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(54) **EXTENDING JUVENILITY IN GRASSES**

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C12N 15/29 (2006.01)
C12N 15/82 (2006.01)

(52) **U.S. Cl.**
CPC **C12N 15/8266** (2013.01); **C12N 15/8261** (2013.01)

(58) **Field of Classification Search**
None
See application file for complete search history.

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(57) **ABSTRACT**

The present invention relates to compositions and methods for modulating the juvenile to adult developmental growth transition in plants, such as grasses (e.g. maize). In particular, the invention provides methods for enhancing agronomic properties in plants by modulating expression of GRMZM2G362718, GRMZM2G096016, or homologs thereof. Modulation of expression of one or more additional genes which affect juvenile to adult developmental growth transition such as Glossy15 or Cg1, in conjunction with such modulation of expression is also contemplated. Nucleic acid constructs for down-regulation of GRMZM2G362718 and/or GRMZM2G096016 are also contemplated, as are transgenic plants and products produced there from, that demonstrate altered, such as extended juvenile growth, and display associated phenotypes such as enhanced yield, improved digestibility, and increased disease resistance. Plants described herein may be used, for example, as improved forage or feed crops or in biofuel production.

45 Claims, 12 Drawing Sheets

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FIG. 1.



FIG. 2 (continued)

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AT5G55390.1          PEVLSQILEWKEKLEKLVYLAFFLHGARYTSFGRHF TWPKEKQQIVGRLHWYADDGGHIVDF
GRMZM2G362718__P01  I-----AVDKYVWGRGKTQ--EDYTRKEAAQRK-D---SS-----E
                        :  :!*  *  .  .!*  *  *!  *
AT5G55390.1          CCGSNDPFCLELHNAKLEETGRKCLYENYDLFFAKNNFNFEREDWHTVSKDELEFGSKL-----
GRMZM2G362718__P01  NQGQNDALELBNLRH-----EMQADEPFLPFGNKRDEK
                        *. *  *  *  :  *  .  .  *  *  *  *
AT5G55390.1          -----INGLNPPFGVNASLANKFITEKALEFRPKILILIVPPETERLDEKRSYVLIUEDRT
GRMZM2G362718__P01  WQHWVYGLGSAAGQMETLSRRE-----NPRSR-----GNVHSMW-----
                        :  **  *  :  !*  :  *  :  *  :  *  :
AT5G55390.1          PLSGNSPYLPGSVNEEDKQLEDWNLVFPPLSLWRSDFAAKHKKIAEKHCHLSRDVGSKK
GRMZM2G362718__P01  ---SKIIYYPRGCTEVN--VDD-----HPLEKQDHQDTESDGSKE--RRPVDMASCGNR
                        *  .  !*  *  .  !  :  !  *  *  .  .  *  !  .  *  !  .  *  .  !
AT5G55390.1          LRIVEEEMASLHPLGASDGWCCDIPMEKDELEVAECVNKILVSEKIDTIVETVAVRHQSD
GRMZM2G362718__P01  -PYLDENKRNLPEDGPA--HYEDNRSEPT--AADTEGYKQSE-EKPVWNTNRTGSR
                        (:  :  :  *  *  *  :  *  *  :  .  *  *  .  *  *  :  *  .  :
AT5G55390.1          HLSRPSQLKREGETDYSGRKLGKSMDSNNVDNKSMDMEEDQCELSAPESIKVRIPEHT
GRMZM2G362718__P01  HSLDRQRIE---GDSYRG---TYMNRQRHEWLHPHAGNSSRIG-----WDR
                        *  *  .  !  :  .  *  *  .  .  :  .  !  *  .  .  .  .  .  .  :
AT5G55390.1          SDWQSPVRESPPDDIYAVCTSISTTTPQRSHEAVEASLPAITRTRKSNLGNIREHOCKVQG
GRMZM2G362718__P01  RQWSSSPSFPFSAEPGQDESCSEANPFEGK-----YRTGGRHHRFQYLG
                        (:  *  *  *  .  .  *  *  :  *  *  *  :  *  .  .  :  *
AT5G55390.1          TGKPEVSRDRPSEVETSREDIYTV-----RPSFENTG--QEPFEAFEPSTYQASLSHFDDG
GRMZM2G362718__P01  LGTPQHGTFRPHHTGWRDIFHDHCHGRFPFHHTGWRDRAFFRDH-----QHGEYDDE
                        *. *  .  .  *  *  .  .  *  :  *  *  .  .  *  :  *  .  .  .  :  *  *
AT5G55390.1          LAARYGGFGGGYRHFDFPFLFDQFFLENGPNEMFDFRGYSDLDRGIGQREYFQQYGGHLD
GRMZM2G362718__P01  RYGEYDATINGPDSAHRPYTAAGVAGRSAPS YQL-AGGYG-----EGSRNR-----
                        .  !  *  .  *  .  *  :  .  *  .  *  :  *  .  *  :
AT5G55390.1          FMLAPPPFNLMDNAFFLQRYAPHFQMN YQRMSSFPQFPPLQPSGHNLLNPHDFPLPF
GRMZM2G362718__P01  -----FVTDKYAP-----WPLP-
                        *  :  !  *  *  *  :  *  *  *
AT5G55390.1          PPSDFEMSPRGFAPGFNPNYPYMSRSGGWIND
GRMZM2G362718__P01  -----

```


FIG. 3 (continued)

Sorghum Sb02g003420.1_Sb02g003430.1	GRRR-----KNIDQSTPEL---SNRLYGAESEQADNVGAKSTPQIVWEPHCAAKVLK
Maize GRM2M2G362718_P01	GRRRKRRKNTDQSTPEL---PNRLCGAESEQADNVGAKSTLQIVWEPHCAAKHLK
Rice LOC_Os08g24946.1 13108.m23057 protein	DRSY-----VSEPLQRAKLNKFNKAGKSKAGVKSEFEVLESEKKTRSLK
	..* : * :*:::*.** : : : : : **
Sorghum Sb02g003420.1_Sb02g003430.1	GPQIEQSIIGV---AGSONGAEITMGHEKQFG-----IS-CVARTETEKRVTY
Maize GRM2M2G362718_P01	GPQIAK--QGV---AARQWGAETMGHENQFG-----ISFCVASTETEKRVTC
Rice LOC_Os08g24946.1 13108.m23057 protein	KRTQPEEPLVECAAAAAMNMRPVREKELGTSSLDMKRIPLSSFPVIVSETEKRISA
	* : * :*. . . : :*:::* * : : :*:::***
Sorghum Sb02g003420.1_Sb02g003430.1	LAQKG-----TCLGTPYDGPSTKMDSCSVQDTPVD-----KDFEL
Maize GRM2M2G362718_P01	LAQFG-----TCLGTQYDGPSTKMGWYDCSVQDTPMD-----DDVEL
Rice LOC_Os08g24946.1 13108.m23057 protein	LVEKEVSLTIVADISRRCVIPSTYACSGRODKIVVRGKLEERSIQAVKAALQKLENGGAV
	* : : : * * * : : : * : * : : * : * : : * : * : : *
Sorghum Sb02g003420.1_Sb02g003430.1	D--NVAYR-----IMEDKYANGREET--QEDYTRKETAHKRDSSENQGGQ
Maize GRM2M2G362718_P01	D--NVACI-----IAYDKYVNGRGKT--QEDYTRKEAAQRDSSENQGGQ
Rice LOC_Os08g24946.1 13108.m23057 protein	DDAKAVGESEVLRQLTRMHNKLRVYLAPF IHGMRVYTSFGRHFTKREK-----
	* : : : : : * : * * : : * * : : * : : * : : * : : * : *
Sorghum Sb02g003420.1_Sb02g003430.1	DVLELD--NLWVEIQAD--GSPLEPGNKRYK--EEMAYGLCSASGHEKET--SSSRRENV
Maize GRM2M2G362718_P01	DALELD--NLRWEMQAD--ERPLEPGNKDRKQKRWYGLGASGGQKE---TLSRRENV
Rice LOC_Os08g24946.1 13108.m23057 protein	-LIEIAEKLHUVYVQPGDKMNVDPETPRR--VMNLRGFCALSQFMKEKLDKVGKRCNF
	:* : * : * : * : * : * : * : * : * : * : * : * : * : *
Sorghum Sb02g003420.1_Sb02g003430.1	QSDRGWVPMNSKTIYRKG--GTTLDNNVYDHS---SEGSYPCQGECS-----HSKCN----
Maize GRM2M2G362718_P01	RSDRGMVHSNSKTIYRKG--GTEVDNVD--DHP---LE-----
Rice LOC_Os08g24946.1 13108.m23057 protein	KMYD-VIQPKNS--FSFEKRDMMTVRQKELPHGSKLIMGLMNPFGPKAMLANKFIDKALT
	.. : : : * : * : : * : * : : * : * : : * : * : : *
Sorghum Sb02g003420.1_Sb02g003430.1	--DGLVALDQDTSDDLKRSQPVEKA-----SDGNKTDLDKKNKHNLKE-----D
Maize GRM2M2G362718_P01	----KODHQDTSDDGSKKRSPPVDNA-----SGGNRPYLDENKRNLR-----D
Rice LOC_Os08g24946.1 13108.m23057 protein	FKPKLIILIVPEAEERLDRKQOYDLVUEDDQRLSKSFLPLGSLDVSQDKQIDQMNKSP
 : : : * : * : * : * : * : * : * : * : *
Sorghum Sb02g003420.1_Sb02g003430.1	GR-DAHVEDRRTERNTAADTSRYKCRDKIQLDREPELVGRMTRASSSEHSPERGMNERD
Maize GRM2M2G362718_P01	GR-YAHVEDWRSERNTAADTSYKAOSE-----EKPVMWNTTCSREHSLDRQRIECG
Rice LOC_Os08g24946.1 13108.m23057 protein	PLYLWSRPDWTQKHKRLAEQCHTKANV--FSHNEEDLVLYLFEDRATQNHVDVNNKMYTSG
	* : : : * : * : * : * : * : * : * : * : * : * : * : *

FIG. 4

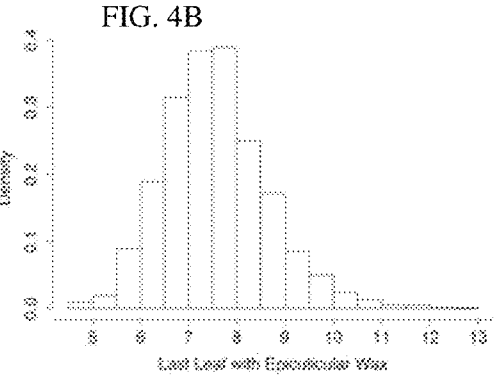
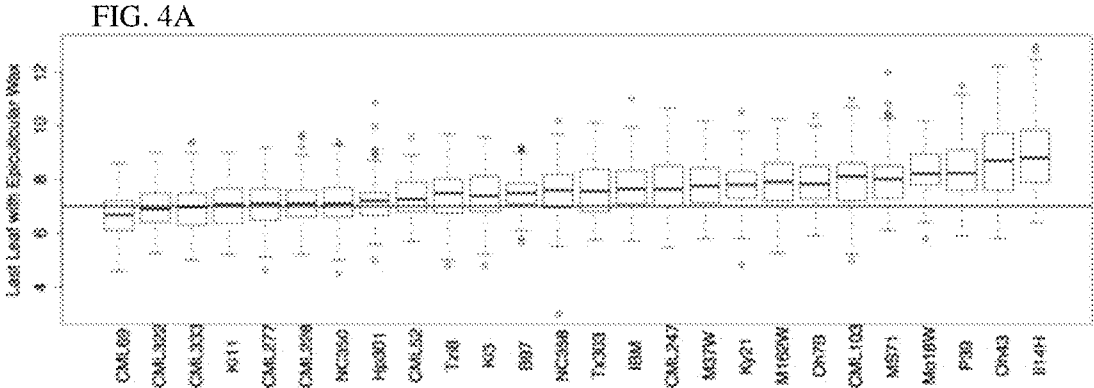


FIG. 4 (continued)

FIG. 4C

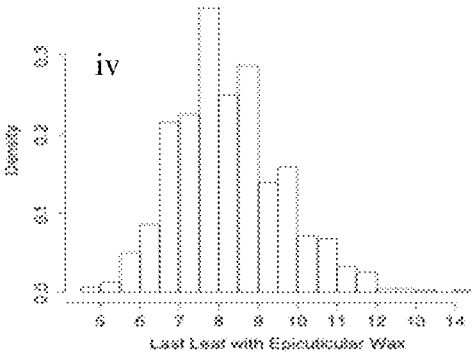
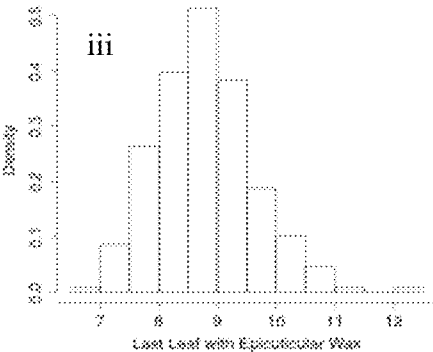
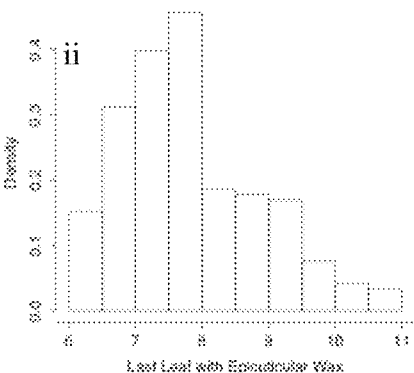
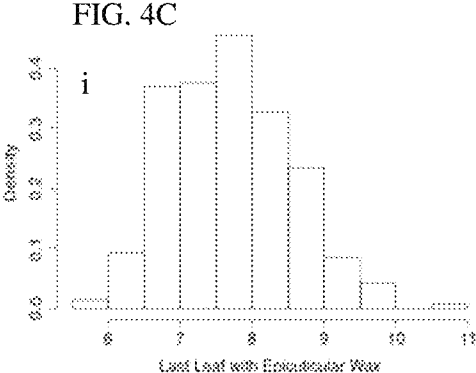


FIG. 5

	NAM		IBM		DyH		OWCI		WIDdy		CYRMIT
No. of lines	3875		302		243		277		573		509
Year ^{rep}	2008 ¹	2009 ¹	2008 ²	2009 ²	2010 ²	2011 ²	2010 ¹	2011 ¹	2009 ²	2010 ²	2010 ¹
Location	WM	ARL	WM	ARL	WM	WM	ARL	WM	ARL	WM	WM
			ARL	WM							
Plot size/ Plants	1/42		2/42		1/15		1/15		2/42		1/15
Traits Measured	Transition DAF GAS	Transition Node # Plant Height	Transition Node # Plant Height DAP		Transition		Transition		Transition Node # Plant Height GDD		Transition

FIG. 6

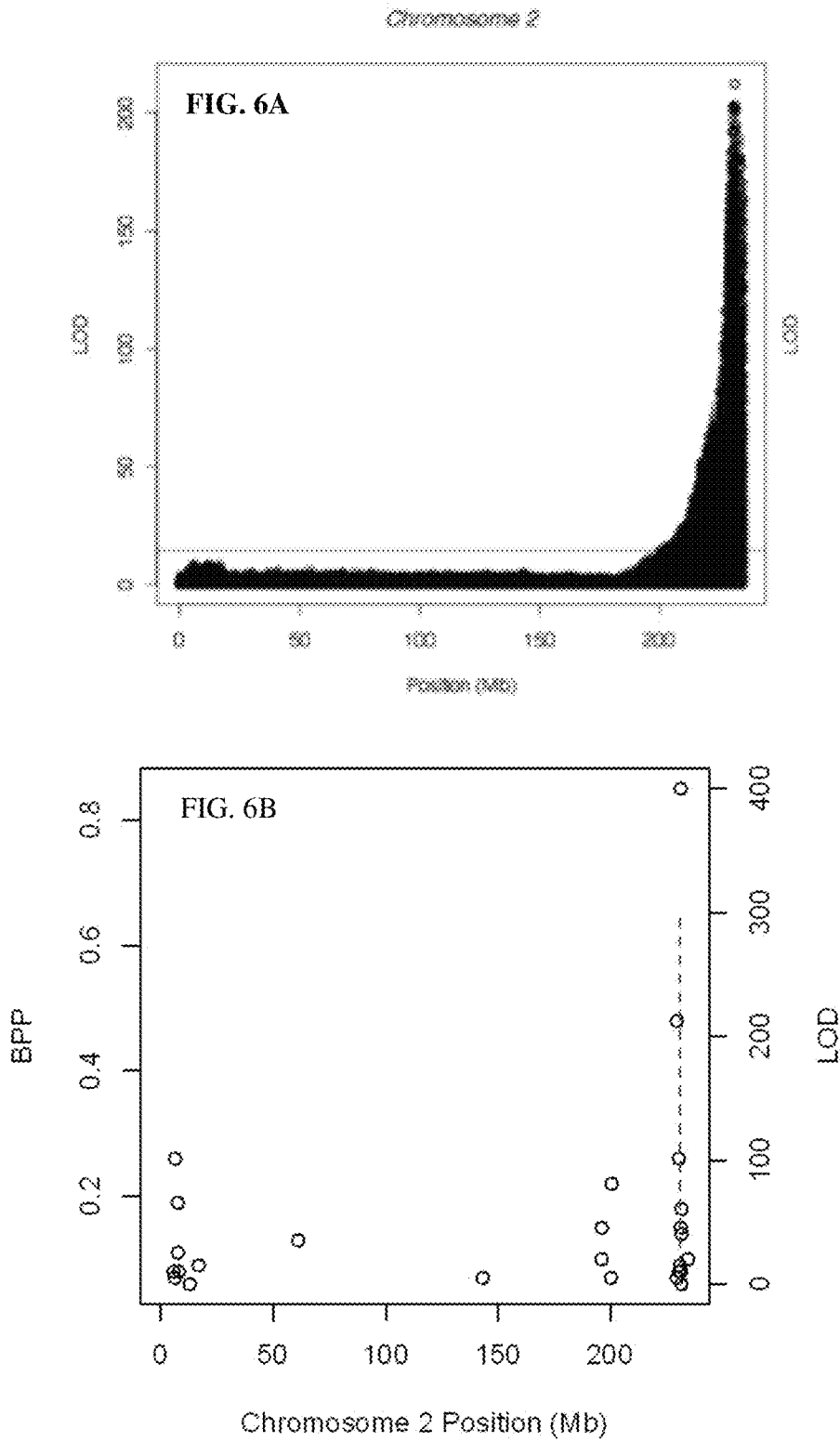


FIG. 7

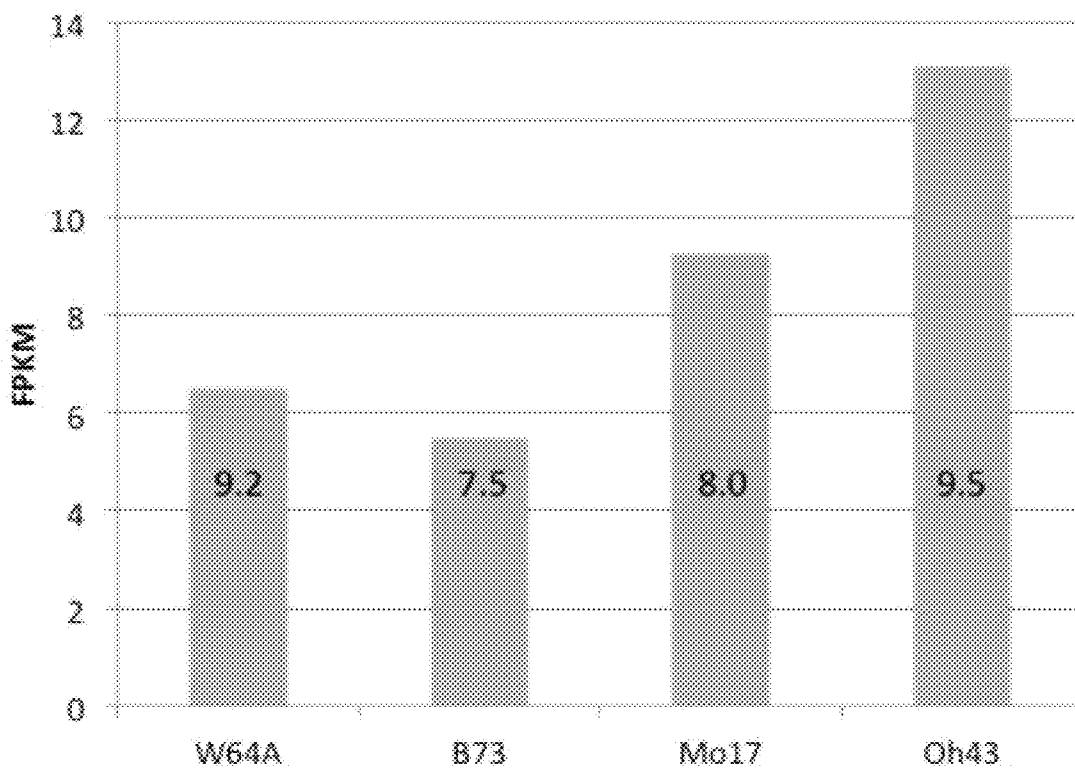
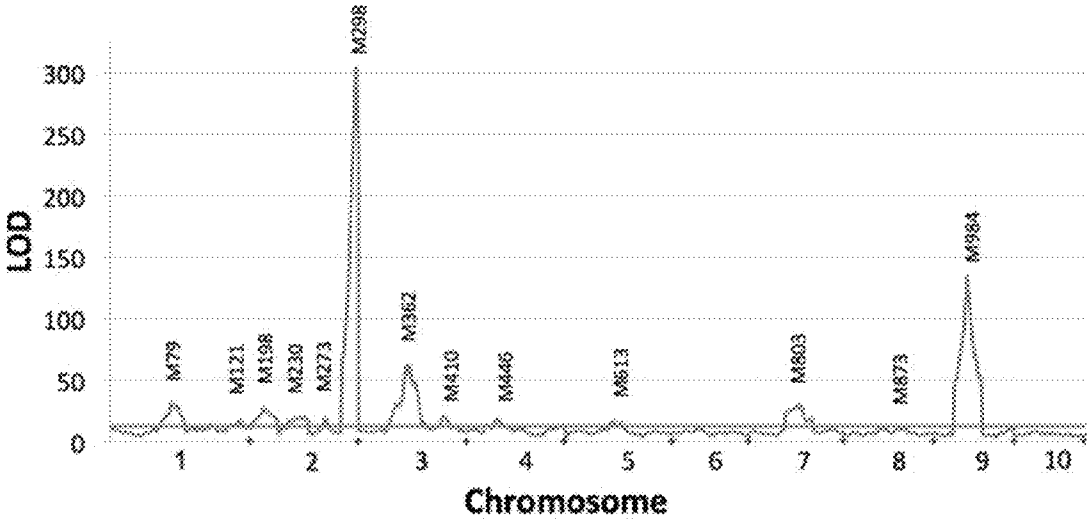


FIG. 8



EXTENDING JUVENILITY IN GRASSES

This application claims the priority of U.S. Provisional Appl. Ser. No. 61/651,540 filed May 24, 2012, the entire disclosure of which is incorporated herein by reference.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

This invention was made with Government support under DE-FC02-07ER64494 awarded by the U.S. Department of Energy. The Government has certain rights in the invention.

INCORPORATION OF SEQUENCE LISTING

The sequence listing that is contained in the file named "WARF103US updated ST25.txt", which is 122,753 bytes (measured in MS-WINDOWS) and was created on Sep. 18, 2013, is filed herewith by electronic submission and incorporated herein by reference.

FIELD OF THE INVENTION

The invention relates to methods and compositions for altering the juvenile phase of growth of plants.

BACKGROUND OF THE INVENTION

Juvenile and adult vegetative tissues in grasses differ dramatically in anatomy, biochemical composition, and in their ability to withstand biotic and abiotic stresses. Juvenile plants cannot flower and are capable of only vegetative growth. Juvenile leaf tissue further has inherent resistance to specific abiotic stresses such as cold and drought, is generally less recalcitrant when used for processing for biofuels, and may be more digestible when used as feed. Researchers have identified certain parameters such as age, leaf number, and certain growth conditions as playing a role in the maturation of juvenile plant tissue to adult plant tissue. However, the genetic triggers controlling the transition between juvenile and adult tissue in plants has not been well understood. Therefore, increasing the proportion of the plant that is juvenile has potential benefit for improving the yield and processing ability of plant biomass, among other agronomic traits.

SUMMARY OF THE INVENTION

In one aspect the invention provides a polynucleotide molecule comprising a sequence selected from the group consisting of: (a) a sequence encoding a polypeptide at least 85% identical to SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, or SEQ ID NO:17; wherein the polypeptide regulates juvenile to adult phase change in grass plant leaves; (b) a sequence comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, or SEQ ID NO:16; (c) a sequence hybridizing to (b) under wash conditions of 0.15 M NaCl and 70° C. for 10 minutes, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves; (d) a sequence comprising at least 85% sequence identity over its full length to the full length of SEQ ID NO:2 or SEQ ID NO:16, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves; and (e) a sequence complementary to (a), (b), (c) or (d), further wherein the polynucleotide molecule is operably linked to a heterologous promoter functional in plants. In a particular embodi-

ment the polynucleotide molecule comprises the coding sequence of SEQ ID NO:2. In another embodiment the polynucleotide molecule comprises the coding sequence of SEQ ID NO:16.

5 Other embodiments of the invention provide a recombinant vector comprising such a polynucleotide molecule. In certain embodiments, the invention provides the recombinant vector, further comprising an additional polynucleotide sequence which, after being transcribed, regulates the timing of the juvenile to adult phase change in a plant. Thus, in particular embodiments the recombinant vector may comprise an additional polynucleotide sequence which encodes all or part of a sequence selected from the group consisting of: Glossy15, Cg1, a homolog of either thereof, and/or a sequence complementary thereto.

10 In some embodiments the recombinant vector further comprises at least one additional sequence chosen from the group consisting of: a regulatory sequence such as a promoter, a selectable marker, a leader sequence and a terminator. The additional sequence may be a heterologous sequence. In some embodiments the promoter is a tissue-specific promoter. In a particular embodiment the promoter directs expression in leaf tissue. In certain embodiments the recombinant vector may be defined as an isolated expression cassette.

25 In other embodiments, the recombinant vector comprises a first sequence selected from the group consisting of: (a) a sequence encoding a polypeptide at least 85% identical to SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, or SEQ ID NO:17; wherein the polypeptide regulates juvenile to adult phase change in grass plant leaves; (b) a sequence comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8; or SEQ ID NO:16; (c) a sequence hybridizing to (b) under wash conditions of 0.15 M NaCl and 70° C. for 10 minutes, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves; (d) a sequence comprising at least 85% sequence identity over its full length to the full length of SEQ ID NO:2 or SEQ ID NO:16, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves; and (e) a sequence complementary to (a), (b), (c) or (d), or a fragment thereof; and a second sequence comprising the reverse complement of the first sequence, wherein the expression of the construct in a plant down regulates the expression of a coding sequence and/or encoded polypeptide in the plant. Some embodiments of the invention provide the recombinant vector further comprising an additional polynucleotide sequence which, after being transcribed, regulates the timing of the juvenile to adult phase change in a plant.

50 Another aspect of the invention is a transgenic plant or seed comprising a recombinant vector comprising a polynucleotide molecule comprising a sequence selected from the group consisting of: (a) a sequence encoding a polypeptide at least 85% identical to SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, or SEQ ID NO:17; wherein the polypeptide regulates juvenile to adult phase change in grass plant leaves; (b) a sequence comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, or SEQ ID NO:16; (c) a sequence hybridizing to (b) under wash conditions of 0.15 M NaCl and 70° C. for 10 minutes, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves; (d) a sequence comprising at least 85% sequence identity over its full length to the full length of SEQ ID NO:2 or SEQ ID NO:16, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves; and (e) a

sequence complementary to (a), (b), (c) or (d), further wherein the polynucleotide molecule is operably linked to a heterologous promoter functional in plants. In yet other embodiments, the transgenic plant may comprise a recombinant vector as described above, comprising an additional polynucleotide sequence which, after being transcribed, regulates the timing of the juvenile to adult phase change in the plant.

Yet another aspect of the invention is a transgenic plant or seed comprising a first sequence selected from the group consisting of: (a) a sequence encoding a polypeptide at least 85% identical to SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, or SEQ ID NO:17; wherein the polypeptide regulates juvenile to adult phase change in grass plant leaves; (b) a sequence comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, or SEQ ID NO:16; (c) a sequence hybridizing to (b) under wash conditions of 0.15 M NaCl and 70° C. for 10 minutes, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves; (d) a sequence comprising at least 85% sequence identity over its full length to the full length of SEQ ID NO:2 or SEQ ID NO:16, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves; and (e) a sequence complementary to (a), (b), (c) or (d), or a fragment thereof; and a second sequence comprising the reverse complement of the first sequence, wherein the expression of the construct in a plant down regulates the expression of a coding sequence and/or encoded polypeptide in the plant.

In some embodiments the transgenic plant may further be defined as a monocotyledonous plant. In particular embodiments the transgenic plant is further defined as a member of the Poaceae. In more particular embodiments the transgenic plant is further defined as a member of the Panicoideae or the Pooideae. In yet more particular embodiments the transgenic plant may further be defined as maize, rice, sorghum, or switchgrass.

The invention also provides a seed or cell of such a transgenic plant wherein the seed or cell comprises recombinant vector.

In certain embodiments the plant is a plant wherein the last leaf with epicuticular wax is produced later during plant development relative to that found in an otherwise isogenic plant lacking the recombinant vector.

In another aspect, the invention provides a method of altering the timing of juvenile to adult phase change in a plant, the method comprising modulating the expression of GRMZM2G362718 or GRMZM2G90616, or a homolog of either thereof, in the plant. Other contemplated embodiments of such methods further comprise modulating the expression of at least a second gene which regulates the timing of the juvenile to adult phase change in a plant. In particular embodiments the second gene is selected from the group consisting of Glossy15 and Cg1. Thus in some embodiments the method comprises expressing a recombinant vector or construct, as defined above, in the plant. In certain embodiments, the timing of the juvenile to adult phase change is extended (delayed) relative to a wild type plant (i.e. an otherwise essentially isogenic plant not comprising such a recombinant construct). In some embodiments the method comprises mutagenizing said GRMZM2G362718 or GRMZM2G90616 or a homolog thereof.

In certain embodiments of the method, the timing of juvenile to adult phase in the plant is extended relative to a wild type plant. In particular embodiments, the timing of

juvenile to adult phase change is calculated by a method comprising counting the last leaf displaying epicuticular wax.

In some embodiments of the method, the plant exhibits a trait selected from the group consisting of: an increase of at least one in the numbering of the last leaf which displays epicuticular wax or which does not contain abaxial trichomes; an altered proportion of juvenile, transitional, or adult leaves; enhanced yield of vegetative tissue; enhanced digestibility of vegetative tissue; enhanced resistance to a plant pest; and enhanced resistance to a plant disease. In certain embodiments of the method, the plant has altered development or morphology when compared to a wild type plant, further wherein the plant displays a trait selected from the group consisting of: enhanced disease resistance, enhanced insect resistance, improved forage digestibility, enhanced abiotic stress tolerance, and improved utility for biofuel production.

Yet another aspect of the invention provides a method of producing plant biomass, the method comprising: (a) obtaining a plant comprising a recombinant vector as described above; and (b) preparing biomass from said plant or a part thereof. In certain embodiments the method further comprises producing biofuel from the biomass. The method may also comprise producing food or feed from the biomass.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 depicts a maize transition leaf, with areas of juvenile tissue, and other areas of adult tissue.

FIG. 2 depicts a CLUSTAL protein alignment of GRMZM2G362718(SEQ ID NO:3) with *Arabidopsis* homolog AT5G55390.1(SEQ ID NO:10).

FIG. 3 shows a CLUSTAL protein alignment of GRMZM2G362718(SEQ ID NO:3) with homologs from sorghum(SEQ ID NO:19) and rice(SEQ ID NO:11).

FIG. 4 depicts bar plots showing variation of transition leaf numbering. (A) Variation for transition leaf by NAM family, labeled by the non-B7 parent on top. The horizontal line at leaf 7 represents the average transition leaf for B73; (B) Phenotypic distribution of the last leaf with epicuticular wax in the NAM population. Leaf number distribution ranged from leaf 4.5 to leaf 13.25; (C) Phenotypic variation for transition leaf in the IBM, NYH, OWRI, and Wisconsin diversity panel populations (plots i-iv, respectively).

FIG. 5 depicts a summary table of studies providing phenotypic data from defined mapping populations.

FIG. 6 shows genome wide association results with 1.6 million polymorphic markers across the NAM population. (A) Position of significant QTL found on the long arm of chromosome 2.; (B) sub sampling analysis confirming location of QTL on chromosome 2. Dashed line represents F-test log(1/P) in the final joint linkage model. Vertical position of points represents bootstrap posterior probability (BPP) of the SNP.

FIG. 7 depicts RNA sequence expression data of GRMZM2G362718 for four maize inbred lines that are parents of RIL mapping populations (Oh43×W64A; B73×Mo17; B73×Oh43). The inbred's transition phenotype is displayed numerically within the bar.

FIG. 8 depicts LOD scores for detecting the presence of QTL located on any of chromosomes 1-10. Stepwise regression with covariates was used in joint QTL mapping of all NAM populations with a threshold value of 12.26 (Buckler et al., *Science* 325:714-718, 2009).

BRIEF DESCRIPTION OF THE SEQUENCES

SEQ ID NO:1 GRMZM2G362718 genomic nucleotide sequence from *Z. mays* B73.

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SEQ ID NO:2 GRMZM2G362718 nucleotide coding sequence from *Z. mays* B73, with UTR.

SEQ ID NO:3 GRMZM2G362718 predicted protein sequence from *Z. mays* B73.

SEQ ID NO:4 GRMZM2G362718 nucleotide coding sequence from *Z. mays* Mo17.

SEQ ID NO:5 GRMZM2G362718 predicted protein sequence from *Z. mays* Mo17.

SEQ ID NO:6 GRMZM2G362718 nucleotide coding sequence from *Z. mays* Oh43.

SEQ ID NO:7 GRMZM2G362718 predicted protein sequence from *Z. mays* Oh43.

SEQ ID NO:8 GRMZM2G362718 nucleotide coding sequence from *Z. mays* W64A.

SEQ ID NO:9 GRMZM2G362718 predicted protein sequence from *Z. mays* W64A.

SEQ ID NO:10 Predicted protein sequence of AT5G55390.1 from *Arabidopsis thaliana*.

SEQ ID NO:11 Predicted protein sequence of Os08g24946.1 from *Oryza sativa*.

SEQ ID NO:12 Predicted protein sequence of Sb02g003420.1 from *Sorghum bicolor*.

SEQ ID NO:13 Predicted protein sequence of Bradi4g27190.1 from *Brachypodium distachyon*.

SEQ ID NO:14 Glossy15 nucleotide coding sequence from *Z. mays* W64A (GenBank U41466).

SEQ ID NO:15 Glossy15 predicted protein sequence from *Z. mays* W64A.

SEQ ID NO:16 GRMZM2G096016 nucleotide coding sequence from *Z. mays*.

SEQ ID NO:17 GRMZM2G096016 predicted protein sequence from *Z. mays*.

SEQ ID NO:18 Cg1 nucleotide coding sequence for miR156 transcripts.

DETAILED DESCRIPTION OF THE INVENTION

The invention provides a gene, and methods for its use, to modulate the transition of plant tissue from the juvenile to the adult phase of growth. By modulate is meant to either hasten or delay such transition. A plant or product comprising a recombinant DNA construct comprising such a gene may exhibit improved properties relating to, for instance, biofuel production and/or processing, use as animal feed, and resistance to a plant pest or plant disease, and is also an aspect of the invention. Seed of such a plant is also an aspect of the invention. Thus, for instance, one or more agronomic traits of a grass, such as a member of the Poaceae including corn, sorghum, rice, and switchgrass, among others, may be enhanced. Such traits may include one or more of: improved vegetative yield; reduced recalcitrance during biofuel processing; improved resistance to a plant pest such as European Corn Borer; improved resistance to a plant disease such as a rust disease; enhanced cold tolerance; enhanced digestibility of an animal feed ingredient such as plant vegetative tissue; and improved nutritional content of plant vegetative tissue.

GRMZM2G362718 is a gene of previously unknown function in corn (maize) which was identified through chromosomal mapping of juvenile plant tissue, and apparently functions as a trigger of juvenile to adult growth phase change. Predicted protein alignments (e.g. FIGS. 2-3) show that this gene encodes a protein with some similarity to the enhanced downy mildew 2-transcription factor (EDM2) of *Arabidopsis*, rice, *Brachypodium*, and sorghum (displaying approximately 52.9, 56.7, 42.9, 63.2, percent similarity,

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respectively). Modulating, such as disrupting, the expression of GRMZM2G362718 may alter, such as extend, the temporal duration during which a plant is in a juvenile phase of growth. Homologs of GRMZM2G362718 exist in other plant species such as *Arabidopsis*, rice (*Oryza sativa*), *Brachypodium*, and sorghum (*Sorghum bicolor*), among others; see exemplary sequence database accession numbers AT5G55390.1, Os08g24946.1, Bradi4g27190.1, and Sb02g003420.1, respectively (SEQ ID NOs: 10-13), so this effect may be seen in other plants, e.g. monocotyledonous plants such as grass plants (e.g. members of the Poaceae such as maize, rice sorghum, or switchgrass), as well as dicotyledonous plants.

