> LENGTH: 5
<212> TYPE: PRT

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<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 70

Thr Leu Arg Ala Pro
1 5

<210> SEQ ID NO 71

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 71

Leu Gln Tyr Met Ala
1 5

<210> SEQ ID NO 72

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 72

Ser Gln Tyr Met Ala
1 5

<210> SEQ ID NO 73

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 73

Ala Gln Tyr Met Ser
1 5

<210> SEQ ID NO 74

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 74

Ala Gln Tyr Met Thr
1 5

<210> SEQ ID NO 75

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 75

Thr Gln Tyr Met Ala
1 5

<210> SEQ ID NO 76

<211> LENGTH: 6

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 76

His Pro Arg Ser Ser Phe
1 5

<210> SEQ ID NO 77

<211> LENGTH: 6

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 77

Asn Pro Arg Ser Ser Phe
1 5

What is claimed is:

1. An isolated polynucleotide comprising a contiguous coding sequence encoding a polypeptide having at least 95% identity to at least one amino acid sequence selected from SEQ ID NOs: 1-22, and having an amino acid other than tyrosine at the position corresponding to Y361 of SEQ ID NO:1, wherein the polypeptide confers increased light sensitivity in a plant expressing the polypeptide relative to a control plant lacking the polypeptide.
2. A vector comprising the polynucleotide of claim 1.
3. A polynucleotide construct comprising a promoter not natively associated with the polynucleotide of claim 1 operably linked to the polynucleotide of claim 1.
4. A plant cell comprising the polynucleotide of claim 1 operably linked to a promoter not natively associated with the polynucleotide of claim 1.
5. A plant comprising the plant cell of claim 4.
6. The plant of claim 5, wherein the plant exhibits increased light sensitivity relative to a control plant lacking the polynucleotide.
7. The plant of claim 5, wherein the plant exhibits a decreased height, decreased diameter or a combination thereof relative to a control plant lacking the polynucleotide.
8. The plant of claim 5, wherein the plant exhibits at least one characteristic selected from, increased hyponasty, decreased petiole length, decreased internode length, and decreased hypocotyl length under an R fluence rate of less than $1 \mu\text{mole m}^{-2} \text{sec}^{-1}$, relative to a control plant lacking the polynucleotide.
9. The plant of claim 5, wherein the plant exhibits enhanced germination relative to the control plant.
10. The plant of claim 9, wherein the plant is corn, soybean or rice.
11. The plant of claim 9, wherein the plant is an ornamental plant.
12. A method of producing a transgenic plant comprising:
 - (a) introducing into a plant cell a polynucleotide encoding a polypeptide comprising an amino acid sequence having at least 95% identity to at least one amino acid sequence selected from SEQ ID NOs: 1-22 and having an amino acid other than tyrosine at the position corresponding to Y361 of SEQ ID NO:1, wherein the polypeptide confers increased light sensitivity in a plant expressing the polypeptide relative to a control plant lacking the polypeptide; and
 - (b) regenerating the transformed cell to produce a transgenic plant.
13. The method of claim 12, wherein the transgenic plant exhibits increased light sensitivity relative to a control plant lacking the polynucleotide.
14. The method of claim 13, wherein the transgenic plant exhibits decreased height, decreased diameter, or a combination thereof relative to a control plant lacking the polynucleotide.
15. The method of claim 13, wherein the transgenic plant exhibits at least one characteristic selected from decreased petiole length, decreased internode number, increased hyponasty, and decreased hypocotyl length under an R fluence rate of less than $1 \mu\text{mole m}^{-2} \text{sec}^{-1}$, relative to a control plant lacking the polynucleotide.
16. The method of claim 12, wherein the transgenic plant exhibits enhanced germination relative to the control plant.
17. The method of claim 16, wherein the transgenic plant is a corn, soybean or rice plant.
18. The method of claim 16, wherein the transgenic plant is an ornamental plant.
19. A transgenic plant produced by the method of claim 12.
20. An isolated polypeptide comprising an amino acid sequence having at least 95% identity to at least one amino acid sequence selected from SEQ ID NOs: 1-22, and having an amino acid other than tyrosine at the position corresponding to Y361 of SEQ ID NO:1, wherein the polypeptide confers increased light sensitivity in a plant expressing the polypeptide relative to a control plant lacking the polypeptide.
21. The isolated polynucleotide of claim 1, further comprising at least one of (i) an amino acid other than aspartate (D) at the position corresponding to 307 of SEQ ID NO:1, (ii) an amino acid other than arginine (R) at the position corresponding to 322 of SEQ ID NO: 1, (iii) an amino acid other than arginine (R) at the position corresponding to 352 of SEQ ID NO: 1, and (iv) an amino acid other than arginine (R) at the position corresponding to 582 of SEQ ID NO: 1.

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