An additional genome wide association analysis, using transcript presence/absence as the dependent variable, identified GRMZM2G096016 (LOC100285984; Maize Genome Sequencing Project; MaizeSequence.org; Schnable et al. *Science*, 326:1112, 2009) on chromosome 2 as also being associated with a change in the timing of production of the last juvenile leaf, e.g. when vegetative phase change was scored by identifying the last leaf with epicuticular wax. Although close in proximity (~24.5 Kb) to the first candidate gene underlying this QTL (i.e. GRMZM2G362718), GRMZM2G096016, which encodes a predicted nuclear transcription factor Y-subunit A-10, is not in linkage disequilibrium with EDM2. Thus, in particular embodiments, the invention provides methods and compositions for modulating expression of GRMZM2G362718 and/or GRMZM2G096016, each found on maize chromosome 2, or homologs thereof, in order to alter the timing of vegetative phase change in maize, rice, sorghum, switchgrass, or other plants.

MicroRNAs play an important role in regulating the timing of plant developmental transitions. By regulating transcripts of developmental genes, miRNAs control some aspects of leaf morphology, polarity and floral organ identity, and some stress responses (Willmann and Poethig, *Curr. Opin. Plant Biol.* 8:548-552, 2005) as well as the timing of juvenile to adult vegetative phase change. The maize and *Arabidopsis* signaling pathway and miRNA expression cascade are similar (Nonogaki, *Plant Cell Physiol.* 51:1840-1846, 2010). In maize, the Corngrass1 (Cg1) mutant retains juvenile traits resulting in initiation of tillers at each leaf axil causing a bush-like appearance. This phenotype is due to the ectopic overexpression of two tandem miR156 genes (Chuck et al., *Nature Genetics* 39:544-549, 2007; Chuck et al., *PNAS* 108:17550-17555, 2011; GenBank: GQ905502.1). miR156 targets SBP-domain transcription factors—teosinte glume architecture1 (*tga1*) in maize and SPL13 in *Arabidopsis*. SPB transcription factors up regulate miR172 in both species and miR172 targets AP2-like transcription factors such as glossy15 in maize and SCHN-ARCHSAPFEN (SNZ) in *Arabidopsis*. Glossy15 maintains expression of juvenile traits in the leaf epidermis and suppresses adult traits. Mutants of glossy15 (G115) show premature vegetative phase change to the adult state (Evans et al., *Devel.* 120:1971-1981, 1994). In Cg1 mutants of maize, the overexpression of miR156 causes a decrease in *tga1* and miR172 (Chuck, 2007, *ibid*), which cause an increase in expression of Glossy15.

In further embodiments, the invention provides methods and compositions for modulating the expression of one or more additional genes involved in regulating the juvenile to adult growth phase change, in conjunction with modulating expression of GRMZM2G362718 and/or GRMZM2G096016, or homologs thereof. Thus, for instance, the expression of Glossy15 (G115;

GRMZM2G160730), or Cg1, or a homolog thereof, may be modulated along with modulation of expression of GRMZM2G362718 and/or GRMZM2G096016, or a homolog thereof, in a plant.

I. Nucleic Acids, Polypeptides and Plant Transformation Constructs

Certain embodiments of the current invention concern polynucleotide sequences comprising a GRMZM2G362718 coding sequence, or a GRMZM2G096016 coding sequence. Exemplary coding sequences for use with the invention include SEQ ID NO: 2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, and SEQ ID NO:16, encoding the polypeptides of SEQ ID NO: 3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, and SEQ ID NO:17, respectively. Constructs may also be designed that are complementary to all or part of the promoter and other control regions, exons, introns or even exon-intron boundaries of a gene.

Other contemplated constructs may be designed which, in addition to a GRMZM2G362718 coding sequence, GRMZM2G096016 coding sequence, or homolog thereof, also comprise all or part of a Glossy15 or Cg1 and/or other coding sequence, wherein such additional sequence also modulates the juvenile to adult growth phase change. Thus for instance, such constructs, in addition to comprising all or part of a GRMZM2G362718 coding sequence, or homolog thereof, may further comprise, for instance, a Glossy15 coding sequence, or homolog thereof. Exemplary coding sequences for use with the invention therefore include SEQ ID NO:14, encoding the polypeptide of SEQ ID NO:15, and SEQ ID NO:18.

The invention provides a nucleic acid sequence identical over its entire length to each coding sequence provided herein. The invention further provides a nucleic acid sequence displaying at least 85%, 90%, 95%, or 99% identity over its entire length to a the full length, or a fragment, of the coding sequence provided herein. The invention also provides the coding sequence for the polypeptide or a fragment thereof in a reading frame with other coding sequences, such as those encoding a leader or secretory sequence, a pre-, pro-, or prepro-protein sequence. The nucleic acid can also include non-coding sequences, including for example, but not limited to, non-coding 5' and 3' sequences, such as the transcribed, untranslated sequences, termination signals, ribosome binding sites, sequences that stabilize mRNA, introns, polyadenylation signals, and additional coding sequence that encodes additional amino acids. For example, a marker sequence can be included to facilitate the purification of a fused polypeptide. Nucleic acids of the present invention also include nucleic acids comprising a structural gene and the naturally associated sequences that control gene expression.

"Identity," as is well understood in the art, is a relationship between two or more polypeptide sequences or two or more polynucleotide sequences, as determined by comparing the sequences. In the art, "identity" also means the degree of sequence relatedness between polypeptide or polynucleotide sequences, as determined by the match between strings of such sequences. Methods to determine "identity" are designed to give the largest match between the sequences tested. Moreover, methods to determine identity are codified in publicly available programs. "Identity" can be readily calculated by known methods. Computer programs can be used to determine "identity" between two sequences these programs include but are not limited to, GCG; suite of

BLAST programs, three designed for nucleotide sequences queries (BLASTN, BLASTX, and TBLASTX) and two designed for protein sequence queries (BLASTP and TBLASTN). The BLASTX program is publicly available from NCBI and other sources (BLAST Manual, Altschul, S., et al., NCBI NLM NIH, Bethesda, Md. 20894; Altschul, S., et al., *J. Mol. Biol.* 215:403-410, 1990). The well known Smith Waterman algorithm can also be used to determine identity.

Parameters for polypeptide sequence comparison include the following: Algorithm: Needleman and Wunsch (*J. Mol. Biol.* 48:443-453, 1970); Comparison matrix: BLOSUM62 from Hentikoff and Hentikoff, (*PNAS* 89:10915-10919, 1992); Gap Penalty: 12; and Gap Length Penalty: 4. A program which can be used with these parameters is publicly available as the "gap" program from Genetics Computer Group, Madison Wis. The above parameters along with no penalty for end gap may serve as default parameters for peptide comparisons.

Parameters for nucleic acid sequence comparison include the following: Algorithm: Needleman and Wunsch (1970); Comparison matrix: matches=+10; mismatches=0; Gap Penalty: 50; and Gap Length Penalty: 3. A program which can be used with these parameters is publicly available as the "gap" program from Genetics Computer Group, Madison Wis. The above parameters may serve as the default parameters for nucleic acid comparisons.

The present inventors have identified chromosomal regions responsible for such growth, and in particular a specific candidate gene termed GRMZM2G362718 that may trap a plant in a juvenile phase of growth. Marker assisted breeding as well as methods of genetic modification may thus be used to introduce or introgress this gene, a modified version of this gene, or the described linkage group, into a plant to alter the timing of the juvenile to adult growth transition to achieve agronomic improvement. In certain embodiments of the invention, the process for producing such plants or lines comprises introducing a recombinant copy of GRMZM2G362718 or GRMZM2G096016, or a variant thereof into a plant. In other embodiments, the method comprises introgressing at least one chromosomal locus mapping to QTL bounded by markers mmc2184 and mmp183 on maize chromosome 2 into a plant. In other embodiments the function of a gene controlling the juvenile to adult phase change may be disrupted, allowing for enhanced juvenile growth, such as by delaying the juvenile to adult growth phase transition.

Vectors used for plant transformation may include, for example, plasmids, cosmids, YACs (yeast artificial chromosomes), BACs (bacterial artificial chromosomes) or any other suitable cloning system, as well as fragments of DNA there from. Thus when the term "vector" or "expression vector" is used, all of the foregoing types of vectors, as well as nucleic acid sequences isolated there from, are included. It is contemplated that utilization of cloning systems with large insert capacities will allow introduction of large DNA sequences comprising more than one selected gene. In accordance with the invention, this could be used to introduce genes corresponding to an entire biosynthetic pathway into a plant. Introduction of such sequences may be facilitated by use of bacterial or yeast artificial chromosomes (BACs or YACs, respectively), or even plant artificial chromosomes.

II. Antisense and RNAi Constructs

A polynucleotide construct of the present invention may comprise a DNA for expression of an antisense RNA,

siRNA or miRNA, which modulates expression of a GRMZM2G362718 or GRMZM2G096016 coding sequence. By “modulates expression” is meant an increase or a decrease in such expression. Techniques for RNAi are well known in the art. Antisense and RNAi treatments represent one way of altering agronomic characteristics in accordance with the invention (e.g., by down regulation of a GRMZM2G362718 and/or GRMZM2G096016 coding sequence). In particular, constructs comprising a GRMZM2G362718 coding sequence, including fragments thereof (or a GRMZM2G096016 coding sequence or fragments thereof), in antisense orientation, or combinations of sense and antisense orientation, may be used to decrease or effectively eliminate the expression of a GRMZM2G362718 or GRMZM2G096016 coding sequence in a plant and to alter agronomic characteristics (e.g., leaf morphology or disease resistance). Accordingly, each of these may be used to “knock-out” the function of a GRMZM2G362718 or GRMZM2G096016 coding sequence or homologous sequences thereof.

III. Genetic Transformation

Suitable methods for transformation of plant or other cells for use with the current invention are believed to include virtually any method by which DNA can be introduced into a cell, such as by direct delivery of DNA such as by PEG-mediated transformation of protoplasts. These methods and their use are well known in the art.

After effecting delivery of exogenous DNA to recipient cells, the next steps generally concern identifying the transformed cells for further culturing and plant regeneration. In order to improve the ability to identify transformants, one may desire to employ a selectable or screenable marker gene with a transformation vector prepared in accordance with the invention. In this case, one would then generally assay the potentially transformed cell population by exposing the cells to a selective agent or agents, or one would screen the cells for the desired marker gene trait.

Cells that survive the exposure to the selective agent, or cells that have been scored positive in a screening assay, may be cultured in media that supports regeneration of plants. In an exemplary embodiment, MS media may be modified by including further substances such as growth regulators. Examples of such growth regulators are dicamba and 2,4-D. However, other growth regulators may be employed, including NAA, NAA+2,4-D or picloram. Media improvement in these and like ways has been found to facilitate the growth of cells at specific developmental stages. Tissue may be maintained on a basic media with growth regulators until sufficient tissue is available to begin plant regeneration efforts, or following repeated rounds of manual selection, until the morphology of the tissue is suitable for regeneration, then transferred to media conducive to maturation of embryoids. Cultures are transferred as needed on this medium. Shoot development will signal the time to transfer to medium lacking growth regulators.

The transformed cells, identified by selection or screening and cultured in an appropriate medium that supports regeneration, will then be allowed to mature into plants. Developing plantlets are transferred to soilless plant growth mix, and hardened, e.g., in an environmentally controlled chamber, for example, at about 85% relative humidity, 600 ppm CO₂, and 25-250 microeinsteins m⁻² s⁻¹ of light. Plants may be matured in a growth chamber or greenhouse. Plants can be regenerated from about 6 wk to 10 months after a transformant is identified, depending on the initial tissue.

During regeneration, cells are grown on solid media in tissue culture vessels. Illustrative embodiments of such vessels are petri dishes and Plant Cons. Regenerating plants can be grown at a suitable temperature, for instance about 19 to 28° C. After the regenerating plants have reached the stage of shoot and root development, they may be transferred to a greenhouse for further growth and testing.

To confirm the presence of the exogenous DNA or “transgene(s)” in the regenerating plants, a variety of assays may be performed. Such assays include, for example, “molecular biological” assays, such as Southern and northern blotting and PCR™; “biochemical” assays, such as detecting the presence of a protein product, e.g., by immunological means (ELISAs and western blots) or by enzymatic function; plant part assays, such as leaf or root assays; and also, by analyzing the phenotype of the whole regenerated plant.

Very frequently the expression of a gene product is determined by evaluating the phenotypic results of its expression. These assays also may take many forms including but not limited to analyzing changes in the chemical composition, morphology, or physiological properties of the plant. Morphological changes may include ones known to demonstrate juvenile characteristics in plant vegetative tissues, such as presence or absence of wax production, or trichome formation. Most often changes in response of plants or plant parts to imposed treatments are evaluated under carefully controlled conditions termed bioassays.

The present invention provides for a seed of a plant capable of producing a plant having enhanced juvenile growth. In one aspect, the plant can be an open-pollinated variety, a hybrid parent inbred line, or a male sterile line. In another aspect, the invention provides seed of a plant capable of producing a plant having enhanced juvenile growth.

Seeds on transformed plants may occasionally require embryo rescue due to cessation of seed development and premature senescence of plants. To rescue developing embryos, they are excised from surface-disinfected seeds 10-20 days post-pollination and cultured. An embodiment of media used for culture at this stage comprises MS salts, 2% sucrose, and 5.5 g/l agarose. In embryo rescue, large embryos (defined as greater than 3 mm in length) are germinated directly on an appropriate media. Embryos smaller than that may be cultured for 1 wk on media containing the above ingredients along with 10⁻⁵ M abscisic acid and then transferred to growth regulator-free medium for germination.

In yet another aspect, tissue culture of the plants described herein relates to the culture of protoplasts, calli, or plant cells, that are isolated from, or present in, intact parts of the plants described herein.

Once plants are produced which display an enhanced, e.g. extended, juvenile phase of growth, the plants themselves can be cultivated in accordance with conventional procedures, including via tissue culture and by sexual reproduction. The seeds resulting from sexual reproduction can be recovered and planted or otherwise grown as a means of propagation. Plants may also be obtained through asexual reproduction. Protoplast or propagules (e.g., cuttings, scions or rootstocks) can be recovered from plants or parts thereof and may be employed to propagate additional plants.

The present invention also provides for and includes a container of seeds.

One aspect of the invention relates to vegetative tissues, including tissues harvested, dried, or otherwise processed,

biomass produced by a plant having a genome that comprises at least one genetic locus giving rise to an enhanced juvenile phase of growth.

The present invention also provides progeny of plants displaying extended juvenile growth. As used herein, progeny include not only, without limitation, the products of any cross (be it a backcross or otherwise) between two plants, but all progeny whose pedigree traces back to the original cross.

One embodiment of the present invention provides for a plant that contains a genetic marker linked to one or more locus allowing for extended juvenile growth. By "extended juvenile growth locus" or "enhanced juvenile growth locus" is meant a locus that contributes to such extended or enhanced juvenile growth either alone or in combination with one more other locus.

IV. Definitions

As used herein, a "desirable trait" or "desirable traits" include, but are not limited to: increased vegetative growth, improved vegetative yield, improved digestibility when used as animal feed, and improved processing of biomass for preparation of, for instance, biofuel, among others.

As used herein, "polymorphism" means the presence of one or more variations of a nucleic acid sequence at one or more loci in a population of one or more individuals. The variation may comprise but is not limited to one or more base changes, the insertion of one or more nucleotides or the deletion of one or more nucleotides. A polymorphism may arise from random processes in nucleic acid replication, through mutagenesis, as a result of mobile genomic elements, from copy number variation and during the process of meiosis, such as unequal crossing over, genome duplication and chromosome breaks and fusions. The variation can be commonly found, or may exist at low frequency within a population, the former having greater utility in general plant breeding and the latter may be associated with rare but important phenotypic variation. Useful polymorphisms may include single nucleotide polymorphisms (SNPs), insertions or deletions in DNA sequence (Indels), simple sequence repeats of DNA sequence (SSRs) a restriction fragment length polymorphism, and a tag SNP. A genetic marker, a gene, a DNA-derived sequence, a haplotype, a RNA-derived sequence, a promoter, a 5' untranslated region of a gene, a 3' untranslated region of a gene, microRNA, siRNA, a QTL, a satellite marker, a transgene, mRNA, dsRNA, a transcriptional profile, and a methylation pattern may comprise polymorphisms. In addition, the presence, absence, or variation in copy number of the preceding may comprise a polymorphism.

As used herein, "genotype" is the actual nucleic acid sequence at a locus in an individual plant. As used herein, "phenotype" means the detectable characteristics (e.g. number of juvenile leaves, or timing of production of leaves displaying adult morphological characteristics, such as the presence of waxes) of a cell or organism which can be influenced by genotype.

As used herein, linkage of two nucleic acid sequences, including a nucleic acid marker sequence and a nucleic acid sequence of a genetic locus imparting a desired trait may be genetic or physical or both. In one aspect of the invention, the nucleic acid marker and genetic locus conferring an enhanced juvenile growth trait are genetically linked, and exhibit a LOD score of greater than 2.0, as judged by interval mapping for the trait based on maximum likelihood methods described by Lander and Botstein, 1989 (*Genetics*,

121:185-199), and implemented in the software package MAPMAKER (e.g. Lander et al., *Genomics* 1:174-181, (1987); default parameters). Alternatively, other software such as QTL Cartographer v1.17 (Basten et al., Zmap—a QTL cartographer. In: Proceedings of the 5th World Congress on Genetics Applied to Livestock Production Computing Strategies and Software, edited by C. Smith, J. S. Gavora, B. Benkel, J. Chesnais, W. Fairfull, J. P. Gibson, B. W. Kennedy and E. B. Burnside. Volume 22, pages 65-66. Organizing Committee, 5th World Congress on Genetics Applied to Livestock Production, Guelph, Ontario, Canada, 1994; and Basten et al., QTL Cartographer, Version 1.17. Department of Statistics, North Carolina State University, Raleigh, N.C., 2004) may be used. Mapping of QTLs is well-described (e.g. WO 90/04651; U.S. Pat. Nos. 5,492, 547, 5,981,832, 6,455,758; reviewed in Flint-Garcia et al. 2003 (*Ann. Rev. Plant Biol.* 54:357-374, the disclosures of which are hereby incorporated by reference). In other embodiments, the marker and region conferring enhanced juvenile growth are genetically linked and exhibit a LOD score of greater than 3.0, or a LOD score of greater than 6.0, 9.0, 12.0, 15.0, or 18.0. In one embodiment, the marker and region contributing to such growth are genetically linked and exhibit a LOD score of between about 14 and about 20. When assigning the presence of a QTL, the LOD threshold score associated with a QTL analysis as described herein may be determined to be significant for instance at the 95% confidence level, or higher, such as at the 98% or 99% confidence level.

In another aspect, the nucleic acid marker is genetically linked at a distance of between about 0 and about 50 centimorgans (cM) to the locus of interest, e.g. a GRMZM2G362718 or GRMZM2G096016 coding sequence. In other embodiments, the distance between the nucleic acid marker and the locus of interest is between about 0 and about 35 cM, or between about 0 and about 25 cM, or between about 0 and about 15 cM, or between about 0 and about 10 cM, or between about 0 and about 5 cM, including less than about 4, 3, 2 or 1 cM.

As used herein, two nucleic acid molecules are said to be capable of hybridizing to one another if the two molecules are capable of forming an anti-parallel, double-stranded nucleic acid structure. Conventional stringency conditions are described by Sambrook et al., *Molecular Cloning*, A Laboratory Manual, 2nd Ed., Cold Spring Harbor Press, Cold Spring Harbor, N.Y. (1989) and by Haymes et al., *Nucleic Acid Hybridization, A Practical Approach*, IRL Press, Washington, D.C. (1985). Departures from complete complementarity are therefore permissible, as long as such departures do not completely preclude the capacity of the molecules to form a double-stranded structure. Thus, in order for a nucleic acid molecule to serve as a primer or probe it need only be sufficiently complementary in sequence to be able to form a stable double-stranded structure under the particular solvent and salt concentrations employed.

Appropriate stringency conditions which promote DNA hybridization are known in the art, for example 6.0x sodium chloride/sodium citrate (SSC) at about 45° C., followed by a wash of 2.0xSSC at 50° C.; or can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. In some embodiments, hybridization conditions can be high, moderate or low stringency conditions. Preferred conditions include those using 50% formamide, 5.0xSSC, 1% SDS and incubation at 42° C. for 14 hours, followed by a wash using 0.2xSSC, 1% SDS and

incubation at 65° C. Alternative wash conditions, such as of 0.15 M NaCl and 70° C. for 10 minutes may also be used.

The specificity of hybridization can be affected by post-hybridization washes. For example, the salt concentration in the wash step can be selected from a low stringency of about 2.0×SSC at 50° C. to a moderate stringency of about 1.0×SSC at 50° C. to a high stringency of about 0.2×SSC at 50° C.; or 0.15 M NaCl and 70° C. In addition, the temperature in the wash step can be increased from low stringency conditions at room temperature, about 22° C., to moderate stringency conditions at about 50° C., to high stringency conditions at about 65° C. Both temperature and salt concentration may be varied, or either the temperature or the salt concentration may be held constant while the other variable is changed. In some aspects, the wash step can be performed for 5, 10, 15, 20, 25, 30, or more minutes. In another aspect, the wash step is performed for about 20 minutes. In yet another aspect, the wash step can be repeated 1, 2, 3, 4, or more times using the selected salt concentration, temperature, and time. In another aspect, the wash step is repeated twice.

A genetic marker profile of a plant may be predictive of the agronomic traits of a hybrid produced using that inbred. For example, if an inbred plant of known genetic marker profile and phenotype is crossed with a second inbred of known genetic marker profile and phenotype it is possible to predict the phenotype of the F₁ hybrid based on the combined genetic marker profiles of the parent inbreds. Methods for prediction of hybrid performance from genetic marker data are disclosed in U.S. Pat. No. 5,492,547, the disclosure of which is specifically incorporated herein by reference in its entirety. Such predictions may be made using any suitable genetic marker, for example, SSRs, INDELS, RFLPs, AFLPs, SNPs, ISSRs, or isozymes.

Additional markers, such as SSRs, AFLP markers, RFLP markers, RAPD markers, phenotypic markers, SNPs, isozyme markers, or microarray transcription profiles that are genetically linked to or correlated with the juvenile growth trait can be utilized (Walton, *Seed World* 22-29 (July, 1993); Burow and Blake, *Molecular Dissection of Complex Traits*, 13-29, Eds. Paterson, CRC Press, New York (1988)). Methods to isolate such markers and to design probes or primers useful in following the presence of such markers are known in the art. For example, locus-specific SSRs can be obtained by screening a genomic library for SSRs, sequencing of "positive" clones, designing primers which flank the repeats, and amplifying genomic DNA with these primers. Likewise, SNP markers may be identified as well.

The genetic linkage of marker molecules to the loci described herein can be established by a gene mapping model such as, without limitation, the flanking marker model, and the interval mapping, based on maximum likelihood methods described by Lander and Botstein, 1989 (*Genetics*, 121:185-199), and implemented in the software packages MAPMAKER (Whitehead Institute for Biomedical Research, Cambridge Mass., USA) or QTL Cartographer (North Carolina State University, Bioinformatics Research Center) or the like.

A maximum likelihood estimate (MLE) for the presence of a marker is calculated, together with an MLE assuming no trait effect, to avoid false positives. A log₁₀ of an odds ratio (LOD) is then calculated as: LOD=log₁₀ (MLE for the presence of a trait (MLE given no linked trait)).

The LOD score essentially indicates how much more likely the data are to have arisen assuming the presence of a resistance allele rather than in its absence. The LOD threshold value for avoiding a false positive with a given

confidence, say 95%, depends on the number of markers and the length of the genome. Graphs indicating LOD thresholds are set forth in Lander and Botstein (1989), and further described by Ars and Moreno-Gonzalez, *Plant Breeding*, Hayward, Bosemark, Romagosa (eds.) Chapman & Hall, London, pp. 314-331 (1993), and van Ooijen (*Heredity* 83:613-624, 1999).

Selection of appropriate mapping or segregation populations is important in trait mapping. The choice of appropriate mapping population depends on the type of marker systems employed (Tanksley et al., *Molecular mapping plant chromosomes. Chromosome structure and function: Impact of new concepts* J. P. Gustafson and R. Appels (eds.), Plenum Press, New York, pp. 157-173 (1988)). Consideration must be given to the source of parents (adapted vs. exotic) used in the mapping population. Chromosome pairing and recombination rates can be severely disturbed (suppressed) in wide crosses (adapted×exotic) and generally yield greatly reduced linkage distances. Wide crosses will usually provide segregating populations with a relatively large array of polymorphisms when compared to progeny in a narrow cross (adapted×adapted).

Advanced breeding lines are collected from breeding programs. These are tested for their phenotype (e.g. their disease score reactions, the presence of adult leaves, an alteration in the relative proportion of juvenile vs. adult tissues, or an alteration in the timing of production of adult tissues, among others), and genotyped for markers in the QTL intervals described herein. From these data, the smallest genetic interval is identified within each QTL containing the donor parent (DP) favorable allele among the tested lines.

Considerable genetic information can be obtained from a completely classified F₂ population using a codominant marker system (Mather, *Measurement of Linkage in Heredity*: Methuen and Co., (1938)). An F₂ population is the first generation of self or sib pollination after the hybrid seed is produced. Usually a single F₁ plant is self or sib pollinated to generate a population segregating for the nuclear-encoded genes in a Mendelian (1:2:1) fashion.

In contrast to the use of codominant markers, using dominant markers often requires progeny tests (e.g., F₃ or back cross self families) to identify heterozygous individuals. The information gathered can be equivalent to that obtained in a completely classified F₂ population. This procedure is, however, often prohibitive because of the cost and time involved in progeny testing. Progeny testing of F₂ individuals is often used in map construction where error is associated with single plant phenotyping, or when sampling the plants for genotyping affects the ability to perform accurate phenotyping, or where trait expression is controlled by a QTL. Segregation data from progeny test populations (e.g., F₃ or backcrossed or selfed families) can be used in trait mapping. Marker-assisted selection can then be applied to subsequent progeny based on marker-trait map associations (F₂, F₃), where linkage has not been completely disassociated by recombination events (i.e., maximum disequilibrium).

Recombinant inbred lines (RILs) (genetically related lines; usually >F₅) can be used as a mapping population. RILs can be developed by selfing F₂ plants, then selfing the resultant F₃ plants, and repeating this generational selfing process, thereby increasing homozygosity. Information obtained from dominant markers can be maximized by using RILs because all loci are homozygous or nearly so. Under conditions of tight linkage (i.e., about <10% recombination), dominant and co-dominant markers evaluated in RIL popu-

lations provide more information per individual than either marker type in backcross populations (e.g. Reiter et al., 1992; *Proc. Natl. Acad. Sci. (U.S.A.)* 89:1477-1481). However, as the distance between markers becomes larger (i.e., loci become more independent), the information in RIL populations decreases dramatically when compared to codominant markers.

Backcross populations can be utilized as mapping populations. A backcross population (BC) can be created by crossing an F_1 to one of its parents. Typically, backcross populations are created to recover the desirable traits (which may include most of the genes) from one of the recurrent parental (the parent that is employed in the backcrosses) while adding one or a few traits from the second parental, which is often referred to as the donor. A series of backcrosses to the recurrent parent can be made to recover most of the recurrent parent's desirable traits. Thus a population is created consisting of individuals nearly like the recurrent parent, wherein each individual carries varying amounts or a mosaic of genomic regions from the donor parent. Backcross populations can be useful for mapping dominant markers particularly if all loci in the recurrent parent are homozygous and the donor and recurrent parent have contrasting polymorphic marker alleles (Reiter et al., 1992; *Proc. Natl. Acad. Sci. (U.S.A.)* 89:1477-1481).

Information obtained from backcross populations using either codominant or dominant markers is less than that obtained from completely classified F_2 populations because recombination events involving one, rather than two, gametes are sampled per plant. Backcross populations, however, are more informative (at low marker saturation) when compared to RILs as the distance between linked loci increases in RIL populations (i.e., about 15% recombination). Increased recombination can be beneficial for resolution of tight linkages, but may be undesirable in the construction of maps with low marker saturation.

Near-isogenic lines (NIL) created by many backcrosses to produce an array of individuals that are nearly identical in genetic composition except for the trait or genomic region under interrogation can be used as a mapping population. In mapping with NILs, only a portion of the loci polymorphic between the parentals are expected to segregate in the highly homozygous NIL population. Those loci that are polymorphic in a NIL population, however, are likely to be linked to the trait of interest.

Bulk segregant analysis (BSA) is a method developed for the rapid identification of linkage between markers and traits of interest (Michelmore, et al., 1991; *Proc. Natl. Acad. Sci. (U.S.A.)* 88:9828-9832). In BSA, two bulk DNA samples are drawn from a segregating population originating from a single cross. These bulk samples contain individuals that are identical for a particular trait (e.g., resistant or susceptible to a particular pathogen) or genomic region but arbitrary at unlinked regions (i.e., heterozygous). Regions unlinked to the target trait will not differ between the bulked samples of many individuals in BSA.

In another aspect, the present invention provides a method of producing a plant displaying enhanced juvenile growth comprising: (a) crossing a plant displaying such growth with a plant lacking such growth to form a segregating population; (b) screening the population for amount and/or duration of juvenile growth; and (c) selecting one or more members of the population having said enhanced or extended juvenile growth.

For highly heritable traits, a choice of superior individual plants evaluated at a single location will be effective, whereas for traits with low heritability, selection should be

based on statistical analyses (e.g., mean values) obtained from replicated evaluations of families of related plants. Popular selection methods commonly include pedigree selection, modified pedigree selection, mass selection, and recurrent selection. In a preferred embodiment a backcross or recurrent breeding program is undertaken.

The complexity of inheritance influences choice of the breeding method. Backcross breeding can be used to transfer one or a few favorable genes for a highly heritable trait into a desirable cultivar. This approach has been used extensively for breeding disease-resistant cultivars. Various recurrent selection techniques are used to improve quantitatively inherited traits controlled by numerous genes. The use of recurrent selection in self-pollinating crops depends on the ease of pollination, the frequency of successful hybrids from each pollination, and the number of hybrid offspring from each successful cross.

Breeding lines can be tested and compared to appropriate standards in environments representative of the commercial target area(s) for two or more generations. The best lines are candidates as parents for new commercial cultivars; those still deficient in traits may be used as parents for hybrids, or to produce new populations for further selection.

One method of identifying a superior plant is to observe its performance relative to other experimental plants and to a widely grown standard cultivar. If a single observation is inconclusive, replicated observations can provide a better estimate of its genetic worth. A breeder can select and cross two or more parental lines, followed by repeated self or sib pollinating and selection, producing many new genetic combinations.

The development of new plant lines requires the development and selection of varieties, the crossing of these varieties and selection of superior hybrid crosses. The hybrid seed can be produced by manual crosses between selected male-fertile parents or by using male sterility systems. Hybrids can be selected for certain single gene traits such as flower color, seed yield or herbicide resistance that indicate that the seed is truly a hybrid. Additional data on parental lines, as well as the phenotype of the hybrid, influence the breeder's decision whether to continue with the specific hybrid cross.

Pedigree breeding and recurrent selection breeding methods can be used to develop cultivars from breeding populations. Breeding programs combine desirable traits from two or more cultivars or various broad-based sources into breeding pools from which cultivars are developed by selfing and selection of desired phenotypes into parent lines. These lines are used to produce new cultivars. New cultivars can be evaluated to determine which have commercial potential.

Pedigree breeding is used commonly for the improvement of self-pollinating crops. Two parents who possess favorable, complementary traits are crossed to produce an F_1 . An F_2 population is produced by selfing one or several F_1 's. Selection of the best individuals in the best families is performed. Replicated testing of families can begin in the F_4 generation to improve the effectiveness of selection for traits with low heritability. At an advanced stage of inbreeding (i.e., F_6 and F_7), the best lines or mixtures of phenotypically similar lines are tested for potential release as new cultivars.

Backcross breeding and cross breeding have been used to transfer genes for a simply inherited, highly heritable trait into a desirable homozygous cultivar or inbred line, which is the recurrent parent. The source of the trait to be transferred is called the donor parent. The resulting plant obtained from a successful backcrossing program is

expected to have the attributes of the recurrent parent (e.g., cultivar) and the desirable trait transferred from the donor parent. After the initial cross, individuals possessing the phenotype of the donor parent are selected and repeatedly crossed (backcrossed) to the recurrent parent. After multiple backcrossing generations with selection, the resulting line is expected to have the attributes of the recurrent parent (e.g., cultivar) and the desirable trait transferred from the donor parent.

Descriptions of other breeding methods that are commonly used for different traits and crops can be found in one of several available reference books (e.g., Fehr, *Principles of Cultivar Development* Vol. 1, pp. 2-3 (1987)).

The present invention also provides for parts of the plants produced by a method of the present invention. Parts of grass plants, without limitation, include plant cells or parts of plant cells, seed, endosperm, meristem, flower, anther, ovule, pollen, fruit, flowers, stems, roots, stalks or leaves, scions, and root stocks. Plant parts also include the parts of a fruit. In one embodiment of the present invention, the plant part is a seed.

In other aspects of the invention, the plants bearing one or more desirable traits in addition to enhanced juvenile growth may display a greater than 10%, or a greater than 30%, or a greater than 60%, or a greater than 80% reduction in foliar symptoms of, for instance, European corn borer damage on the second leaf above the ear (Riedeman, et al., 2008; *Crop Sci.* 48:1723-1731), relative to an otherwise isogenic control plant. Additionally, juvenile leaves from plants displaying enhanced juvenile growth may comprise increased content of total uronosyl acids, arabinose, and galactose; decreased lignification, decreased neutral sugars, decreased glucose and xylose; decreased ester-linked monomers of p-coumaric acid, and decreased levels of ferulates, among other changes. Such changes may, for instance, beneficially allow for improved efficiency for biofuel production or allow for enhanced feed digestibility or nutritional content.

V. Examples

The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventor to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

Example 1

Genetic Structure of Juvenile to Adult Phase Change in Maize

Juvenile and adult vegetative tissues in grasses differ dramatically in anatomy, biochemical composition, and in their ability to withstand biotic and abiotic stresses. A maize transition leaf, with juvenile tissue distinguished by the presence of epicuticular wax with a dull blueish appearance is shown in FIG. 1. Dark glossy green portions of the leaf are adult tissue.

The molecular network controlling the process of developmental transition has been poorly understood. The present study utilizes the dramatic variation in the timing of juvenile

to adult vegetative transition in different maize populations to identify genes and pathways controlling this fundamental biological process. This work evaluates structured populations and diverse collections of maize that have been characterized extensively for allelic variation, for instance at the GRMZM2G362718 locus, in order to provide a genetic basis for the extensive observed natural variation for developmental timing in plants such as maize. Exemplary phenotypic variation for timing of production of a transition leaf in the maize NAM population is provided in FIG. 4A-4C, with variation for transition leaf by NAM family, labeled by the non-B7 parent in FIG. 4A. The horizontal line at leaf 7 represents the average transition leaf for B73. Phenotypic distribution of the last leaf with epicuticular wax in the NAM population is shown in FIG. 4B. Transition leaf number distribution ranged from leaf 4.5 to leaf 13.25. Phenotypic variation for transition leaf in the IBM, NYH, OWRI, and Wisconsin diversity panel populations is shown in FIG. 4C, plots i-iv.

QTL discovery was accomplished by analysis of a collection of structured biparental mapping populations and a diversity panel of maize inbred lines (summarized in FIG. 5). These included the publicly available Nested Association Mapping (NAM) resource (Flint-Garcia et al *Plant J* 44(6): 1054-64, 2005) and the intermated B73×Mo17 (IBM) RIL mapping population (Lee et al., *Plant Mol Biol* 48(5-6):453-61, 2002). In addition, Oh43×W64A (OWRI) and Ny821×H99 (NyH) populations were evaluated. The diversity panel included a set of northern adapted inbreds described by Hansey et al (*Bioenergy Res.* 3:28-37, 2010) plus 512 lines released by CIMMYT (International Maize and Wheat Improvement Center; Texcoco, Mexico) that are of tropical, subtropical, and highland origin. In total, 5779 unique genotypes were evaluated in at least one location and season, with many of the materials replicated across years.

Example 2

Phenotypic Analysis

The primary trait that was scored to reflect the timing of juvenile to adult transition was the last leaf with juvenile wax (FIG. 1). Maize leaves, in order of emergence, can be fully juvenile, part juvenile and part adult (termed transition leaves), and fully adult. Since the earliest emerging juvenile leaves can senesce and become no longer visible at the time that the uppermost transition leaf can be scored, leaf 5 was marked at the young seedling stage (~V7) by punching a hole in the leaf with a leaf punch. At the ~V10 stage, a paper collar was secured around the stalk between leaf 8 and 9 to mark that internode before the punched leaf 5 fully senesced. The last leaf with juvenile wax was scored on 5 plants per plot with the exact node from which it emerged determined by the position of the leaf collar. At flowering time or thereafter, the total number of leaves (nodes) was determined by counting 5 plants per plot. Node number is both a measure of the duration of plant development (highly correlated with flowering time), but also allowing for calculation of the proportion of nodes which were juvenile versus adult. Days to pollen shed and days to silk emergence were scored by visual assessment of the day that 50% or greater of the plants in a plot had visible pollen shed and visible silk emergence, respectively.

The following linear model was used for phenotypic analysis of the NAMs:

$$Y_{ik} = \mu + G_i + Y_k + e_{ik}$$

where Y is the last leaf with epicuticular wax of the ith genotype (G) in the kth year (Y) and μ is the overall mean with residual error e_{ik} . All effects were considered random.

Repeatability in the NAM, NyH, and OWRI populations were calculated as:

$$R^2 = \frac{\sigma^2(G)}{\sigma^2(E) + \sigma^2(G)}$$

where $\sigma^2(G)$ is the genotypic variance and $\sigma^2(E)$ is the error variance.

The following linear model was used for phenotypic analysis of the IBM, NyH and OWRI populations as well as the Wisconsin Diversity Panel:

$$Y_{ijk} \sim \mu + G_i + R_{j(k)} + Y_k Y_i \times G_i + e_{ijk}$$

where Y is the last leaf with epicuticular wax of the ith genotype (G) in the jth rep (R) within the kth year (Y) and μ is the overall mean. All effects were considered random.

Heritability on an entry mean basis was calculated in the IBM population and the WiDiv panel using the following formula:

$$H^2 = \frac{\sigma^2(G)}{\frac{\sigma^2(E)}{ry} + \frac{\sigma^2(GY)}{r} + \sigma^2(G)}$$

where $\sigma^2(G)$ is the genotypic variance, $\sigma^2(GY)$ is the genotype by year variance and $\sigma^2(E)$ is the error variance.

Significant Pearson and Spearman rank correlations between years were calculated and allowed analysis of averages across years. Following correlation analysis, means across years (and replications for the IBM, NyH, and OWRI populations) were used for QTL mapping. Phenotypic Pearson correlations were performed for transition and flowering time, node number, and internode length.

Example 3

QTL Analysis and Integration of QTL Results Across Materials

1. Nested Association Mapping (NAM) Population:

1106 single nucleotide polymorphisms (SNPs) markers on the 3875 NAM lines (Buckler et al., *Science* 325:714-718, 2009) were used for composite interval mapping with Windows QTL Cartographer v2.5 (Wang, <http://statgen.nesu.edu/qtlcart/WQTCart.htm>, 2011). One thousand permutations were performed to determine an appropriate significance threshold.

QTL were then mapped in a combined analysis of all 25 NAM populations by joint stepwise regression of transition leaf on the same 1106 SNP makers. Because stepwise regression cannot use individuals with missing marker data, an initial step was to impute missing markers. In the joint stepwise regression, a population and marker by population effect was fit. Using the SAS experimental procedure, GLM-SELECT, covariates were determined by forward regression ($p=0.0001$) and SQL was subsequently used to calculate a likelihood ratio for all markers, as per Buckler et al (2009), to determine a genome-wide error rate of 12.26 by permutation.

The 1.6 million SNPs identified in the HapMap project were imputed in the offspring of the NAM RILs based on

founder genotypes. Genome wide association was conducted on top of the joint linkage mapping from above. First, residuals for each chromosome were calculated from the full joint linkage model and with the removal of any QTL located on that chromosome. Single marker analysis was then performed on the residuals across all 1.6 million SNPs to determine significance at each locus. A threshold was also set using 1000 permutation scans.

The last leaf with epicuticular wax varied in the NAM RILs ranging from leaf 4.5 to leaf 13.25 with a repeatability of 0.72. The phenotypic distribution of the NAM families in FIG. 4 shows the trait centering near leaf 7, which is the average transition leaf of B73 and the common parent among the NAMs. Although node number is highly correlated with flowering time, transition leaf was not found to be correlated with flowering time, node number, or internode length with Pearson correlation coefficients of -0.18, -0.10, and 0.07 respectively in the NAM populations.

Through single-population composite interval mapping, 56 total QTL were detected across all NAM populations. A QTL on the long arm of chromosome two in bin ten was detected in 22 of the 25 NAM populations explaining between 5-55% of the variation. The LOD scores ranged from 6.4 to 32.9, while the significance threshold was 2.5.

Similar QTL were detected with the joint-linkage composite interval mapping. The major QTL located on chromosome two had LOD scores of 303.9. The combined average additive effects of the three most significant QTL equate to almost a three-leaf difference in transition, or near 40% of the variation observed in the NAM population. Interestingly, the additive effect of all non-B73 alleles at the chromosome two QTL extends the juvenile wax phase compared to B73.

Using the genome-wide association scan, the most significant SNP is located at 234,407,421 on chromosome two (FIG. 6A) reaching a maximum LOD score of 212.4. FIG. 6B shows results of chromosome two from a similar genome-wide analysis using sub sampling. The results are in agreement with the single marker genome wide scan; the most significant SNP is at position 234,407,421 on chromosome two.

2. Intermated B73xMo17, Ny821xH99, and Oh43xW64A populations:

1340 markers on the recombinant inbred lines of the IBM population (Lee et al *Plant Mol Biol* 48(5-6):453-61, 2002), 78 markers on the NyH RILs, and 169 markers on the OWRI RILs were used for composite interval mapping with Windows QTL Cartographer v2.5 (Wang, 2011). One thousand permutations were performed to determine an appropriate significance threshold. Updated genetic maps of these populations are developed with over 1480 SNP markers identified through genotyping-by-sequencing, and composite interval mapping of transition leaf is analyzed. The increased marker density improves the precision of QTL detection in these populations.

The last leaf with epicuticular wax ranged from leaf 5.4 to 11 in the IBM RILs, from 4.6 to 14.2 in the diversity panel with a heritability of 0.53, 0.6 respectively. The NyH population ranged in transition from leaf 6 to 11 and from leaf 6.9 to 12.2 in the OWRI population (FIG. 4).

The same QTL on chromosome two detected in 23 NAM populations was also detected in the IBM population, having a LOD score of 18.7. This QTL explains 16% of the variation in the IBM population.

Four QTL were detected in the NyH mapping population, one located on chromosome 2. The QTL on chromosome 2 is consistent with the chromosome 2 QTL detected in NAM

and IBM. This QTL explains 11.6% of the variation observed in the NyH population.

3. Wisconsin Diversity Panel (WiDiv):

Over 100,000 SNPs have been identified in this diversity panel through genotyping-by-sequencing (Elshire et al *PLoS One* 6(5): e19379, 2011). Association analysis including appropriate kinship and population structure matrices is performed; and genome-wide association analysis of transition leaf is analyzed on the WiDiv data set.

A summary of all QTL mapping results can be found in Table 1. Numbers indicate LOD score. Overlapping QTL based on the physical position of QTL support intervals are italicized. NAM QTL are presented from joint-linkage composite interval mapping (LOD threshold 12.26). IBM, NyH, and OWRI results are from composite interval mapping (LOD threshold 2.5).

TABLE 1

Summary of QTL detected across all RIL mapping populations.	
Mapping Population	LOD score of QTL on chromosome 2
NAM	21.3, 20.1, 18.2, <i>303.9</i>
IBM	<i>18.7</i>
NyH	3.3

Numbers indicate LOD score. Overlapping QTL based on physical position of QTL support intervals are italicized.

Two common QTL were detected across multiple mapping populations. The QTL on the long arm of chromosome two was detected in NAM, IBM, and NyH populations. No previously known genes affecting vegetative phase change or miRNA targets are located in the chromosome two QTL peak.

The putative chromosome two peak was initially defined as covering a 1.1 Mb region containing over 50 predicted gene models (MaizeGDB; world wide web.maizegdb.org). However, the most significant polymorphism from 1.6 million loci, was narrowed to a single SNP at position 234,407, 421 on chromosome two (AGP_v2). These results demonstrate that a major QTL on chromosome 2 underlies natural variation for this important developmental trait of juvenile-adult transition.

Example 4

Candidate Gene GRMZM2G362718

The gene model nearest the most significant SNP on chromosome two is GRMZM2G362718 whose predicted protein contains a DNMT1 and PHD-finger domain. A protein BLAST shows this gene is highly similar to the enhanced downy mildew 2 (EDM2-encoding) transcription factor of *Arabidopsis*, rice, *Brachypodium*, and sorghum (52.9, 56.7, 42.9, 63.2, percent similarity respectively).

Although the function of GRMZM2G362718 is unknown, several known functions of EDM2 in other species point to its potential significance in underlying the chromosome two QTL. Mutations in EDM2 show a delay in flowering and elevated transcripts of the flowering suppressor FLC (Tsuchiya and Eulgem *Plant. J.* 62:518-528, 2010). These authors reported EDM2's function in regulating the vegetative to floral transition in an FLC-dependent manner; EDM2 also has a direct effect on the juvenile to adult vegetative phase change in *Arabidopsis*.

edm2 plants appear to skip the early juvenile phase of development by not producing the initial pair of rosette

leaves. The effect of edm2-2 on trichome production was also examined by these authors. In wild-type *Arabidopsis*, juvenile leaves lack trichomes on the abaxial side, while adult leaves gradually produce an increasing number of trichomes. Mutant edm2-2 plants delay the onset of trichome production and, therefore, EDM2 seems to have a role in promoting the transition from the juvenile to adult vegetative phase (Tsuchiya and Eulgem *BMC Plant Bio.* 10:203-217, 2010). Further, Willmann and Poethig (*Devel.* 138:677-685, 2011) show FLC has both flowering-dependent and flowering-independent effects on vegetative transition. EDM2 does not appear to affect expression of the trans-acting siRNAs (HASTY, ZIPPY, SGS3, RDR6) or the other five genes (ARF3, ARF4, SPL3, At1g63130, At5g18040) of this pathway that have previously been shown to control vegetative phase change in *Arabidopsis* (Peragine et al *Genes Devel.* 18:2368-2379, 2004). This suggests EDM2's role in vegetative phase change may be independent of the siRNA pathway, and GRMZM2G362718 may act similarly.

Sekhon et al (*Plant J.* 66:553-563, 2011) developed a maize B73 gene atlas showing gene expression levels across all 11 major organs at varying developmental time points (60 total tissue samples). The atlas shows some level of GRMZM2G362718 expression in all tissue sampled, such as a pooled leaf sample as well as in tissue at the base of stage two leaves and immature leaves (v9). Neighboring gene models 500 kb up and downstream of GRMZM2G362718 were therefore studied in the gene atlas to determine if any could be ruled out as candidates due to inappropriate tissue expression. All predicted neighboring gene models were either not present in the atlas data set or were expressed at some level in the shoot apical meristem.

RNA-seq expression levels on a subset of the Wisconsin diversity panel (Hansey et al *PLoS ONE* 7(3):e33071, 2011) were thus used to determine if a relationship exists between expression of GRMZM2G362718 and timing of vegetative phase change. In this analysis, diverse inbreds were ordered from early to late transition and their gene expression pattern is plotted. Either categorical differences (i.e. as shown by groups of early or late transitioning inbreds have a shared expression level), or quantitative differences (i.e. via a progressive increase or decrease in expression level trending with timing of phase change) would indicate a relationship between the expression of GRMZM2G362718 and phenotype. Analysis of RNA-seq information is performed to demonstrate such differences.

Specific allelic contrasts between B73, Mo17, Oh43, and W64A show some association whereby later transitioning plants displayed higher expression levels of GRMZM2G362718 (FIG. 7). For example, B73 has an average transition leaf of 7.5 and an expression level of 5 fragments per kilobase per million reads (FPKM) compared to Oh43 which transitions at leaf 9.5 on average, and has an expression level of 13 FPKM. In this comparison, the later transition corresponds with a higher expression level. However the comparison is between plants with different GRMZM2G362718 alleles which may differ in function or activity, and thus correlating function and expression level may not be straightforward. It is also important to consider the tissue sampled (whole seedling) when making these comparisons; thus further expression analysis of the shoot apical meristem at various developmental time points is performed.

Candidate Gene Glossy 15

QTL mapping performed with the NAM population detected three major QTL located on chromosomes two, three, and nine, which had LOD scores of 303.9, 87.5 and 141.2 respectively (FIG. 8). The gene model nearest the most significant SNP on chromosome nine is Glossy 15 (“G115” (GRMZM2G160730); e.g. Moose and Sisco, *Genes Dev.* 10:3018-3027, 1996). Glossy15 encodes an AP2-like transcription factor which is responsible for the expression of adult traits in the leaf epidermis. Additional mapping populations were also analyzed. Based on overlapping LOD confidence intervals, the QTL detected on chromosomes 2 and 9 after composite interval mapping of the IBM population are consistent with the QTL detected in the NAM population. For the Wisconsin Diversity Panel population a mixed linear model including relatedness and population structure was used to perform a genome wide association study. After an experiment wide Bonferroni correction for multiple tests, one genomic region was significantly associated with changes in the production of the last juvenile leaf (“LJL”) and was located on chromosome nine with an additive effect of -0.43 , relative to the minor allele. The most significant SNPs in this region are located within the gene Glossy15.

Glossy15 is thus a candidate gene, modulation of expression or activity of which can result in altering the timing of juvenile to adult phase change in plants. For instance, Glossy15 may be utilized in conjunction with GRMZM2G362718, and/or GRMZM2G096016 (see Example 6), to modulate, e.g. delay, the transition of a plant from a juvenile to an adult phase of growth.

Candidate Gene GRMZM2G096016

Sequencing of whole seedling RNA was conducted from a set of 503 diverse maize inbred lines to evaluate the maize seedling pan-transcriptome as a proxy to the maize pan genome. Using de novo assembly of reads unmapped to the B73 reference genome, 8,681 novel representative transcript assemblies (RTAs) were identified. Genomic Presence/Absence Variation Analysis was performed, and pooled reads were cleaned using the fastx_clipper program within the FASTX toolkit. The minimum sequence length was set to 15

bp after clipping using both Illumina single end adapter sequences. Sequence reads were parsed into individual genotype files requiring a perfect match to the barcode and ApeKI cut site, and the barcode sequences were removed.

Sequence reads were mapped to AGPv2 using Bowtie version 0.12.7 (Langmead, *Genome Biol.* 10:R25, 2009) requiring a unique alignment and allowing up to two mismatches. SAMtools version 0.1.7 (Li et al., *Bioinformatics* 25:2078-2079, 2009) was used to generate unfiltered pileup files. Representative genes/RTAs with at least two uniquely aligned reads were considered present at the genome level.

Sequence reads for each library were mapped to an AGPv2 formatted maize reference genome plus the 8,681 unfiltered RTAs using Bowtie version 0.12.7 (Langmead, 2009, *ibid*) and TopHat version 1.4.1 (Trapnell et al., *Nature Protocols* 7:562-578, 2012). Normalized gene expression levels were determined using Cufflinks version 1.3.0 (Trapnell, *ibid*). To characterize transcript presence/absence variation (PAV), sequence reads were also mapped to AGPv2 plus the 8,681 unfiltered RTAs requiring a unique alignment. A gene/RTA was then defined as expressed if the fragments per kilobase of exon model per million fragments mapped (FPKM) low confidence interval as described by Cufflinks was greater than zero. The 503 included inbred lines were clustered with hierarchical clustering using a Pearson correlation distance metric and average linkage using Multiple Experiment Viewer Software (MeV) version 4.5 (Saeed et al., *Biotechniques* 34:374-378, 2003).

Vegetative phase change was scored by identifying the last leaf with epicuticular wax in a subset of the 503 inbred lines. Significant natural variation for the last juvenile leaf was observed, ranging from leaf 3.45 to leaf 13.4. 186,733 SNPs were subjected to genome wide association analysis (GWAS) which was performed using a mixed linear model accounting for both familial relatedness (Q) and population structure (K) (Yu et al., *Nature Genetics* 38:203-208, 2006). GWAS was also performed with transcript presence/absence state for all of the reference genes and RTAs for last juvenile leaf. The association analysis was done using the same mixed model as described above but instead of using a SNP as the dependent variable, transcript presence/absence was used as the genetic marker. In the presence/absence analysis, GRMZM2G096016 (GenBank EU975023.1) which encodes predicted nuclear transcription factor Y-subunit A-10, was found to be significantly associated with regulation of the timing of vegetative phase change transition, and may be utilized to modulate, e.g. delay, the transition of a plant from a juvenile to an adult phase of growth.

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<211> LENGTH: 1006

<212> TYPE: PRT

<213> ORGANISM: Zea mays

<400> SEQUENCE: 3

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 Asn Arg Tyr Tyr Phe Glu Asp Gly Glu Glu Lys Pro Val Cys Phe Ser
 20 25 30
 Ile Leu Pro Phe Gln Phe Gly Glu Asp Asp Ser Glu Ala Val Phe Leu
 35 40 45
 Arg Lys Asp Val Phe Leu Cys Gly Phe Val Asp Lys Asn Leu Pro Val
 50 55 60
 Tyr Lys Glu Val Val Ala Trp Lys Ile Arg Leu Asp Ser Glu His Pro
 65 70 75 80
 Asn Ile Tyr Val Leu Ser Ile Glu His Lys Trp Ile Lys Leu Leu Lys
 85 90 95
 Pro Arg Lys Cys Tyr Gly Asp Ile Val Arg Ser Thr Leu Ile Thr Val
 100 105 110
 Gln Met Leu His Phe Phe Gly Arg Gly Glu Gln Arg Ser Ser Asn His
 115 120 125
 Leu Trp Asp His Leu Asp Glu Val Phe Gly Lys Tyr Asn Pro Lys Pro
 130 135 140
 Val Glu Asp Asp Leu Met Lys His His Thr Leu Ile Lys Leu Phe Val
 145 150 155 160
 Glu Lys Asp Gln Thr Leu Met Lys Ser Lys Ile Leu Gln Arg Leu Ile
 165 170 175
 Glu Asn Gly Phe Lys Arg Thr Lys Lys Ala Leu Gly Met Glu Ala Gln
 180 185 190
 Ser Ile Val Ser Asp Gly Trp Arg Ala Arg Lys Asn Asp Asp Asn Asn
 195 200 205
 Tyr Gly Asn Lys Asp Asp Ser Gly Asp Asp Cys Asp Gly Asp Gly Ser
 210 215 220
 Ser Asp Asp Gly Asp Gly Ser Ser Asp Asp Asp Val Thr Asp Gln Ile
 225 230 235 240
 Cys Ala Leu Cys Asp Asp Gly Gly His Leu Leu Ser Cys Asp Gly Pro
 245 250 255
 Cys Lys Arg Ser Phe His Pro Thr Lys Lys Asp Gly Arg Glu Ser Lys
 260 265 270
 Cys Glu Ser Leu His Tyr Thr Ser Ala Glu Val Lys Arg Ile Gly Thr
 275 280 285
 Tyr Leu Cys Ala Asn Cys Lys Asn Lys Gln His Gln Cys Phe Arg Cys
 290 295 300
 Gly Glu Leu Glu Pro Ser His Gly Pro Asn Ala Lys Val Phe Gln Cys
 305 310 315 320
 Asn Gln Ala Ser Cys Gly Tyr Phe Tyr His Pro Lys Cys Ile Ala Gln
 325 330 335
 Leu Leu Asp Pro Asn Ala Thr Asp Gly Glu Leu Glu Arg Arg Ile Met
 340 345 350
 Ser Gly Met Ser Phe Pro Cys Pro Ile His Trp Cys Phe Lys Cys Gly
 355 360 365
 His Met Glu Asn Lys Ala Gln Arg Ala Leu Gln Leu Ala Val Cys Arg
 370 375 380
 Arg Cys Pro Arg Ala Tyr His Arg Glu Cys Leu Pro Arg Asp Leu Ser

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385	390	395	400
Phe Gly Thr Lys Asp Lys Asp Gly Asn Gln Arg Ala Trp Lys Leu Ser	405	410	415
Asp Thr Ile Phe Ile Tyr Cys Leu Asp His Glu Ile Asp Lys Asp Thr	420	425	430
Gly Thr Thr Ser Arg Asn His Ile Lys Phe Pro Ala Thr Pro Glu Tyr	435	440	445
Thr Lys Thr Lys Gly Leu Gly Asn Ser Lys Gly Arg Met Thr Gly Lys	450	455	460
Arg Arg Lys Asn Lys Arg Arg Lys Asn Thr Asp Gln Ser Thr Lys Pro	465	470	475
Thr Asp Leu Pro Asn Arg Leu Cys Gly Ala Glu Ser Glu Gln Ala Asp	485	490	495
Asn Val Gly Ala Lys Ser Thr Leu Pro Gln Ile Val Val Glu Pro His	500	505	510
Cys Ala Ala Lys His Leu Lys Gly Asp Pro Gln Ile Ala Lys Gln Gly	515	520	525
Val Ala Ala Arg Gln Asn Gly Ala Glu Thr Met Lys Gly His Glu Asn	530	535	540
Gln Phe Gly Ile Ser Phe Cys Val Ala Ser Thr Glu Thr Glu Lys Arg	545	550	555
Val Thr Cys Leu Ala Gln Arg Gly Thr Cys Leu Gly Thr Gln Tyr Asp	565	570	575
Gly Pro Ser Thr Lys Gly Met Tyr Asp Cys Ser Val Gln Asp Thr Pro	580	585	590
Met Asp Asp Asp Val Glu Leu Asp Asn Val Ala Cys Ile Ile Ala Val	595	600	605
Asp Lys Tyr Val Asn Gly Arg Gly Lys Thr Gln Glu Asp Tyr Thr Arg	610	615	620
Lys Glu Ala Ala Gln Arg Lys Asp Ser Ser Glu Asn Gln Gly Gln Asn	625	630	635
Asp Ala Leu Glu Leu Asp Asn Leu Arg Met Glu Met Gln Ala Asp Glu	645	650	655
Arg Pro Leu Glu Pro Gly Asn Lys Arg Asp Arg Lys Trp Gln Lys Asn	660	665	670
Val Tyr Gly Leu Gly Ser Ala Ser Gly Gln Lys Glu Thr Leu Ser Arg	675	680	685
Arg Glu Asn Pro Arg Ser Asp Arg Gly Met Val His Ser Asn Asp Ser	690	695	700
Lys Thr Ile Tyr Tyr Arg Lys Gly Gly Thr Glu Val Asp Asn Val Asp	705	710	715
Asp His Pro Leu Glu Lys Gln Asp His Gln Asp Thr Ser Ser Asp Gly	725	730	735
Ser Lys Lys Arg Ser Arg Pro Val Asp Asn Ala Ser Gly Gly Asn Arg	740	745	750
Pro Tyr Leu Asp Glu Asn Lys Lys Arg Asn Leu Arg Glu Asp Gly Arg	755	760	765
Tyr Ala His Tyr Glu Asp Trp Arg Ser Glu Arg Asn Thr Ala Ala Asp	770	775	780
Thr Ser Gly Tyr Lys Ala Gln Ser Glu Glu Lys Pro Val Trp Thr Asn	785	790	795
Thr Arg Thr Gly Ser Arg Glu His Ser Leu Asp Arg Gln Arg Ile Glu	805	810	815

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Cys Gly Asp Ser Tyr Arg Gly Thr Tyr Asn Asn Arg Gln Arg His Glu
 820 825 830
 Trp Leu His Pro His Ala Ser Gly Asn Ser Ser Arg Ile Gly Trp Asp
 835 840 845
 Asp Arg Arg Gln Trp Ser Ser Ser Arg Ser Pro Phe Pro Ser Ala Glu
 850 855 860
 Phe Gly Gly Asp Arg Ser Cys Ser Arg Ala His Pro Arg Gly Ser Lys
 865 870 875 880
 Tyr Arg Thr Gly Gly Arg His Asp His Pro Gln Tyr Leu Gly Leu Gly
 885 890 895
 Thr Pro Gln His Gly Thr Ser Arg Pro His His Thr Met Gly Trp Asp
 900 905 910
 Arg Asp Thr Phe His Asp His Gln His Gly Arg Arg Pro Pro His His
 915 920 925
 Thr Met Gly Trp Asp Arg Ala Pro Phe Arg Asp His Gln His Gly Glu
 930 935 940
 Tyr Asp Asp Ser Arg Tyr Gly Glu Tyr Asp Ala Thr Asp Asn Gly Pro
 945 950 955 960
 Asp Ser Ala His Arg Pro Tyr Thr Ala Ala Gly Val Ala Gly Arg Ser
 965 970 975
 Ala Pro Ser Tyr Gln Leu Ala Gly Gly Tyr Gly Glu Gly Ser Arg Ala
 980 985 990
 Trp Arg Pro Val Thr Asp Lys Tyr Ala Pro Trp Pro Leu Pro
 995 1000 1005

<210> SEQ ID NO 4
 <211> LENGTH: 3021
 <212> TYPE: DNA
 <213> ORGANISM: Zea mays

<400> SEQUENCE: 4

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 gacgatagcg aggcagtttt cctcagaaag gatgttttct tgtgtggatt tgtggataaa 180
 aatcttcctg tgtacaagga ggtggtagct tggagataa ggcttgacag tgagcatccc 240
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 tatggagaca ttgttcgatc aacattgatt acggtgcaaa tgctccactt tttggggaga 360
 ggggagcaaa gaagtttgaa tcacctttgg gatcaccttg atgaagtttt tggtaaatac 420
 aatcctaaac ccgtggagga tgacttgatg aagcaccata ccctaataca gttgtttgta 480
 gagaaagatc aaaccttgat gaagtcaaag attcttcaaa ggctcattga gaatggcttt 540
 aagagaacta aaaaggcctt aggtatggaa gcacaatcca ttgttagtga cgggtggcgt 600
 gctagaaaaa atgatgataa caattatggt aacaaagatg acagtggatg tgattgtgat 660
 ggtgatggta gcagtgatga tggatgatgc agcagtgatg atgatgttac cgatcaataa 720
 tgtgcgctat gtgatgatgg aggacatttg cttagctgtg acggtccatg caagaggctc 780
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<210> SEQ ID NO 5
<211> LENGTH: 1005
<212> TYPE: PRT
<213> ORGANISM: Zea mays

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<400> SEQUENCE: 5

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Ile	Leu	Pro	Phe	Gln	Phe	Gly	Glu	Asp	Asp	Ser	Glu	Ala	Val	Phe	Leu
		35					40					45			
Arg	Lys	Asp	Val	Phe	Leu	Cys	Gly	Phe	Val	Asp	Lys	Asn	Leu	Pro	Val
	50					55					60				
Tyr	Lys	Glu	Val	Val	Ala	Trp	Lys	Ile	Arg	Leu	Asp	Ser	Glu	His	Pro
65					70					75					80
Asn	Ile	Tyr	Val	Leu	Ser	Ile	Glu	His	Lys	Trp	Ile	Lys	Leu	Leu	Lys
			85						90					95	
Pro	Arg	Lys	Cys	Tyr	Gly	Asp	Ile	Val	Arg	Ser	Thr	Leu	Ile	Thr	Val
			100					105						110	
Gln	Met	Leu	His	Phe	Phe	Gly	Arg	Gly	Glu	Gln	Arg	Ser	Leu	Asn	His
		115					120					125			
Leu	Trp	Asp	His	Leu	Asp	Glu	Val	Phe	Gly	Lys	Tyr	Asn	Pro	Lys	Pro
	130					135						140			
Val	Glu	Asp	Asp	Leu	Met	Lys	His	His	Thr	Leu	Ile	Lys	Leu	Phe	Val
145					150					155					160
Glu	Lys	Asp	Gln	Thr	Leu	Met	Lys	Ser	Lys	Ile	Leu	Gln	Arg	Leu	Ile
				165					170						175
Glu	Asn	Gly	Phe	Lys	Arg	Thr	Lys	Lys	Ala	Leu	Gly	Met	Glu	Ala	Gln
			180					185						190	
Ser	Ile	Val	Ser	Asp	Gly	Trp	Arg	Ala	Arg	Lys	Asn	Asp	Asp	Asn	Asn
		195					200					205			
Tyr	Gly	Asn	Lys	Asp	Asp	Ser	Gly	Asp	Asp	Cys	Asp	Gly	Asp	Gly	Ser
	210					215					220				
Ser	Asp	Asp	Gly	Asp	Gly	Ser	Ser	Asp	Asp	Asp	Val	Thr	Asp	Gln	Ile
225					230					235					240
Cys	Ala	Leu	Cys	Asp	Asp	Gly	Gly	His	Leu	Leu	Ser	Cys	Asp	Gly	Pro
				245					250					255	
Cys	Lys	Arg	Ser	Phe	His	Pro	Thr	Lys	Lys	Asp	Gly	Arg	Glu	Ser	Lys
			260					265					270		
Cys	Glu	Ser	Leu	His	Tyr	Thr	Ser	Ala	Glu	Val	Lys	Arg	Ile	Gly	Thr
		275					280					285			
Tyr	Leu	Cys	Ala	Asn	Cys	Lys	Asn	Lys	Gln	His	Gln	Cys	Phe	Arg	Cys
	290					295					300				
Gly	Glu	Leu	Glu	Pro	Ser	His	Gly	Pro	Asn	Ala	Lys	Val	Phe	Gln	Cys
305					310					315					320
Asn	Gln	Ala	Ser	Cys	Gly	Tyr	Phe	Tyr	His	Pro	Lys	Cys	Ile	Ala	Gln
				325					330					335	
Leu	Leu	Asp	Pro	Asn	Ala	Thr	Asp	Gly	Glu	Leu	Glu	Arg	Arg	Ile	Met
			340					345					350		
Ser	Gly	Met	Ser	Phe	Pro	Cys	Pro	Ile	His	Trp	Cys	Phe	Lys	Cys	Gly
		355					360					365			
His	Met	Glu	Asn	Lys	Ala	Gln	Arg	Ala	Leu	Gln	Leu	Ala	Val	Cys	Arg
	370					375					380				
Arg	Cys	Pro	Arg	Ala	Tyr	His	Arg	Glu	Cys	Leu	Pro	Arg	Asp	Leu	Ser
385					390					395					400
Phe	Gly	Thr	Lys	Asp	Lys	Asp	Gly	Asn	Gln	Arg	Ala	Trp	Lys	Leu	Ser
				405					410					415	
Asp	Thr	Ile	Phe	Ile	Tyr	Cys	Leu	Asp	His	Glu	Ile	Asp	Lys	Asp	Thr
		420						425					430		
Gly	Thr	Thr	Ser	Arg	Asn	His	Ile	Lys	Phe	Pro	Ala	Thr	Pro	Glu	Tyr

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Gly Gly Asp Arg Ser Cys Ser Arg Ala His Pro Arg Gly Ser Lys Tyr
 865 870 875 880
 Arg Thr Gly Gly Arg His Asp His Pro Gln Tyr Leu Gly Leu Gly Thr
 885 890 895
 Pro Gln His Gly Thr Ser Arg Pro His His Thr Met Gly Trp Asp Arg
 900 905 910
 Asp Thr Phe His Asp His Gln His Gly Arg Arg Pro Pro His His Thr
 915 920 925
 Met Gly Trp Asp Arg Ala Pro Phe Arg Asp His Gln His Gly Glu Tyr
 930 935 940
 Asp Asp Ser Arg Tyr Gly Glu Tyr Asp Ala Thr Asp Asn Gly Pro Asp
 945 950 955 960
 Ser Ala His Arg Pro Tyr Thr Ala Ala Gly Val Ala Gly Arg Ser Ala
 965 970 975
 Pro Ser Tyr Gln Leu Ala Gly Gly Tyr Gly Glu Gly Ser Arg Ala Trp
 980 985 990
 Arg Pro Val Thr Asp Lys Tyr Ala Pro Trp Pro Leu Pro
 995 1000 1005

<210> SEQ ID NO 6
 <211> LENGTH: 3021
 <212> TYPE: DNA
 <213> ORGANISM: Zea mays

<400> SEQUENCE: 6

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 aatcttcctg tgtacaagga ggtggtagct tggaaagataa ggcttgacag tgagcatccc 240
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 aatcctaaac cctgtggagga tgacttgatg aagcaccata ccctaataca gttgtttgta 480
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 gctagaaaaa atgatgataa caattatggt aacaaagatg acagtgggta tgatttgatg 660
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 aatgccactg atggtgagtt ggaagaaggg attatgtcgg gaatgtcatt tccgtgcccc 1080
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gaccaccctt tagaaaagca agaccaccag gatacatcaa gtgacggatc taaaagaga 2220
agccgatctg tggacaacgc atctgggtgc aacagaccat acttggatga gagcaagaag 2280
cgtaatctca gagaagatgg aagatgtct cattatgaag actggagaag tgaaggaat 2340
acagcagcag acacgtctgg atataaggcc caatcagaag agaagcctgt atggacaaac 2400
actcgaacag gatcaagggc gcattcactg gacaggcaaa ggatagatg tggtgacagc 2460
tatcgtggaa cctataacaa tagacaaga catgaatgga cgcacccgca cgctagtggg 2520
aattcctcga gaattggttg ggtgacagg aggcagtgga gttcatctcg gtcaccattt 2580
ccttcggctg aatttgggtg tgaccgttcc tgttctctg cccatccgag aggttctaaa 2640
tacagaaccg gcgggaggca tgatcaccac cagtacctgg gactgggaac acctcaacat 2700
ggtacaagta gaccgcacca cacaatgggc tgggacaggg acacctccca tgatcaccag 2760
catggcagaa gaccgccgca ccacacaatg ggctgggaca gggccccctt ccgtgatcac 2820
cagcatggcg aataccacga ctccaggtat ggtgaatatg atgcaactga caatggtcct 2880
gacagcgcgc atcgacccta caeggtgctt ggctggctg gacgttcagc accgagttat 2940
cagcttctg gtggttatgg agagggatca aggcttggc ggccagttac ggacaagtac 3000
gccccatggc ccttgccttg a 3021

```

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<210> SEQ ID NO 7
<211> LENGTH: 1005
<212> TYPE: PRT
<213> ORGANISM: Zea mays

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<400> SEQUENCE: 7

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Met Phe Asp Asp Asp Asp Gly Val Asp Pro Gln Ile Glu Asp Val
 1             5             10             15
Asn Arg Tyr Tyr Phe Glu Asp Gly Glu Glu Lys Pro Val Cys Phe Ser
 20             25             30
Ile Leu Pro Phe Gln Phe Gly Glu Asp Asp Ser Glu Ala Val Phe Leu
 35             40             45
Arg Lys Asp Val Phe Leu Cys Gly Phe Val Asp Lys Asn Leu Pro Val
 50             55             60

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Tyr Lys Glu Val Val Ala Trp Lys Ile Arg Leu Asp Ser Glu His Pro
 65 70 75 80
 Asn Ile Tyr Val Leu Ser Ile Glu His Lys Trp Ile Lys Leu Leu Lys
 85 90 95
 Pro Arg Lys Cys Tyr Gly Asp Ile Val Arg Ser Thr Leu Ile Thr Val
 100 105 110
 Gln Met Leu His Phe Phe Gly Arg Gly Glu Gln Arg Ser Leu Asn His
 115 120 125
 Leu Trp Asp His Leu Asp Glu Val Phe Gly Lys Ser Asn Pro Lys Pro
 130 135 140
 Val Glu Asp Asp Leu Met Lys His His Thr Leu Ile Lys Leu Phe Val
 145 150 155 160
 Glu Lys Asp Gln Thr Leu Met Lys Ser Lys Ile Leu Gln Arg Leu Ile
 165 170 175
 Glu Asn Gly Phe Lys Arg Thr Lys Lys Ala Leu Gly Met Glu Ala Gln
 180 185 190
 Ser Ile Val Ser Asp Gly Trp Arg Ala Arg Lys Asn Asp Asp Asn Asn
 195 200 205
 Tyr Gly Asn Lys Asp Asp Ser Gly Asp Asp Cys Asp Gly Asp Gly Ser
 210 215 220
 Ser Asp Asp Gly Asp Gly Ser Ser Asp Asp Asp Val Thr Asp Gln Ile
 225 230 235 240
 Cys Ala Leu Cys Asp Asp Gly Gly His Leu Leu Ser Cys Asp Gly Pro
 245 250 255
 Cys Lys Arg Ser Phe His Pro Thr Lys Lys Asp Gly Arg Glu Ser Lys
 260 265 270
 Cys Glu Ser Leu His Tyr Thr Ser Ala Glu Val Lys Arg Ile Gly Thr
 275 280 285
 Tyr Leu Cys Ala Asn Cys Lys Asn Lys Gln His Gln Cys Phe Arg Cys
 290 295 300
 Gly Glu Leu Glu Pro Ser His Gly Pro Asn Ala Lys Val Phe Gln Cys
 305 310 315 320
 Asn Gln Ala Ser Cys Gly Tyr Phe Tyr His Pro Lys Cys Ile Ala Gln
 325 330 335
 Leu Leu Asp Pro Asn Ala Thr Asp Gly Glu Leu Glu Arg Arg Ile Met
 340 345 350
 Ser Gly Met Ser Phe Pro Cys Pro Ile His Trp Cys Phe Lys Cys Gly
 355 360 365
 His Met Glu Asn Lys Ala Gln Arg Ala Leu Gln Leu Ala Val Cys Arg
 370 375 380
 Arg Cys Pro Arg Ala Tyr His Arg Glu Cys Leu Pro Arg Asp Leu Ser
 385 390 395 400
 Phe Gly Thr Lys Asp Lys Asp Gly Asn Gln Arg Ala Trp Lys Leu Ser
 405 410 415
 Asp Thr Ile Phe Ile Tyr Cys Leu Asp His Glu Ile Asp Lys Asp Thr
 420 425 430
 Gly Thr Thr Ser Arg Asn His Ile Lys Phe Pro Ala Thr Pro Glu Tyr
 435 440 445
 Thr Lys Thr Lys Gly Leu Gly Asn Ser Lys Arg Met Thr Gly Lys Arg
 450 455 460
 Arg Lys Asn Lys Arg Arg Lys Asn Thr Asp Gln Ser Thr Lys Pro Thr
 465 470 475 480
 Asp Leu Pro Asn Arg Leu Cys Gly Ala Glu Ser Glu Gln Ala Asp Asn

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485				490				495							
Val	Gly	Ala	Lys	Ser	Thr	Leu	Pro	Gln	Ile	Val	Val	Glu	Pro	His	Cys
			500					505				510			
Ala	Ala	Lys	His	Leu	Lys	Gly	Asp	Pro	Gln	Ile	Ala	Lys	Gln	Gly	Val
		515					520					525			
Ala	Ala	Arg	Gln	Asn	Gly	Ala	Glu	Thr	Met	Lys	Gly	His	Glu	Asn	Gln
		530				535					540				
Phe	Gly	Ile	Ser	Phe	Cys	Val	Ala	Ser	Thr	Glu	Thr	Glu	Lys	Arg	Val
545				550						555					560
Thr	Cys	Leu	Ala	Gln	Arg	Gly	Thr	Cys	Leu	Gly	Thr	Gln	Tyr	Asp	Gly
				565						570				575	
Pro	Ser	Thr	Lys	Gly	Met	Tyr	Asp	Cys	Ser	Val	Gln	Asp	Thr	Pro	Met
			580					585				590			
Asp	Asp	Asp	Val	Glu	Leu	Asp	Asn	Val	Ala	Cys	Ile	Ile	Ala	Val	Asp
		595					600					605			
Lys	Tyr	Val	Asn	Glu	Arg	Glu	Lys	Thr	Gln	Glu	Asp	Tyr	Thr	Arg	Lys
	610					615					620				
Glu	Ala	Ala	Gln	Arg	Lys	Asp	Ser	Ser	Glu	Asn	Gln	Gly	Gln	Asn	Asp
625					630					635					640
Ala	Leu	Glu	Leu	Asp	Asn	Leu	Arg	Met	Glu	Met	Gln	Ala	Asp	Lys	Arg
				645					650					655	
Pro	Leu	Glu	Pro	Gly	Asn	Lys	Arg	Asp	Arg	Lys	Trp	Gln	Lys	Asn	Ala
			660					665				670			
Tyr	Gly	Leu	Gly	Ser	Ala	Ser	Gly	Gln	Lys	Glu	Thr	Leu	Ser	Arg	Arg
		675					680					685			
Glu	Asn	Pro	Pro	Ser	Asp	Arg	Gly	Met	Val	His	Ser	Asn	Asp	Ser	Lys
	690					695					700				
Thr	Ile	Tyr	Tyr	Arg	Lys	Gly	Gly	Thr	Glu	Val	Asp	Asn	Val	Asp	Asp
705					710					715					720
His	Pro	Leu	Glu	Lys	Gln	Asp	His	Gln	Asp	Thr	Ser	Ser	Asp	Gly	Ser
				725					730					735	
Lys	Lys	Arg	Ser	Arg	Ser	Val	Asp	Asn	Ala	Ser	Gly	Gly	Asn	Arg	Pro
			740					745				750			
Tyr	Leu	Asp	Glu	Ser	Lys	Lys	Arg	Asn	Leu	Arg	Glu	Asp	Gly	Arg	Tyr
		755					760					765			
Ala	His	Tyr	Glu	Asp	Trp	Arg	Ser	Glu	Arg	Asn	Thr	Ala	Ala	Asp	Thr
		770				775					780				
Ser	Gly	Tyr	Lys	Ala	Gln	Ser	Glu	Glu	Lys	Pro	Val	Trp	Thr	Asn	Thr
				785		790				795					800
Arg	Thr	Gly	Ser	Arg	Glu	His	Ser	Leu	Asp	Arg	Gln	Arg	Ile	Glu	Cys
				805					810					815	
Gly	Asp	Ser	Tyr	Arg	Gly	Thr	Tyr	Asn	Asn	Arg	Gln	Arg	His	Glu	Trp
			820					825				830			
Pro	His	Pro	His	Ala	Ser	Gly	Asn	Ser	Ser	Arg	Ile	Gly	Trp	Asp	Asp
		835					840					845			
Arg	Arg	Gln	Trp	Ser	Ser	Ser	Arg	Ser	Pro	Phe	Pro	Ser	Ala	Glu	Phe
		850				855					860				
Gly	Gly	Asp	Arg	Ser	Cys	Ser	Arg	Ala	His	Pro	Arg	Gly	Ser	Lys	Tyr
				865		870				875				880	
Arg	Thr	Gly	Gly	Arg	His	Asp	His	Pro	Gln	Tyr	Leu	Gly	Leu	Gly	Thr
				885					890					895	
Pro	Gln	His	Gly	Thr	Ser	Arg	Pro	His	His	Thr	Met	Gly	Trp	Asp	Arg
			900					905						910	

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Asp Thr Ser His Asp His Gln His Gly Arg Arg Pro Pro His His Thr
 915 920 925
 Met Gly Trp Asp Arg Ala Pro Phe Arg Asp His Gln His Gly Glu Tyr
 930 935 940
 His Asp Ser Arg Tyr Gly Glu Tyr Asp Ala Thr Asp Asn Gly Pro Asp
 945 950 955 960
 Ser Ala His Arg Pro Tyr Thr Ala Ala Gly Val Ala Gly Arg Ser Ala
 965 970 975
 Pro Ser Tyr Gln Leu Ala Gly Gly Tyr Gly Glu Gly Ser Arg Ala Trp
 980 985 990
 Arg Pro Val Thr Asp Lys Tyr Ala Pro Trp Pro Leu Pro
 995 1000 1005

<210> SEQ ID NO 8
 <211> LENGTH: 3021
 <212> TYPE: DNA
 <213> ORGANISM: Zea mays

<400> SEQUENCE: 8

atgtttgatg atgatgatga tggagtggac ccacaaattg aggatgtcaa cagatactac 60
 tttgaggatg gtgaagagaa accagtttgt ttcagtatct tgcctttcca gtttgggtgag 120
 gacgatagcg aggcagtttt cctcagaaag gatgttttct tgtgtggatt tgtggataaa 180
 aatcttcctg tgtacaagga ggtggtagct tggaaagataa ggcttgacag tgagcatccc 240
 aacatctatg tgctttctat tgagcacaag tggataaagc tggttgaaacc acgaaaatgc 300
 tatggagaca ttgttcgatc aacattgatt acgggtgcaaa tgctccactt ttttgggaga 360
 ggggagcaaa gaagtttgaa tcacctttgg gatcaccttg atgaagtttt tggtaaatcc 420
 aatcctaaac ccggtggagga tgacttgatg aagcaccata ccctaataca gttgtttgta 480
 gagaaagatc aaaccttgat gaagtcaaag attcttcaaa ggctcattga gaatggcttt 540
 aagagaacta aaaaggcctt aggtatggaa gcacaatcca ttgttagtga cgggtggcgt 600
 gctagaaaaa atgatgataa caattatggt aacaaagatg acagtggatg tgatttgatg 660
 ggtgatggta gcagtgatga tggatgatggc agcagtgatg atgatgttac cgatcaaata 720
 tgtgcgctat gtgatgatgg aggacatttg cttagctgtg acggccatg caagaggctc 780
 ttccacccca caaagaaaga tggcagagaa tctaaatgtg aaagtcttca ttacacttca 840
 gcagaagtaa agagaattgg tacttatcta tgtgcaaaact gcaaaaataa gcaacaccaa 900
 tgttttagat gtggagagct tgaacatcc catgggcca atgctaaggc ctttcaatgc 960
 aatcaagcat cttgtggata tttttaccac cctaagtgca ttgcacaatt attggatcct 1020
 aatgccactg atggtgagtt ggaagaagg attatgtcgg gaatgtcatt tccgtgcccc 1080
 atacattggt gtttcaaag tggccacatg gagaacaaag ctcaaagagc acttcagctt 1140
 gcagtgtgta gacgtgttcc aagagcatat cacagggaat gccttccaag ggacttatcc 1200
 tttgaaaca aggacaagga tggtaaacca cgcgcttggga agctttccga cacaattttc 1260
 atttactgcc tagatcatga aatagacaag gatactggca caactagtag gaacctata 1320
 aaatttcag ctacacctga atacaccaa acaaaagggc ttggtaacag caaagtaagg 1380
 atgactggca aaaggagaaa gaacaaaagg agaaagaaca ctgaccaatc aacaaaacct 1440
 acagatttgc caaacaggtt gtgtggagca gaaagtgagc aagctgacaa tgtaggtgca 1500
 aaaagcacat tgccccagat tgtgttagag cctcactgtg cagcaaaagca ctggaagggt 1560

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gatccacaaa ttgccaaaaca ggggtgttgc gctcgtcaaa atgggtgcaga aactatgaaa 1620
gggcatgaaa atcaatttgg catttcattt tgtgttgcaa gtactgaaac agagaagagg 1680
gtaacatggt tggcacaaga ggggacatgt ttagggacac aatatgatgg gccatcaacc 1740
aagggcattg atgattgttc tgttcaggac accccaatgg acgacgatgt tgagtggat 1800
aatgtggcct gcataatcgc ggtggataaa tatgtcaatg gaaggggaaa aacacaagag 1860
gactacacta gaaaagaagc tgctcagcgc aaagactcga gtgaaaatca agggcagaat 1920
gatgctctag agctagacaa cctccggatg gagatgcaag ctgacaaaacg tccgttagaa 1980
ccaggaaaca agagggacag gaagtggcag aaaaatgcat atggactcgg atcagcttcg 2040
ggacagaagg aaacctgtgc caggagagaa aatccaccgt cagatagagg gatgggccac 2100
agtaacgaca gaaaacaat ttattacagg aagggtggga cggaagtcga taatggtgat 2160
gaccaccctt tagaaaagca agaccaccag gatacatcaa gtgacggatc taaaagaga 2220
agccgaccag tggacaacgc atctgggtggc aacagaccat acttggatga gaacaagaag 2280
cgtaatttct gagaagatgg aagatatgct cattatgaag actggagaag tgaaggaat 2340
acagcagcag acacgtctgg atataaggcc caatcagaag agaagcctgt atggacaaac 2400
actcgaacag gatcaaggga gcattcactg gacaggcaaa ggatagatg cggtgacagc 2460
tatcgtggaa cctataacaa tagacaaga catgaatggt tgcacccgca cgctagtggg 2520
aattcctcga gaattggttg ggtgacagg aggcagtgga gttcatctcg gtcaccattt 2580
ccttcggctg aatttgggtg tgaccgttcc tgttctctg cccatccgag aggttctaaa 2640
tacagaaccg gggggaggca tgatcaccoc cagtacctgg gactgggaac acctcaacat 2700
ggtacaagta gaccgcacca cacaatgggc tgggacaggg acaccttcca tgatcaccag 2760
catggcagaa gaccgccgca ccacacaatg ggctgggaca gggccccctt ccgtgatcac 2820
cagcatggcg aatacgacga ctccaggtat ggtgaatatg atgcaactga caatggtcct 2880
gacagcgcgc atcgacccta caeggctgct ggcgtggctg gacgttcagc accgagttat 2940
cagcttgctg gtggttatgg agagggatca agggcttggc ggccagttac ggacaagtac 3000
gccccatggc ccttgccttg a 3021

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<210> SEQ ID NO 9
<211> LENGTH: 1005
<212> TYPE: PRT
<213> ORGANISM: Zea mays

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<400> SEQUENCE: 9

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Met Phe Asp Asp Asp Asp Gly Val Asp Pro Gln Ile Glu Asp Val
 1             5             10             15
Asn Arg Tyr Tyr Phe Glu Asp Gly Glu Glu Lys Pro Val Cys Phe Ser
                20             25             30
Ile Leu Pro Phe Gln Phe Gly Glu Asp Asp Ser Glu Ala Val Phe Leu
            35             40             45
Arg Lys Asp Val Phe Leu Cys Gly Phe Val Asp Lys Asn Leu Pro Val
 50             55             60
Tyr Lys Glu Val Val Ala Trp Lys Ile Arg Leu Asp Ser Glu His Pro
65             70             75             80
Asn Ile Tyr Val Leu Ser Ile Glu His Lys Trp Ile Lys Leu Leu Lys
            85             90             95
Pro Arg Lys Cys Tyr Gly Asp Ile Val Arg Ser Thr Leu Ile Thr Val
100             105             110

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Gln Met Leu His Phe Phe Gly Arg Gly Glu Gln Arg Ser Leu Asn His
 115 120 125
 Leu Trp Asp His Leu Asp Glu Val Phe Gly Lys Ser Asn Pro Lys Pro
 130 135 140
 Val Glu Asp Asp Leu Met Lys His His Thr Leu Ile Lys Leu Phe Val
 145 150 155 160
 Glu Lys Asp Gln Thr Leu Met Lys Ser Lys Ile Leu Gln Arg Leu Ile
 165 170 175
 Glu Asn Gly Phe Lys Arg Thr Lys Lys Ala Leu Gly Met Glu Ala Gln
 180 185 190
 Ser Ile Val Ser Asp Gly Trp Arg Ala Arg Lys Asn Asp Asp Asn Asn
 195 200 205
 Tyr Gly Asn Lys Asp Asp Ser Gly Asp Asp Cys Asp Gly Asp Gly Ser
 210 215 220
 Ser Asp Asp Gly Asp Gly Ser Ser Asp Asp Asp Val Thr Asp Gln Ile
 225 230 235 240
 Cys Ala Leu Cys Asp Asp Gly Gly His Leu Leu Ser Cys Asp Gly Pro
 245 250 255
 Cys Lys Arg Ser Phe His Pro Thr Lys Lys Asp Gly Arg Glu Ser Lys
 260 265 270
 Cys Glu Ser Leu His Tyr Thr Ser Ala Glu Val Lys Arg Ile Gly Thr
 275 280 285
 Tyr Leu Cys Ala Asn Cys Lys Asn Lys Gln His Gln Cys Phe Arg Cys
 290 295 300
 Gly Glu Leu Glu Pro Ser His Gly Pro Asn Ala Lys Val Phe Gln Cys
 305 310 315 320
 Asn Gln Ala Ser Cys Gly Tyr Phe Tyr His Pro Lys Cys Ile Ala Gln
 325 330 335
 Leu Leu Asp Pro Asn Ala Thr Asp Gly Glu Leu Glu Arg Arg Ile Met
 340 345 350
 Ser Gly Met Ser Phe Pro Cys Pro Ile His Trp Cys Phe Lys Cys Gly
 355 360 365
 His Met Glu Asn Lys Ala Gln Arg Ala Leu Gln Leu Ala Val Cys Arg
 370 375 380
 Arg Cys Pro Arg Ala Tyr His Arg Glu Cys Leu Pro Arg Asp Leu Ser
 385 390 395 400
 Phe Gly Thr Lys Asp Lys Asp Gly Asn Gln Arg Ala Trp Lys Leu Ser
 405 410 415
 Asp Thr Ile Phe Ile Tyr Cys Leu Asp His Glu Ile Asp Lys Asp Thr
 420 425 430
 Gly Thr Thr Ser Arg Asn His Ile Lys Phe Pro Ala Thr Pro Glu Tyr
 435 440 445
 Thr Lys Thr Lys Gly Leu Gly Asn Ser Lys Val Arg Met Thr Gly Lys
 450 455 460
 Arg Arg Lys Asn Lys Arg Arg Lys Asn Thr Asp Gln Ser Thr Lys Pro
 465 470 475 480
 Thr Asp Leu Pro Asn Arg Leu Cys Gly Ala Glu Ser Glu Gln Ala Asp
 485 490 495
 Asn Val Gly Ala Lys Ser Thr Leu Pro Gln Ile Val Val Glu Pro His
 500 505 510
 Cys Ala Ala Lys His Ser Lys Gly Asp Pro Gln Ile Ala Lys Gln Gly
 515 520 525
 Val Ala Ala Arg Gln Asn Gly Ala Glu Thr Met Lys Gly His Glu Asn

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530			535			540									
Gln	Phe	Gly	Ile	Ser	Phe	Cys	Val	Ala	Ser	Thr	Glu	Thr	Glu	Lys	Arg
545					550						555				560
Val	Thr	Cys	Leu	Ala	Gln	Arg	Gly	Thr	Cys	Leu	Gly	Thr	Gln	Tyr	Asp
			565						570					575	
Gly	Pro	Ser	Thr	Lys	Gly	Met	Tyr	Asp	Cys	Ser	Val	Gln	Asp	Thr	Pro
			580					585					590		
Met	Asp	Asp	Asp	Val	Glu	Leu	Asp	Asn	Val	Ala	Cys	Ile	Ile	Ala	Val
		595					600					605			
Asp	Lys	Tyr	Val	Asn	Gly	Arg	Gly	Lys	Thr	Gln	Glu	Asp	Tyr	Thr	Arg
	610					615					620				
Lys	Glu	Ala	Ala	Gln	Arg	Lys	Asp	Ser	Ser	Glu	Asn	Gln	Gly	Gln	Asn
625					630						635				640
Asp	Ala	Leu	Glu	Leu	Asp	Asn	Leu	Arg	Met	Glu	Met	Gln	Ala	Asp	Lys
			645						650					655	
Arg	Pro	Leu	Glu	Pro	Gly	Asn	Lys	Arg	Asp	Arg	Lys	Trp	Gln	Lys	Asn
			660						665				670		
Ala	Tyr	Gly	Leu	Gly	Ser	Ala	Ser	Gly	Gln	Lys	Glu	Thr	Leu	Ser	Arg
		675						680					685		
Arg	Glu	Asn	Pro	Pro	Ser	Asp	Arg	Gly	Met	Val	His	Ser	Asn	Asp	Ser
	690					695					700				
Lys	Thr	Ile	Tyr	Tyr	Arg	Lys	Gly	Gly	Thr	Glu	Val	Asp	Asn	Val	Asp
705					710						715				720
Asp	His	Pro	Leu	Glu	Lys	Gln	Asp	His	Gln	Asp	Thr	Ser	Ser	Asp	Gly
			725						730					735	
Ser	Lys	Lys	Arg	Ser	Arg	Pro	Val	Asp	Asn	Ala	Ser	Gly	Gly	Asn	Arg
			740					745					750		
Pro	Tyr	Leu	Asp	Glu	Asn	Lys	Lys	Arg	Asn	Phe	Glu	Asp	Gly	Arg	Tyr
		755					760					765			
Ala	His	Tyr	Glu	Asp	Trp	Arg	Ser	Glu	Arg	Asn	Thr	Ala	Ala	Asp	Thr
		770				775					780				
Ser	Gly	Tyr	Lys	Ala	Gln	Ser	Glu	Glu	Lys	Pro	Val	Trp	Thr	Asn	Thr
785					790					795					800
Arg	Thr	Gly	Ser	Arg	Glu	His	Ser	Leu	Asp	Arg	Gln	Arg	Ile	Glu	Cys
			805						810					815	
Gly	Asp	Ser	Tyr	Arg	Gly	Thr	Tyr	Asn	Asn	Arg	Gln	Arg	His	Glu	Trp
			820					825					830		
Leu	His	Pro	His	Ala	Ser	Gly	Asn	Ser	Ser	Arg	Ile	Gly	Trp	Asp	Asp
		835					840						845		
Arg	Arg	Gln	Trp	Ser	Ser	Ser	Arg	Ser	Pro	Phe	Pro	Ser	Ala	Glu	Phe
		850				855					860				
Gly	Gly	Asp	Arg	Ser	Cys	Ser	Arg	Ala	His	Pro	Arg	Gly	Ser	Lys	Tyr
865					870					875					880
Arg	Thr	Gly	Gly	Arg	His	Asp	His	Pro	Gln	Tyr	Leu	Gly	Leu	Gly	Thr
			885						890					895	
Pro	Gln	His	Gly	Thr	Ser	Arg	Pro	His	His	Thr	Met	Gly	Trp	Asp	Arg
			900					905					910		
Asp	Thr	Phe	His	Asp	His	Gln	His	Gly	Arg	Arg	Pro	Pro	His	His	Thr
		915						920					925		
Met	Gly	Trp	Asp	Arg	Ala	Pro	Phe	Arg	Asp	His	Gln	His	Gly	Glu	Tyr
		930				935					940				
Asp	Asp	Ser	Arg	Tyr	Gly	Glu	Tyr	Asp	Ala	Thr	Asp	Asn	Gly	Pro	Asp
945					950					955					960

-continued

Ser Ala His Arg Pro Tyr Thr Ala Ala Gly Val Ala Gly Arg Ser Ala
 965 970 975

Pro Ser Tyr Gln Leu Ala Gly Gly Tyr Gly Glu Gly Ser Arg Ala Trp
 980 985 990

Arg Pro Val Thr Asp Lys Tyr Ala Pro Trp Pro Leu Pro
 995 1000 1005

<210> SEQ ID NO 10
 <211> LENGTH: 1297
 <212> TYPE: PRT
 <213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 10

Met Thr Phe Val Asp Asp Asp Glu Glu Glu Asp Phe Ser Val Pro Gln
 1 5 10 15

Ser Ala Ser Asn Tyr Tyr Phe Glu Asp Asp Asp Lys Glu Pro Val Ser
 20 25 30

Phe Ala Arg Leu Pro Ile Gln Trp Ser Val Glu Glu Lys Val Asp Gly
 35 40 45

Ser Gly Leu Gly Phe Tyr Leu Arg Gly Arg Ser Asp Asn Gly Leu Leu
 50 55 60

Pro Leu His Lys Leu Val Lys Ala Trp Arg Tyr Asp Leu Ser Asn Phe
 65 70 75 80

Gln Pro Glu Ile Ser Val Leu Thr Lys Asp Asn Ile Trp Ile Lys Leu
 85 90 95

Glu Glu Pro Arg Lys Ser Tyr Gly Glu Leu Ile Arg Thr Val Leu Val
 100 105 110

Thr Leu His Ser Ile Gln Phe Leu Arg Arg Asn Pro Gln Ala Ser Glu
 115 120 125

Lys Ala Leu Trp Glu Lys Leu Thr Arg Ser Leu Arg Ser Tyr Asp Val
 130 135 140

Lys Pro Ser Gln Asn Asp Leu Val Asp His Ile Gly Leu Ile Ala Glu
 145 150 155 160

Ala Ala Lys Arg Asp Arg Asn Leu Ala Asn Ser Lys Phe Ile Leu Ala
 165 170 175

Phe Leu Thr Lys Lys Pro Thr Lys Arg Arg Leu Pro Asp Glu Asp Asn
 180 185 190

Ala Lys Asp Asp Phe Ile Val Gly Asp Glu Asp Thr Tyr Val Ala Ser
 195 200 205

Asp Glu Asp Glu Leu Asp Asp Glu Asp Asp Asp Phe Phe Glu Ser Val
 210 215 220

Cys Ala Ile Cys Asp Asn Gly Gly Glu Ile Leu Cys Cys Glu Gly Ser
 225 230 235 240

Cys Leu Arg Ser Phe His Ala Thr Lys Lys Asp Gly Glu Asp Ser Leu
 245 250 255

Cys Asp Ser Leu Gly Phe Asn Lys Met Gln Val Glu Ala Ile Gln Lys
 260 265 270

Tyr Phe Cys Pro Asn Cys Glu His Lys Ile His Gln Cys Phe Ile Cys
 275 280 285

Lys Asn Leu Gly Ser Ser Asp Asn Ser Ser Gly Ala Ala Glu Val Phe
 290 295 300

Gln Cys Val Ser Ala Thr Cys Gly Tyr Phe Tyr His Pro His Cys Val
 305 310 315 320

Thr Arg Arg Leu Arg Leu Gly Asn Lys Glu Glu Ser Glu Ala Leu Glu

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325				330				335							
Arg	Gln	Ile	Ile	Ala	Gly	Glu	Tyr	Thr	Cys	Pro	Leu	His	Lys	Cys	Ser
			340					345					350		
Val	Cys	Glu	Asn	Gly	Glu	Val	Lys	Thr	Asp	Ser	Asn	Leu	Gln	Phe	Ala
		355					360					365			
Val	Cys	Arg	Arg	Cys	Pro	Lys	Ser	Tyr	His	Arg	Lys	Cys	Leu	Pro	Arg
		370				375					380				
Glu	Ile	Ser	Phe	Glu	Asp	Ile	Glu	Asp	Glu	Asp	Ile	Leu	Thr	Arg	Ala
385					390					395					400
Trp	Asp	Gly	Leu	Leu	His	Asn	Arg	Val	Leu	Ile	Tyr	Cys	Gln	Glu	His
			405					410						415	
Glu	Ile	Asp	Glu	Glu	Leu	Leu	Thr	Pro	Val	Arg	Asp	His	Val	Lys	Phe
		420						425					430		
Pro	Phe	Thr	Glu	Glu	Gln	Lys	Val	Phe	Val	Lys	Glu	Gln	Arg	Arg	Ile
		435					440					445			
Leu	Glu	Ser	His	Val	Gly	Arg	Asp	Lys	Ala	Arg	Leu	Lys	Val	Lys	Asp
450						455					460				
Pro	Ala	Leu	Gln	Asp	Thr	Cys	Gly	Lys	Ala	Ser	Lys	Asn	Ser	Phe	Arg
465					470					475					480
Ser	Ser	Phe	Pro	Ser	Ser	Lys	Asp	Gly	Phe	Ser	Thr	Lys	Lys	His	Gly
			485					490						495	
Leu	Val	Ser	Ser	Val	Pro	Asp	His	Ser	Arg	Lys	Arg	Lys	Asp	Ile	Asp
		500						505					510		
Pro	Ser	Ile	Lys	His	Lys	Met	Val	Pro	Gln	Lys	Ser	Gln	Lys	Met	Met
		515					520					525			
Glu	Asp	Ser	Arg	Glu	Ala	Gly	Lys	Asn	Lys	Leu	Gly	Val	Lys	Glu	Ala
530						535					540				
Arg	Asp	Ala	Gly	Lys	Ser	Lys	Ile	Ser	Leu	Gly	Glu	Arg	Leu	Phe	Ser
545					550					555					560
Tyr	Thr	Gln	Glu	Pro	Asn	Pro	Val	Lys	Pro	Gly	Arg	Val	Ile	Pro	Val
			565						570					575	
Asp	Ser	Lys	His	Asn	Lys	Thr	Asp	Ser	Ile	Ala	Ser	Lys	Glu	Pro	Gly
			580					585					590		
Ser	Glu	Ile	Pro	Thr	Leu	Asp	Asn	Asp	Ser	Gln	Arg	Arg	Leu	Leu	Ala
		595					600					605			
Val	Met	Lys	Lys	Ala	Thr	Glu	Glu	Ile	Thr	Met	Gly	Thr	Ile	Leu	Lys
	610					615					620				
Lys	Phe	Lys	Ile	Gln	Ser	Thr	Met	Ser	Thr	His	Ser	Thr	Arg	Asn	Val
625					630					635					640
Val	Asp	Lys	Thr	Ile	Thr	Met	Gly	Lys	Val	Glu	Gly	Ser	Val	Gln	Ala
			645						650					655	
Ile	Arg	Thr	Ala	Leu	Lys	Lys	Leu	Glu	Glu	Gly	Gly	Asn	Ile	Glu	Asp
		660						665					670		
Ala	Lys	Ala	Val	Cys	Glu	Pro	Glu	Val	Leu	Ser	Gln	Ile	Leu	Lys	Trp
		675					680						685		
Lys	Asp	Lys	Leu	Lys	Val	Tyr	Leu	Ala	Pro	Phe	Leu	His	Gly	Ala	Arg
	690					695						700			
Tyr	Thr	Ser	Phe	Gly	Arg	His	Phe	Thr	Asn	Pro	Glu	Lys	Leu	Gln	Gln
705					710					715					720
Ile	Val	Asp	Arg	Leu	His	Trp	Tyr	Ala	Asp	Asp	Gly	Asp	Met	Ile	Val
			725						730					735	
Asp	Phe	Cys	Cys	Gly	Ser	Asn	Asp	Phe	Ser	Cys	Leu	Met	Asn	Ala	Lys
			740					745						750	

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Leu Glu Glu Thr Gly Lys Lys Cys Leu Tyr Lys Asn Tyr Asp Leu Phe
 755 760 765

Pro Ala Lys Asn Asn Phe Asn Phe Glu Arg Lys Asp Trp Met Thr Val
 770 775 780

Ser Lys Asp Glu Leu Glu Pro Gly Ser Lys Leu Ile Met Gly Leu Asn
 785 790 795 800

Pro Pro Phe Gly Val Asn Ala Ser Leu Ala Asn Lys Phe Ile Thr Lys
 805 810 815

Ala Leu Glu Phe Arg Pro Lys Ile Leu Ile Leu Ile Val Pro Pro Glu
 820 825 830

Thr Glu Arg Leu Asp Lys Lys Lys Ser Ser Tyr Val Leu Ile Trp Glu
 835 840 845

Asp Lys Thr Phe Leu Ser Gly Asn Ser Phe Tyr Leu Pro Gly Ser Val
 850 855 860

Asn Glu Glu Asp Lys Gln Leu Glu Asp Trp Asn Leu Val Pro Pro Pro
 865 870 875 880

Leu Ser Leu Trp Ser Arg Ser Asp Phe Ala Ala Lys His Lys Lys Ile
 885 890 895

Ala Glu Lys His Cys His Leu Ser Arg Asp Val Gly Ser Ser Lys Leu
 900 905 910

Lys Ile Val Glu Glu Glu Ala Asn Ala Ser Leu His Pro Leu Gly Ala
 915 920 925

Ser Asp Gly Met Cys Asp Asp Ile Pro Met Glu Lys Asp Glu Leu Glu
 930 935 940

Val Ala Glu Cys Val Asn Lys Ile Leu Val Ser Glu Lys Ile Asp Thr
 945 950 955 960

Val Glu Thr Val Ala Arg Val His Gln Ser Asp His Leu Ser Arg Arg
 965 970 975

Ser Gln Leu Lys Lys Glu Gly Lys Thr Lys Asp Tyr Ser Gly Arg Lys
 980 985 990

Leu Gly Lys Ser Met Asp Ser Asn Asn Val Asp Trp Lys Ser Asn Asp
 995 1000 1005

Met Glu Glu Asp Gln Gly Glu Leu Ser Arg Ala Pro Glu Ser Ile
 1010 1015 1020

Lys Val Lys Ile Pro Glu Met Thr Ser Asp Trp Gln Ser Pro Val
 1025 1030 1035

Arg Ser Ser Pro Asp Asp Ile Tyr Ala Val Cys Thr Ser Ile Ser
 1040 1045 1050

Thr Thr Thr Pro Gln Arg Ser His Glu Ala Val Glu Ala Ser Leu
 1055 1060 1065

Pro Ala Ile Thr Arg Thr Lys Ser Asn Leu Gly Lys Asn Ile Arg
 1070 1075 1080

Glu His Gly Cys Lys Val Gln Gly Thr Gly Lys Pro Glu Val Ser
 1085 1090 1095

Arg Asp Arg Pro Ser Ser Val Arg Thr Ser Arg Glu Asp Ile Tyr
 1100 1105 1110

Thr Val Arg Pro Ser Pro Glu Asn Thr Gly Gln Lys Pro Phe Glu
 1115 1120 1125

Ala Phe Glu Pro Ser Tyr Gly Ala Ser Leu Ser His Phe Asp Asp
 1130 1135 1140

Gly Leu Ala Ala Lys Tyr Gly Gly Phe Gly Gly Gly Tyr Arg Met
 1145 1150 1155

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Pro Asp Pro Pro Phe Leu Pro Asp Gln Phe Pro Leu Arg Asn Gly
 1160 1165 1170
 Pro Asn Glu Met Phe Asp Phe Arg Gly Tyr Ser Asp Leu Asp Arg
 1175 1180 1185
 Gly Ile Gly Gln Arg Glu Tyr Pro Gln Gln Tyr Gly Gly His Leu
 1190 1195 1200
 Asp Pro Met Leu Ala Pro Pro Pro Pro Asn Leu Met Asp Asn
 1205 1210 1215
 Ala Phe Pro Leu Gln Gln Arg Tyr Ala Pro His Phe Asp Gln Met
 1220 1225 1230
 Asn Tyr Gln Arg Met Ser Ser Phe Pro Pro Gln Pro Pro Leu Gln
 1235 1240 1245
 Pro Ser Gly His Asn Leu Leu Asn Pro His Asp Phe Pro Leu Pro
 1250 1255 1260
 Pro Pro Pro Pro Ser Asp Phe Glu Met Ser Pro Arg Gly Phe Ala
 1265 1270 1275
 Pro Gly Pro Asn Pro Asn Tyr Pro Tyr Met Ser Arg Ser Gly Gly
 1280 1285 1290
 Trp Ile Asn Asp
 1295

<210> SEQ ID NO 11
 <211> LENGTH: 1323
 <212> TYPE: PRT
 <213> ORGANISM: *Oryza sativa*

<400> SEQUENCE: 11

Met Met Ser Ser Asp Asp Asp Leu Glu Pro Gln Leu Lys Ala Val Glu
 1 5 10 15
 Asn Tyr Tyr Phe Val Asp Asp Asn Asp Val Pro Val Ser Phe Asp Val
 20 25 30
 Leu Pro Phe Gln Phe Asp Ala Ala Glu Gly Val Ala Ser Phe Lys Lys
 35 40 45
 Asp Val Tyr Leu Arg Gly Phe Thr Asp Gly Gly Leu Gln Lys Val Tyr
 50 55 60
 Lys Gln Val Val Ala Trp Lys Leu Val Leu Asp Gly Asp Ser Pro Glu
 65 70 75 80
 Ile Ala Val Leu Ser Thr Glu Gly Ser Trp Ile Ala Leu Leu Lys Pro
 85 90 95
 Arg Pro Ser Tyr Glu Glu Thr Ile Arg Ser Val Leu Ile Thr Val Glu
 100 105 110
 Met Leu His Phe Val Arg Arg Arg Pro Thr Asp Ser Glu Lys Asp Met
 115 120 125
 Trp Asp His Leu Tyr Gly Val Phe Glu Arg Phe Val Val Arg Pro Leu
 130 135 140
 Glu Asp Asp Phe Ala Asn His Gln Asn Leu Ile Lys Leu Phe Ala Gln
 145 150 155 160
 Arg Asp Pro Asp Leu Ala Asn Ser Gln Val Leu Gln Val Phe Ile Lys
 165 170 175
 Asp Lys Ile Met Glu Lys Thr Asn Glu Val Gly Ser Asn Asn Leu Asp
 180 185 190
 Asn Lys Arg Glu Pro Asp Ile Lys Gln Glu Pro Asp Ile Lys Gln Glu
 195 200 205
 Pro Val Ala Ala Gly Asp Glu Met Glu Glu Ile Val Glu Glu Gly Ile
 210 215 220

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Pro Asp Ala Pro Ser Asn Asp Asp Asp Asp Asp Glu Glu Asp Glu Glu
 225 230 235 240
 Asp Gly Asp Leu Phe Asp Ser Val Cys Ala Ile Cys Asp Asn Gly Gly
 245 250 255
 Glu Leu Leu Cys Cys Glu Gly Pro Cys Met Arg Ser Phe His Ala Lys
 260 265 270
 Ile Arg Asp Gly Glu Asp Ser Tyr Cys Ala Thr Leu Gly Tyr Thr Lys
 275 280 285
 Ala Glu Val Lys Ala Leu Lys Asn Phe Val Cys Lys Asn Cys Asp His
 290 295 300
 Lys Gln His Gln Cys Phe Val Cys Gly Glu Leu Glu Pro Ser Asp Gly
 305 310 315 320
 Pro Asn Ala Lys Val Phe Leu Cys Asn Asn Ala Thr Cys Gly His Phe
 325 330 335
 Tyr His Pro Arg Cys Val Ala Gln Leu Leu His Pro Asn Ser Arg Asn
 340 345 350
 Glu Ala Ser Glu Met Glu Lys Lys Ile Met Ala Gly Phe Ser Phe Thr
 355 360 365
 Cys Pro Val His Trp Cys Phe His Cys Lys Gly Leu Glu Asp Arg Thr
 370 375 380
 Gln Glu Pro Leu Gln Phe Ala Val Cys Arg Arg Cys Pro Arg Ser Tyr
 385 390 395 400
 His Arg Lys Cys Leu Pro Arg Glu Ile Ser Phe Glu Asp Ile Asn Thr
 405 410 415
 Gln Gly Ile Ile Thr Arg Ala Trp Glu Leu Ser Lys Arg Ile Leu Ile
 420 425 430
 Tyr Cys Leu Asp His Glu Ile Asp Leu Asp Ile Gly Thr Pro Pro Arg
 435 440 445
 Asp His Ile Lys Phe Pro His Val Glu Lys Ser Ala Tyr Ser Ala Lys
 450 455 460
 Lys Lys Val Lys Glu Leu Ala Glu Lys Lys Arg Arg Ile Cys Asp Asp
 465 470 475 480
 Ser Tyr Val Ser Glu Pro Leu Gln Lys Arg Ala Lys Leu Asn Glu Lys
 485 490 495
 Phe Asn Ala Lys Gly Asp Lys Ser Lys Lys Ala Gly Val Lys Ser Glu
 500 505 510
 Phe Glu Glu Val Leu Glu Ser Glu Lys Lys Lys Thr Arg Ser Leu Lys
 515 520 525
 Lys Arg Thr Gln Pro Glu Glu Pro Leu Val Glu Cys Ala Ala Ala Ala
 530 535 540
 Ala Ala Asn Asn Ala Asn Arg Pro Val Lys Glu Arg Glu Lys Glu Leu
 545 550 555 560
 Gly Thr Ser Ser Leu Asp Met Gly Lys Ile Pro Leu Ser Ser Phe Pro
 565 570 575
 Ile Val Asp Ser Glu Thr Glu Lys Arg Ile Ser Ala Leu Val Glu Lys
 580 585 590
 Glu Val Ser Ser Leu Thr Val Ala Asp Ile Ser Arg Arg Cys Val Ile
 595 600 605
 Pro Ser Thr Tyr Ala Cys Ser Gly Arg Gln Ile Asp Lys Ile Val Val
 610 615 620
 Arg Gly Lys Leu Glu Arg Ser Ile Gln Ala Val Lys Ala Ala Leu Gln
 625 630 635 640

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Lys Leu Glu Asn Gly Gly Ala Val Asp Asp Ala Lys Ala Val Cys Glu
645 650 655

Ser Glu Val Leu Arg Gln Leu Thr Arg Trp His Asn Lys Leu Arg Val
660 665 670

Tyr Leu Ala Pro Phe Ile His Gly Met Arg Tyr Thr Ser Phe Gly Arg
675 680 685

His Phe Thr Lys Lys Glu Lys Leu Ile Glu Ile Ala Glu Lys Leu His
690 695 700

Trp Tyr Val Gln Pro Gly Asp Met Lys Ser Asn Asn Val Asp Pro Glu
705 710 715 720

Thr Arg Pro Arg Arg Val Asn Met Leu Arg Gly Phe Gly Ala Leu Ser
725 730 735

Gln Phe Met Lys Glu Lys Leu Asp Lys Val Gly Lys Arg Cys Asn Phe
740 745 750

Lys Asn Tyr Asp Val Ile Gln Pro Lys Asn Ser Phe Ser Phe Glu Lys
755 760 765

Arg Asp Trp Met Thr Val Arg Gln Lys Glu Leu Pro His Gly Ser Lys
770 775 780

Leu Ile Met Gly Leu Asn Pro Pro Phe Gly Pro Lys Ala Met Leu Ala
785 790 795 800

Asn Lys Phe Ile Asp Lys Ala Leu Thr Phe Lys Pro Lys Leu Ile Ile
805 810 815

Leu Ile Val Pro Lys Glu Ala Glu Arg Leu Asp Arg Lys Gln Gln Pro
820 825 830

Tyr Asp Leu Val Trp Glu Asp Asp Gln Arg Leu Ser Gly Lys Ser Phe
835 840 845

Tyr Leu Pro Gly Ser Leu Asp Val Ser Asp Lys Gln Ile Asp Gln Trp
850 855 860

Asn Lys Ser Pro Pro Pro Leu Tyr Leu Trp Ser Arg Pro Asp Trp Thr
865 870 875 880

Gln Lys His Lys Arg Ile Ala Glu Gln His Gly His Thr Lys Ala Asn
885 890 895

Val Phe Ser His Asn Glu Glu Asp Leu Val Tyr Leu Phe Glu Asp Arg
900 905 910

Ala Thr Gln Asn His Asp Val Asn Asn Lys Asn Tyr Thr Ser Gly Asn
915 920 925

Gly Asn Phe Thr Ala Glu Lys Pro Val Gln Ala Asp Ala Phe Pro Pro
930 935 940

Glu Lys Leu Val Glu Val Ala Tyr Glu Glu Met Lys Val Ala Ser Asn
945 950 955 960

Arg Ser Ser Met Tyr Gln Ser Asp Gln Ile Ser Val His Asp Glu Arg
965 970 975

Asp Ala His Ser Asp Leu Pro Met Ser Arg His Asn Ser Met Lys Ala
980 985 990

Lys Glu Val Ser Asn Ser Ser Arg Asp Arg Arg Lys Ser Asp Lys Thr
995 1000 1005

Gly His Glu Ala Asp Ser Asp Met Ser Ile Leu Pro Ser Asp Ser
1010 1015 1020

Arg Asn Phe Leu His Lys Ser Gly Asn Leu Glu Pro Pro Ile Ser
1025 1030 1035

Ser Arg Ser Gly Tyr Thr Leu Glu Arg Leu Arg Tyr His Asp Asn
1040 1045 1050

His Phe Asp His Leu Val Gly Glu His Ser Ser Ser Ser Leu Gln

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1055	1060	1065
Met Pro Ile Phe Glu Asp Ser Tyr Phe Arg Ser Val Asn Glu Tyr 1070	1075	1080
Gly Val Ala Ser Val Glu Asn Asn Ile Ala Leu Ser Thr Asp Asn 1085	1090	1095
Val Gly Ala Gly Ser Arg Met Tyr Ser Pro Asp Pro Glu Leu Asn 1100	1105	1110
Gly Tyr Ala Val Asp Pro Thr Val Asn Ala Tyr Gly Ser Val Ser 1115	1120	1125
Gly Gly Thr Gly Gly Ser Phe Tyr Arg Arg Gln Asn Leu Glu Asp 1130	1135	1140
Tyr Thr Met Asp Ser Ser Glu Ser Ala Gln Met Asn Pro Val Pro 1145	1150	1155
Gly Arg Asp Val Gln Glu Tyr Ala Arg Thr Tyr Tyr Gly His Asn 1160	1165	1170
Arg Asp Glu Val Pro Gln Thr Ala Ile Asn Thr Pro Ser Met Asp 1175	1180	1185
Ile Arg Thr His Ile Arg Met Tyr Gly Arg His Ile Arg Asp Asp 1190	1195	1200
His Thr Gln Thr Thr Met Asn Pro Pro Ala Asn Asp Ile Arg Ala 1205	1210	1215
Gln Ile Arg Met Tyr Gly Gln His Ala Thr Ser Asp His Gln His 1220	1225	1230
Ala Ser Arg Tyr Ser Ser Gly Ser Pro Asp Ala Arg Phe Glu Gln 1235	1240	1245
Gln Pro Ser Phe Thr Ser Tyr Gly Met Pro Ser Leu Gly Ser Thr 1250	1255	1260
Gly Arg Ser Met Met Asp Arg Tyr Ser Pro Ser Ile Asp Glu Thr 1265	1270	1275
Ser Tyr Arg Thr Gly Gln Arg Gly Pro Tyr Asn Ala Ser Asp Phe 1280	1285	1290
Arg Arg Asp Arg His Pro Asp Asp Met Asn Phe Ala Leu His Asn 1295	1300	1305
Gln Tyr Pro Tyr Pro His Pro Gly Ser Ser Gly Gly Trp His Asp 1310	1315	1320

<210> SEQ ID NO 12

<211> LENGTH: 631

<212> TYPE: PRT

<213> ORGANISM: Sorghum bicolor

<400> SEQUENCE: 12

Met Glu Asn Lys Thr Gln Arg Ala Leu Gln Leu Ala Val Cys Arg Arg 1 5 10 15
Cys Pro Arg Ala Tyr His Trp Glu Cys Leu Pro Arg Glu Leu Ser Leu 20 25 30
Gly Ala Lys Asp Lys Asp Gly Asn Pro Arg Ala Trp Lys Leu Ser Lys 35 40 45
Thr Ile Phe Phe Tyr Cys Leu Asp His Glu Ile Asp Lys Asp Thr Arg 50 55 60
Thr Ala Ser Arg Asn His Ile Lys Phe Pro Ala Thr Pro Glu Cys Thr 65 70 75 80
Lys Thr Lys Glu Leu Gly Asn Arg Lys Gly Arg Met Thr Gly Lys Arg 85 90 95

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Arg Lys Asn Thr Asp Gln Ser Thr Glu Pro Thr Glu Leu Ser Asn Arg
 100 105 110
 Leu Tyr Gly Ala Glu Ser Glu Gln Ala Asp Asn Val Gly Ala Lys Ser
 115 120 125
 Thr Ser Pro Gln Ile Val Val Glu Pro His Cys Ala Ala Lys Val Leu
 130 135 140
 Lys Gly Asp Pro Gln Ile Glu Gln Ser Ile Ile Gly Val Ala Gly Ser
 145 150 155 160
 Gln Asn Gly Ala Glu Thr Met Asn Gly His Glu Lys Gln Phe Gly Ile
 165 170 175
 Ser Cys Val Ala Arg Thr Glu Thr Glu Lys Arg Val Thr Tyr Leu Ala
 180 185 190
 Gln Lys Gly Thr Cys Leu Gly Thr Pro Tyr Asp Gly Pro Ser Thr Lys
 195 200 205
 Asp Met Ser Asp Cys Ser Val Gln Asp Thr Pro Val Asp Lys Asp Phe
 210 215 220
 Glu Leu Asp Asn Val Ala Tyr Arg Ile Met Glu Asp Lys Tyr Ala Asn
 225 230 235 240
 Gly Arg Glu Glu Thr Gln Glu Asp Tyr Thr Arg Lys Glu Thr Ala His
 245 250 255
 Arg Lys Asp Ser Ser Glu Asn Gln Gly Gln Asn Asp Val Leu Glu Leu
 260 265 270
 Asp Asn Leu Trp Val Glu Ile Gln Ala Asp Gly Ser Pro Leu Glu Pro
 275 280 285
 Gly Asn Lys Arg Tyr Lys Glu Glu Asn Ala Tyr Gly Leu Gly Ser Ala
 290 295 300
 Ser Gly His Glu Lys Glu Thr Ser Ser Ser Arg Arg Glu Asn Val Gln
 305 310 315 320
 Ser Asp Arg Gly Met Val Pro Met Asn Asp Ser Lys Thr Ile Asp Tyr
 325 330 335
 Arg Lys Gly Gly Thr Thr Leu Asp Asn Asn Val Tyr Asp His Ser Ser
 340 345 350
 Glu Gly Ser Tyr Pro Cys Gln Gly Glu Cys Ser His Ser Lys Cys Asn
 355 360 365
 Asp Gly Leu Val Ala Ile Asp Gln Asp Thr Ser Ser Asp Arg Leu Lys
 370 375 380
 Lys Arg Ser Gln Pro Val Glu Lys Ala Ser Asp Gly Asn Lys Thr Asp
 385 390 395 400
 Leu Asp Lys Asn Lys Lys His Asn Leu Lys Glu Asp Gly Arg Asp Ala
 405 410 415
 His Tyr Glu Asp Arg Arg Thr Glu Arg Asn Thr Ala Ala Asp Thr Ser
 420 425 430
 Arg Tyr Lys Cys Arg Asp Lys Ile Gln Leu Asp Arg Arg Glu Pro Glu
 435 440 445
 Leu Val Gly Arg Asn Thr Arg Ala Arg Ser Ser Glu His Ser Pro Glu
 450 455 460
 Arg Gln Arg Met Glu Arg Asp Gly Ser Tyr Pro Gly Thr Tyr Asn Arg
 465 470 475 480
 Arg Arg Tyr Glu Ser Leu His Asn Phe Asn Pro Pro Arg Ser Gly Cys
 485 490 495
 Asp Asp Arg Arg Gln Leu Ser Pro Cys Gln Ser Ser Phe Pro Leu Pro
 500 505 510
 Glu Phe Cys Gly Asp His Ser His Leu Tyr Pro Arg Asp Ser Thr Ile

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515					520					525					
Gly	Arg	His	Asn	Pro	His	Arg	Tyr	Leu	Gly	Ile	Pro	Gln	Tyr	Gly	Pro
530					535					540					
Tyr	Met	Ala	Ala	Ser	Ala	Ala	Gly	His	Ser	Ala	Val	Cys	Tyr	Arg	Leu
545				550					555						560
Ala	Gly	Gly	Tyr	Gly	Glu	Gly	Ser	Arg	Ala	Ser	Arg	Pro	Val	Thr	Asp
				565					570					575	
Trp	Tyr	Ala	Pro	His	Leu	Asp	Arg	Thr	Asn	Cys	Gln	Pro	Arg	Ser	Gln
			580					585						590	
Ile	Asp	Leu	Gln	Leu	Gln	Ala	Ser	Arg	Pro	Val	Thr	Asp	Lys	Tyr	Ala
		595				600						605			
Pro	Gln	Leu	Glu	Leu	Thr	Asn	Tyr	Pro	Pro	Arg	Ser	Gln	Ser	Asp	Leu
	610					615					620				
Gln	Tyr	Cys	Thr	Thr	Thr	Ile									
625					630										

<210> SEQ ID NO 13

<211> LENGTH: 1420

<212> TYPE: PRT

<213> ORGANISM: Brachypodium distachyon

<400> SEQUENCE: 13

Met	Met	Met	Ser	Asp	Asp	Asp	Asp	Asp	Ser	Glu	Pro	Gln	Val	Val	Val
1			5					10					15		
Val	Lys	Asp	Tyr	Tyr	Phe	Val	Asp	Ala	Asp	Lys	Asn	Ala	Leu	Cys	Phe
		20						25					30		
Ser	Val	Leu	Pro	Ile	Trp	Phe	Lys	Glu	Asp	Ala	Val	Ala	Val	Pro	Glu
		35					40					45			
Cys	Lys	Thr	Gly	Val	Phe	Leu	Arg	Gly	Thr	Val	Asp	Pro	Gly	Ile	Pro
	50					55					60				
Val	Tyr	Lys	Gln	Val	Val	Ala	Trp	Lys	Leu	Gly	Leu	Asp	Ala	Arg	Gln
65				70					75					80	
Pro	Asp	Leu	Ala	Val	Leu	Ser	Lys	Glu	Gly	Gly	Trp	Ile	Asn	Leu	Ser
			85						90					95	
Lys	Pro	Lys	Asn	Ser	Tyr	Glu	Glu	Ser	Phe	Arg	Thr	Ile	Phe	Ile	Thr
			100					105					110		
Val	Gln	Met	Leu	His	Phe	Leu	Arg	Arg	Lys	Pro	Glu	Glu	Pro	Glu	Lys
		115					120					125			
Asp	Leu	Trp	Ile	His	Leu	Arg	Lys	Val	Phe	Asp	Lys	Phe	Asp	Val	Arg
	130					135					140				
Pro	Ser	Lys	Asp	Asp	Phe	Arg	Asn	His	His	Thr	Leu	Met	Lys	Gln	Phe
145				150					155					160	
Ala	Glu	Lys	Asp	Leu	Arg	Leu	Ala	Asn	Ser	Glu	Ile	Leu	Lys	Val	Phe
			165						170					175	
Ile	Gly	Glu	Arg	Phe	Arg	Lys	Gln	Ile	Ser	Glu	Val	Asp	Ser	Gly	Asn
		180					185						190		
Phe	Glu	Val	Lys	Glu	Ser	Phe	Ile	Ala	Ala	Asp	Glu	Asp	Val	Glu	Asp
		195					200					205			
Ile	Val	Ala	Asp	Asp	Asn	Val	Glu	Ser	Asp	Glu	Asp	Gly	Asp	Asp	Asp
	210					215					220				
Leu	Phe	Asp	Ser	Thr	Cys	Ala	Ile	Cys	Asp	Asn	Gly	Gly	Asp	Leu	Leu
225				230					235					240	
Gly	Cys	Asp	Gly	Pro	Cys	Met	Arg	Ser	Phe	His	Ala	Lys	Ile	Gly	Thr
				245					250					255	

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675				680				685							
Val	Asn	Thr	Ala	Lys	Ala	Thr	Cys	Glu	Pro	Gln	Val	Leu	Lys	Gln	Leu
690						695					700				
Ala	Arg	Trp	His	Met	Lys	Leu	Lys	Val	Tyr	Ile	Ser	Pro	Phe	Ile	Tyr
705				710					715						720
Gly	Ser	Arg	Tyr	Ser	Ser	Phe	Gly	Arg	His	Phe	Thr	Lys	Val	Glu	Lys
				725					730					735	
Leu	Val	Glu	Ile	Val	Asp	Lys	Leu	His	Trp	Tyr	Val	Glu	Pro	Gly	Asp
			740					745				750			
Met	Ile	Val	Asp	Phe	Cys	Cys	Gly	Ala	Asn	Asp	Phe	Ser	Arg	Leu	Met
			755				760					765			
Lys	Glu	Lys	Leu	Asp	Leu	Val	Gln	Lys	Lys	Cys	His	Phe	Lys	Asn	Tyr
	770					775					780				
Asp	Leu	Ile	Gln	Pro	Gln	Asn	Thr	Phe	Cys	Phe	Glu	Arg	Arg	Asp	Trp
785					790					795					800
Met	Thr	Val	Gln	Arg	Asn	Glu	Leu	Pro	Arg	Gly	Ser	Arg	Leu	Val	Met
				805					810					815	
Gly	Leu	Asn	Pro	Pro	Phe	Gly	Val	Lys	Ala	Ala	Leu	Ala	Asn	Lys	Phe
			820					825					830		
Ile	Asp	Lys	Ala	Leu	Ser	Phe	Asn	Pro	Lys	Leu	Ile	Ile	Leu	Ile	Val
	835						840					845			
Pro	Lys	Glu	Thr	Lys	Arg	Leu	Asp	Gln	Lys	Lys	Thr	Pro	Tyr	Asp	Leu
	850				855						860				
Val	Trp	Glu	Asp	Gly	Asp	Cys	Leu	Ala	Gly	Lys	Ser	Phe	Tyr	Leu	Pro
865					870					875					880
Gly	Ser	Val	Asp	Val	Asn	Glu	Lys	Ile	Val	Gln	Gly	Trp	Asn	Ala	Ser
				885					890					895	
Ala	Pro	Pro	Leu	Tyr	Leu	Trp	Ser	His	Pro	Asp	Trp	Thr	Lys	Lys	His
			900					905					910		
Lys	Lys	Val	Ala	Glu	Glu	His	Asn	His	Thr	Ser	Leu	Ala	Lys	Ile	Ala
		915					920						925		
Cys	Arg	Ile	Glu	Glu	Gly	Asn	Leu	Ser	Asp	Asp	Val	Pro	Met	Lys	Lys
	930					935					940				
Glu	Ala	Glu	Ser	Ser	Asp	Val	His	Asn	Ser	Arg	Pro	Arg	Lys	Glu	Asp
945					950					955					960
Glu	Asn	Thr	Gly	Arg	Thr	Ser	Cys	His	Leu	Glu	Glu	Ala	Ser	Leu	Ser
				965					970					975	
Asn	Val	Val	Pro	Val	Gln	Arg	Gln	Ala	Glu	Pro	Lys	Ser	Lys	Gln	Asn
			980					985					990		
Ala	Arg	Ser	Gly	Lys	Ala	Lys	Trp	Thr	Lys	Glu	Arg	Thr	Ser	Cys	Asp
		995					1000						1005		
Val	Arg	Asp	Val	Ile	Pro	Ser	Asp	Glu	Thr	Leu	Ala	Lys	Lys	Gln	
	1010					1015					1020				
Asp	Arg	Ser	Gly	Glu	Asp	Gln	Ala	Lys	Glu	Pro	Asn	His	Leu	Val	
	1025					1030					1035				
Gln	Lys	Gln	Ser	Arg	Ser	Gly	Glu	Asp	Lys	Ala	Lys	Glu	Pro	Asn	
	1040					1045					1050				
Arg	Leu	Val	Lys	Lys	Gln	Ala	Arg	Phe	Gly	Glu	Glu	Lys	Asp	Lys	
	1055					1060						1065			
Glu	Arg	Asn	Arg	Leu	Val	Lys	Lys	Gln	Ala	Arg	Ser	Gly	Glu	Asp	
	1070					1075					1080				
Lys	Tyr	Ser	Asn	Leu	Ala	Gly	Gly	Leu	Ser	Ala	Lys	Asn	Gln	Ala	
	1085					1090					1095				

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Glu Ala Ala Leu Gln Gln Met Cys Arg Ser Gly Lys His Asn Ser
 1100 1105 1110
 Arg Asp Gly Ser Lys Ser Ser Asp Asp Arg Ser Arg Lys Arg Thr
 1115 1120 1125
 Pro Asp Glu Val Asp Ser Leu Pro Pro Glu Lys Gln Val Glu Val
 1130 1135 1140
 Ala Phe Glu Glu Arg Arg Ala Pro Ile Lys Met Ser Ile Gln Arg
 1145 1150 1155
 Glu Gln Arg Asp Ala Phe Cys Glu Asn Leu Arg Asn Asp His Ile
 1160 1165 1170
 Lys Glu Pro Ser Arg Gly Ser Ser Asp Met Asn Met Ser Ser Pro
 1175 1180 1185
 Asp Thr Ser Asn Ala Pro Asn Arg Ser Thr Ser Tyr Ser Pro Tyr
 1190 1195 1200
 Met Pro Thr Glu Gln Pro Ser Glu Phe Arg Pro Thr Ala Tyr Leu
 1205 1210 1215
 Asp Gly Asn Met Ser Tyr Pro Val Lys Glu Pro His Val Ser Ala
 1220 1225 1230
 Phe Ser Ser Ala Thr Tyr Gln Gly Ser Tyr Leu Ala Arg Ser Asp
 1235 1240 1245
 Arg His Asn Asp Ala Leu Gly Val Lys Asn Asp Pro Met Leu Tyr
 1250 1255 1260
 Thr His Ala Val Asp Gly Ser Lys Tyr Ser Pro Ser Phe Glu Glu
 1265 1270 1275
 Leu Thr Met Arg Tyr Ala Ala Asn Pro Ala Gly Asp Gly Tyr Ser
 1280 1285 1290
 Met Gln Ala Gln Gly Asp Asp Tyr Leu Pro Met Ser Arg His Ser
 1295 1300 1305
 Leu Gly Ser Ser Gly Ala Arg Tyr Asp Gln Pro Ser Leu Arg Ser
 1310 1315 1320
 Tyr Tyr Gly Leu Ser Gly Thr Thr Ala Pro Gln Ser Ser Ile Thr
 1325 1330 1335
 Asp Lys Tyr Gly Pro Gly Leu Phe Gly Pro Ser Gly Ser Gly Ala
 1340 1345 1350
 Ser Val Thr Asp Lys Tyr Ala Pro Gly Phe Leu Gly Pro Ser Ala
 1355 1360 1365
 Pro Gly Ser Ser Val Ile Asp Asn Tyr Ala Ala Pro Leu Asn Gly
 1370 1375 1380
 Thr Asn Tyr Ala Thr Gln Ser Val Ile Asp Met Pro Gly Tyr Gly
 1385 1390 1395
 Arg Glu Met Pro Pro Gln Tyr Pro Tyr Arg Gly Pro Gly Ser Ser
 1400 1405 1410
 Gly Gly Gly Leu Pro Tyr Thr
 1415 1420

<210> SEQ ID NO 14

<211> LENGTH: 1341

<212> TYPE: DNA

<213> ORGANISM: Zea mays

<400> SEQUENCE: 14

atggccgcca ccaggagagc cttcttcac agcgccgctg acggcattgc gcgcccggg 60

ccgggggagg ccgagcggct gccggcgccg ccgcaggtcg ggcgaccggt ggaaggcgc 120

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agcagcatgg tcttgggctt ccccgtagcg cgccccacga tgccggaccg ccggccggcc 180
gccgtcaccg agcagttctt tccgccact actacggccg cgcagcaggc gacgatggag 240
gagcaatgcc acgtgcccgc cggtagcgcg gcggagcagt gggtcggctc gtcggcgctc 300
aggaagagcc ggcgcggccc gcggtcacgg agctcgcagt accgcggcgt caccttctac 360
cgccgcaccg gccgtggga gtcccacatc tgggactgtg ggaaacaggt gtacttgggt 420
ggattcgaca cggcgcaggc cgctgcgagg gcctacgacc aagcccgcat caagtccgc 480
ggcctgaacg cggacatcaa cttcaccctg gacgactaca aggacgagat gaagaagatg 540
aaggacttga gcaaggagga gttcgtgttg gtgctccggc gccagggcgc cggcttcgctc 600
aggggagcgt ccaggttcgc gggagtcacc cagcacaagt gcggcaagtg ggaggccagg 660
atcgccagc tcatgggcaa gaagtatgtg tacttgggcc tgtatgacac agagacggag 720
gtgcccagg catatgacaa ggctgccatc aagtgtacg gcaaggaggc ggtgaccaac 780
ttcgatgccc agagctacga caaggagctc cagtgcgagc cctgggagcg cgagctggat 840
ctcgagctca gtctgggctg cgcagcagc gatccgtcca cggtcgccgt cgaggcgttc 900
agccccgca cgagcagcag tagccgcaag cagaggacga tgacgctgac gctcggctctg 960
ccggaggagg aggagacggg cgcggctac cctcaccctg ctgcccgat gttcggcgc 1020
ccggctgatg gccacgtcca cgtagcaccg ccgccacacc ggcaatggca gcagcagcag 1080
cagggacagc acgcagctcc agatgcggcg cctgagcgac gagcggcaga gccagcagat 1140
cggcagcggg ggggcggggg cgcgcgctgg cccatcgcca gcgccagcgg cattaactgg 1200
gcctgggccc cgccgtacgc caccgcccgc gccggcaccg acgacgacga cgcagcagc 1260
gcgcccgtg cagcatcacc aggattccca ctgtggcagc tgggtgcggc gtcgtccagg 1320
tccagctggc ccagctgctg a 1341

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<210> SEQ ID NO 15

<211> LENGTH: 446

<212> TYPE: PRT

<213> ORGANISM: Zea mays

<400> SEQUENCE: 15

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Met Ala Ala Thr Arg Arg Ala Phe Phe His Ser Ala Val Asp Gly Ile
1           5           10          15
Ala Arg Ala Gly Pro Gly Glu Ala Glu Arg Leu Pro Ala Pro Pro Gln
20          25          30
Val Gly Arg Pro Val Glu Gly Ala Ser Ser Met Val Leu Gly Phe Pro
35          40          45
Val Pro Arg Pro Thr Met Pro Asp Arg Arg Pro Ala Ala Val Thr Gln
50          55          60
Gln Phe Phe Pro Pro Thr Thr Thr Ala Ala Gln Gln Ala Thr Met Glu
65          70          75          80
Glu Gln Cys His Val Pro Ala Gly Ser Ala Ala Glu Gln Trp Val Arg
85          90          95
Ser Ser Ala Ser Arg Lys Ser Arg Arg Gly Pro Arg Ser Arg Ser Ser
100         105         110
Gln Tyr Arg Gly Val Thr Phe Tyr Arg Arg Thr Gly Arg Trp Glu Ser
115         120         125
His Ile Trp Asp Cys Gly Lys Gln Val Tyr Leu Gly Gly Phe Asp Thr
130         135         140
Ala Gln Ala Ala Ala Arg Ala Tyr Asp Gln Ala Ala Ile Lys Phe Arg
145         150         155         160

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Gly Leu Asn Ala Asp Ile Asn Phe Thr Leu Asp Asp Tyr Lys Asp Glu
 165 170 175

Met Lys Lys Met Lys Asp Leu Ser Lys Glu Glu Phe Val Leu Val Leu
 180 185 190

Arg Arg Gln Gly Ala Gly Phe Val Arg Gly Ser Ser Arg Phe Arg Gly
 195 200 205

Val Thr Gln His Lys Cys Gly Lys Trp Glu Ala Arg Ile Gly Gln Leu
 210 215 220

Met Gly Lys Lys Tyr Val Tyr Leu Gly Leu Tyr Asp Thr Glu Thr Glu
 225 230 235 240

Ala Ala Gln Ala Tyr Asp Lys Ala Ala Ile Lys Cys Tyr Gly Lys Glu
 245 250 255

Ala Val Thr Asn Phe Asp Ala Gln Ser Tyr Asp Lys Glu Leu Gln Ser
 260 265 270

Gln Pro Trp Asp Gly Glu Leu Asp Leu Glu Leu Ser Leu Gly Cys Ala
 275 280 285

Ser Ser Asp Pro Ser Thr Val Ala Val Glu Ala Phe Ser Pro Ala Thr
 290 295 300

Ser Ser Ser Ser Arg Lys Gln Arg Thr Met Thr Leu Thr Leu Gly Leu
 305 310 315 320

Pro Glu Glu Glu Glu Thr Gly Ala Gly Tyr Pro His Pro Ala Ala Gly
 325 330 335

Met Phe Gly Arg Pro Ala Asp Gly His Val His Val Ala Pro Pro Pro
 340 345 350

His Arg Gln Trp Gln Gln Gln Gln Gly Gln His Ala Ala Pro Asp
 355 360 365

Ala Ala Pro Glu Arg Arg Ala Ala Glu Pro Ala Asp Arg Gln Arg Trp
 370 375 380

Gly Arg Gly Ala Arg Trp Pro Ile Ala Ser Ala Ser Gly Ile Asn Trp
 385 390 395 400

Ala Trp Ala Pro Pro Tyr Ala Thr Ala Arg Ala Gly Thr Asp Asp Asp
 405 410 415

Asp Ala Ser Ser Ala Ala Ala Ala Ser Ser Gly Phe Pro Leu Trp
 420 425 430

Gln Leu Gly Ala Ala Ser Ser Arg Ser Ser Trp Pro Ser Cys
 435 440 445

<210> SEQ ID NO 16
 <211> LENGTH: 588
 <212> TYPE: DNA
 <213> ORGANISM: Zea mays

<400> SEQUENCE: 16

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 cagatctatg gcgccgtgc gtactatccc ttctacggag cccaagctct gcacgggagg 180
 gtgctcctgc cgcggcgat cgcggccgac gagccggtct acgtgaacgc caagcagttc 240
 aacggcatcc tccggggcg cctggcgcgc gccaaagcgc cggccgccac ggaccgcccg 300
 gtctccggga gccgcaagcc gtacctccac gagtcacgga acctgcacgc gctgcccggg 360
 gcgccccgca cggcggcggt cttcctcaac acccgagacc gcgacggcga ccccgaggcc 420
 ggcagcgcgg ggaaggcggc ggcggcggcg gcgaggatgc aggaggaggga ccggcaggcg 480

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 gacgccgtgt tcctctcgtc gttggcgagc atggcgggcg gcgaggccac ccggtggccc 540

agcgcgccgt cgcgggggcg gggctgctgc gacctgctca aggcgtga 588

<210> SEQ ID NO 17

<211> LENGTH: 195

<212> TYPE: PRT

<213> ORGANISM: Zea mays

<400> SEQUENCE: 17

Met Ala Ala His Pro Glu Leu Leu Leu Leu Asp Arg Tyr Gln Tyr His
1 5 10 15His His Gly Arg Phe Asp Leu Thr Val Gly Gln Ser Met Val Leu Asn
20 25 30Asn Asp Ser Ala Ile Ala Ser His Gln Ile Tyr Gly Ala Ala Ala Tyr
35 40 45Tyr Pro Phe Tyr Gly Ala Gln Ala Leu His Gly Arg Val Leu Leu Pro
50 55 60Pro Ala Ile Ala Ala Asp Glu Pro Val Tyr Val Asn Ala Lys Gln Phe
65 70 75 80Asn Gly Ile Leu Arg Arg Arg Leu Ala Arg Ala Lys Arg Ala Ala Ala
85 90 95Thr Asp Arg Arg Val Ser Gly Ser Arg Lys Pro Tyr Leu His Glu Ser
100 105 110Arg His Leu His Ala Leu Arg Arg Ala Arg Gly Thr Gly Gly Arg Phe
115 120 125Leu Asn Thr Arg Ser Arg Asp Gly Asp Pro Glu Ala Gly Ser Ala Gly
130 135 140Lys Ala Ala Ala Ala Ala Ala Arg Met Gln Glu Glu Asp Arg Gln Ala
145 150 155 160Asp Ala Val Phe Leu Ser Ser Leu Ala Ser Met Ala Gly Gly Glu Ala
165 170 175Thr Arg Trp Pro Ser Ala Pro Ser Arg Gly Arg Gly Cys Cys Asp Leu
180 185 190Leu Lys Ala
195

<210> SEQ ID NO 18

<211> LENGTH: 1711

<212> TYPE: DNA

<213> ORGANISM: Zea mays

<400> SEQUENCE: 18

accacccaaa taagcataaa tagtagtggt tgattgtgta attccagaga tataaacgaa 60

tatctctaga gatctgctc atcaacagct gcagtatttg ctageccacat atatatacac 120

agtccgacac gtagttataa cggaagagag aagcaaagag agaggcagag tgactgcaac 180

catcagtagt tctatgattt tattttttac cgttttggtg ctgtttcatg gtgtttatnt 240

gattgtaggg tggaggagag gtgaaagctg acagaagaga gtgagcacac atggtgcctt 300

tcttgcatga tgtatgatcg agagagttca tgctcgaagc tatgcgtgct cacttctctc 360

tctgtcagcc attagaactc ctctatctct caatctcgat ctccctcttt ctttggtgat 420

ctctcccatg gtgatattta tttgcttctc acgtgttggt ttctctttct tcagcacaca 480

cacaacctgt tcatgttacc ttagggttaa agtttttgca ctttcgctga agatggaaag 540

acaaacagta gatgagtttt ttgaaggttt gacagaagag agtgagcaca cacgggtggtt 600

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tcttaccatg agtgtcatgc taggagctgt gcgtgctcac cctctatctg tcagtcactc 660
atcaagccca tctgtcttat tagcttgttt ccgctgctaa taaatattct agactcaagt 720
ttatttgaca cagagatega tcgctatcct gttcgatgca tatatatata tatatataga 780
agtattattht acatgtgctg atctcgtgta tatcttcttc ctgattagct gatgatctta 840
ttctcatgtg tagtttattt gtcttcgcat ctaacttttt cgcagggggc aagccatgat 900
tgogaagaaa ggaatgtaaa agatggctca gaactccatc acaaagtcta catacggctg 960
aaataagatc tccatcagat cggagagctt tatttattga ttgttttttt agagtcttag 1020
agcaaagccc atgtcttagc attaccaaga caaagaata atccatatat ataccctttg 1080
gatggttcaac atccttcaact ccagggttaa gtttaaccaa cacaaataaa aatatataat 1140
taaacgaccg gcacatagag acaaaaaactc aaatctccaa gaacatctcc actgtcagca 1200
gctccaagg taaaccataa attttcaatg gaacgatcat caaccaccag ggaggaggat 1260
atatagcctc tgctcttcaa atcttcaggt cttgtggcca gattcttgag tttctccagc 1320
ttcactttga agtccagttt ctatgcctga aggcagagca gctcagcct tgctttagc 1380
atgactgcca tgttcaagag taaagacctc gtgtctctaa ggaacacatc gaaaacacag 1440
ttgtttctg tcaagtttca aagcctccac agggctgcaa gaatcatttg tgtagtagtg 1500
gtggttgacg tactctctct caaagaaga acatcatggt tcttaggttg ttttttagtg 1560
cgcttcatt tcattctctg aatatgccc taagctttag ttgaacatac tgatagttaa 1620
atgtgctcgt gtggtcacc gttattgcat gtgcagaatt tatggcactg gtctgattta 1680
tatgaaaata ttttatattg ttgtggcttt a 1711

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<210> SEQ ID NO 19

<211> LENGTH: 719

<212> TYPE: PRT

<213> ORGANISM: Sorghum bicolor

<400> SEQUENCE: 19

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Met Ser Asp Asp Asp Gly Val Asp Pro Glu Ile Glu Asp Val Asn
 1           5           10           15
Gly Tyr Tyr Phe Glu Asp Gly Glu Gly Glu Pro Val Cys Phe Ser Ile
          20           25           30
Leu Pro Phe Gln Phe Gly Glu Asn Asp Asn Glu Ala Asp Phe Ser Arg
          35           40           45
Lys Asn Val Phe Leu His Gly Phe Val Asp Gln Ser Pro His Val Tyr
          50           55           60
Lys Glu Val Val Ala Trp Lys Ile Leu Gln Arg Leu Ile Glu Asn Gly
          65           70           75           80
Phe Glu Arg Thr Lys Lys Val Cys Met Glu Asn Lys Thr Gln Arg Ala
          85           90           95
Leu Gln Leu Ala Val Cys Arg Arg Cys Pro Arg Ala Tyr His Trp Glu
          100          105          110
Cys Leu Pro Arg Glu Leu Ser Leu Gly Ala Lys Asp Lys Asp Gly Asn
          115          120          125
Pro Arg Ala Trp Lys Leu Ser Lys Thr Ile Phe Phe Tyr Cys Leu Asp
          130          135          140
His Glu Ile Asp Lys Asp Thr Arg Thr Ala Ser Arg Asn His Ile Lys
          145          150          155          160
Phe Pro Ala Thr Pro Glu Cys Thr Lys Thr Lys Glu Leu Gly Asn Arg
          165          170          175

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Lys Gly Arg Met Thr Gly Lys Arg Arg Lys Asn Thr Asp Gln Ser Thr
 180 185 190
 Glu Pro Thr Glu Leu Ser Asn Arg Leu Tyr Gly Ala Glu Ser Glu Gln
 195 200 205
 Ala Asp Asn Val Gly Ala Lys Ser Thr Ser Pro Gln Ile Val Val Glu
 210 215 220
 Pro His Cys Ala Ala Lys Val Leu Lys Gly Asp Pro Gln Ile Glu Gln
 225 230 235 240
 Ser Ile Ile Gly Val Ala Gly Ser Gln Asn Gly Ala Glu Thr Met Asn
 245 250 255
 Gly His Glu Lys Gln Phe Gly Ile Ser Cys Val Ala Arg Thr Glu Thr
 260 265 270
 Glu Lys Arg Val Thr Tyr Leu Ala Gln Lys Gly Thr Cys Leu Gly Thr
 275 280 285
 Pro Tyr Asp Gly Pro Ser Thr Lys Asp Met Ser Asp Cys Ser Val Gln
 290 295 300
 Asp Thr Pro Val Asp Lys Asp Phe Glu Leu Asp Asn Val Ala Tyr Arg
 305 310 315 320
 Ile Met Glu Asp Lys Tyr Ala Asn Gly Arg Glu Glu Thr Gln Glu Asp
 325 330 335
 Tyr Thr Arg Lys Glu Thr Ala His Arg Lys Asp Ser Ser Glu Asn Gln
 340 345 350
 Gly Gln Asn Asp Val Leu Glu Leu Asp Asn Leu Trp Val Glu Ile Gln
 355 360 365
 Ala Asp Gly Ser Pro Leu Glu Pro Gly Asn Lys Arg Tyr Lys Glu Glu
 370 375 380
 Asn Ala Tyr Gly Leu Gly Ser Ala Ser Gly His Glu Lys Glu Thr Ser
 385 390 395 400
 Ser Ser Arg Arg Glu Asn Val Gln Ser Asp Arg Gly Met Val Pro Met
 405 410 415
 Asn Asp Ser Lys Thr Ile Asp Tyr Arg Lys Gly Gly Thr Thr Leu Asp
 420 425 430
 Asn Asn Val Tyr Asp His Ser Ser Glu Gly Ser Tyr Pro Cys Gln Gly
 435 440 445
 Glu Cys Ser His Ser Lys Cys Asn Asp Gly Leu Val Ala Ile Asp Gln
 450 455 460
 Asp Thr Ser Ser Asp Arg Leu Lys Lys Arg Ser Gln Pro Val Glu Lys
 465 470 475 480
 Ala Ser Asp Gly Asn Lys Thr Asp Leu Asp Lys Asn Lys Lys His Asn
 485 490 495
 Leu Lys Glu Asp Gly Arg Asp Ala His Tyr Glu Asp Arg Arg Thr Glu
 500 505 510
 Arg Asn Thr Ala Ala Asp Thr Ser Arg Tyr Lys Cys Arg Asp Lys Ile
 515 520 525
 Gln Leu Asp Arg Arg Glu Pro Glu Leu Val Gly Arg Asn Thr Arg Ala
 530 535 540
 Arg Ser Ser Glu His Ser Pro Glu Arg Gln Arg Met Glu Arg Asp Gly
 545 550 555 560
 Ser Tyr Pro Gly Thr Tyr Asn Arg Arg Arg Tyr Glu Ser Leu His Asn
 565 570 575
 Phe Asn Pro Pro Arg Ser Gly Cys Asp Asp Arg Arg Gln Leu Ser Pro
 580 585 590
 Cys Gln Ser Ser Phe Pro Leu Pro Glu Phe Cys Gly Asp His Ser His

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595	600	605
Leu Tyr Pro Arg Asp Ser Thr Ile Gly Arg His Asn Pro His Arg Tyr		
610	615	620
Leu Gly Ile Pro Gln Tyr Gly Pro Tyr Met Ala Ala Ser Ala Ala Gly		
625	630	635
His Ser Ala Val Cys Tyr Arg Leu Ala Gly Gly Tyr Gly Glu Gly Ser		
645	650	655
Arg Ala Ser Arg Pro Val Thr Asp Trp Tyr Ala Pro His Leu Asp Arg		
660	665	670
Thr Asn Cys Gln Pro Arg Ser Gln Ile Asp Leu Gln Leu Gln Ala Ser		
675	680	685
Arg Pro Val Thr Asp Lys Tyr Ala Pro Gln Leu Glu Leu Thr Asn Tyr		
690	695	700
Pro Pro Arg Ser Gln Ser Asp Leu Gln Tyr Cys Thr Thr Thr Ile		
705	710	715

What is claimed is:

1. A polynucleotide molecule comprising a sequence selected from the group consisting of:
 - (a) a sequence encoding a polypeptide at least 95% identical to SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, or SEQ ID NO:9; wherein the polypeptide regulates juvenile to adult phase change in grass plant leaves;
 - (b) a sequence comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, or SEQ ID NO:8;
 - (c) a sequence comprising at least 95% sequence identity over its full length to the full length of SEQ ID NO:2, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves; and
 - (d) a sequence complementary to (a), (b), or (c), further wherein the polynucleotide molecule is operably linked to a heterologous promoter functional in plants.
2. The polynucleotide molecule of claim 1, comprising the coding sequence of SEQ ID NO:2.
3. A recombinant vector comprising the polynucleotide molecule of claim 1.
4. The recombinant vector of claim 3, further comprising at least one additional sequence chosen from the group consisting of: a regulatory sequence, a selectable marker, a leader sequence and a terminator.
5. The recombinant vector of claim 4, wherein the additional sequence is a heterologous sequence.
6. The recombinant vector of claim 3, wherein the promoter is a tissue-specific promoter.
7. The recombinant vector of claim 3, wherein the promoter directs expression in leaf tissue.
8. An expression cassette comprising the polynucleotide molecule of claim 1.
9. A recombinant vector comprising a polynucleotide encoding a siRNA, wherein the polynucleotide comprises a first sequence comprising all or a part of the sequence of claim 1, and a second sequence comprising the reverse complement of the first sequence, wherein the expression of the polynucleotide in a plant down regulates the expression of a polypeptide encoded by the polynucleotide molecule of claim 1 in the plant.
10. A transgenic plant comprising the recombinant vector of claim 3.
11. A transgenic plant comprising the recombinant vector of claim 9.
12. The transgenic plant of claim 10 or 11, wherein the plant is a monocotyledonous plant.
13. The transgenic plant of claim 10 or 11, wherein the plant is a member of the Poaceae.
14. The transgenic plant of claim 10 or 11, wherein the plant is a member of the Panicoideae or the Pooideae.
15. The transgenic plant of claim 10 or 11, wherein the plant is a maize, a rice, a sorghum, or a switchgrass plant.
16. A seed of the transgenic plant of claim 10 or 11, wherein the seed comprises recombinant vector.
17. The plant of claim 10 or 11, wherein the last leaf with epicuticular wax is produced later during plant development relative to that found in an otherwise isogenic plant lacking the recombinant vector.
18. A cell transformed with the recombinant vector of claim 3 or 9.
19. A method of altering the timing of juvenile to adult phase change in a plant, the method comprising expressing the construct of claim 3 in the plant, expressing the construct of claim 9 in the plant, or mutagenizing the polynucleotide molecule of claim 1.
20. The method of claim 19, comprising expressing the construct of claim 3 in the plant.
21. The method of claim 19, comprising expressing the construct of claim 9 in the plant.
22. The method of claim 19, comprising mutagenizing the polynucleotide sequence of SEQ ID NO:2.
23. The method of claim 19, wherein the timing of juvenile to adult phase in the plant is extended relative to a wild type plant.
24. The method of claim 23, wherein the timing of juvenile to adult phase change is calculated by a method comprising counting the last leaf displaying epicuticular wax.
25. The method of claim 19, wherein the plant exhibits an altered trait selected from the group consisting of: an increase of at least one in the numbering of the last leaf which displays epicuticular wax or which does not contain abaxial trichomes; an altered proportion of juvenile, transitional, or adult leaves; enhanced yield of vegetative tissue; enhanced digestibility of vegetative tissue; enhanced resistance to a plant pest; and enhanced resistance to a plant disease, wherein the trait exhibited by the plant is altered relative to a wild type plant.

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26. The method of claim 19, wherein the plant has altered development or morphology when compared to a wild type plant, further wherein the plant displays an altered trait selected from the group consisting of: enhanced disease resistance, enhanced insect resistance, improved forage digestibility, enhanced abiotic stress tolerance, and improved utility for biofuel production, wherein the development, morphology, or trait is altered relative to a wild-type plant.

27. A method of producing plant biomass, the method comprising:

- (a) obtaining a plant according to claim 10 or 11; and
- (b) preparing biomass from said plant or a part thereof.

28. The method of claim 27, further comprising producing biofuel from the biomass.

29. The method of claim 27, comprising producing food or feed from the biomass.

30. The recombinant vector of claim 3, further comprising an additional polynucleotide sequence which, after being transcribed, regulates the timing of the juvenile to adult phase change in a plant.

31. The recombinant vector of claim 30 wherein the additional polynucleotide sequence encodes all or part of a sequence selected from the group consisting of: Glossy15, Cg1, a homolog of either thereof, and/or a sequence complementary thereto.

32. The recombinant vector of claim 9, further comprising an additional polynucleotide sequence which, after being transcribed, regulates the timing of the juvenile to adult phase change in a plant.

33. A transgenic plant comprising the recombinant vector of claim 30.

34. A transgenic plant comprising the recombinant vector of claim 32.

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35. A seed of the transgenic plant of claim 33 or claim 34.

36. The method of claim 19, further comprising modulating the expression of at least a second gene which regulates the timing of the juvenile to adult phase change in a plant.

37. The method of claim 36, wherein the second gene is selected from the group consisting of: Glossy15 and Cg1.

38. The method of claim 36, comprising expressing the recombinant vector of claim 30 or 32 in the plant.

39. The method of claim 36, wherein the timing of juvenile to adult phase in the plant is extended relative to a wild type plant.

40. A method of producing plant biomass, the method comprising:

- (a) obtaining a plant according to claim 33 or 34; and
- (b) preparing biomass from said plant or a part thereof.

41. The method of claim 40, further comprising producing biofuel from the biomass.

42. The method of claim 40, comprising producing food or feed from the biomass.

43. The polynucleotide molecule of claim 1, wherein the sequence encodes a polypeptide that is at least 95% identical to SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, or SEQ ID NO:9; wherein the polypeptide regulates juvenile to adult phase change in grass plant leaves.

44. The polynucleotide molecule of claim 1, wherein the sequence comprises SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, or SEQ ID NO:8.

45. The polynucleotide molecule of claim 1, wherein the sequence comprises at least 95% sequence identity over its full length to the full length of SEQ ID NO:2, wherein the sequence encodes a protein that regulates juvenile to adult phase change in grass plant leaves.

* * * * *